Contract No. and Disclaimer:

This manuscript has been authored by Savannah River Nuclear Solutions, LLC under Contract No. DE-AC09-08SR22470 with the U.S. Department of Energy. The United States Government retains and the publisher, by accepting this article for publication, acknowledges that the United States Government retains a non-exclusive, paid-up, irrevocable, worldwide license to publish or reproduce the published form of this work, or allow others to do so, for United States Government purposes.

1	
2	Rapid Determination of Radiostrontium in Large Soil Samples
3	Sherrod L. Maxwell ¹ , Brian K. Culligan and Patrick J. Shaw
4	Savannah River Nuclear Solutions, LLC, Building 735-B, Aiken, SC 29808, USA
5	
6	¹ Author for correspondence (email: <u>sherrod.maxwell@srs.gov</u>)
7	phone 803-952-7473
8	fax 803-952-7881
9	Keywords: Strontium, ⁹⁰ Sr, ⁸⁹ Sr, large soil, strontium, rapid, Fukushima
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	

27

28 Abstract

A new method for the determination of radiostrontium in large soil samples has 29 been developed at the Savannah River Environmental Laboratory (Aiken, SC, USA) that 30 allows rapid preconcentration and separation of strontium in large soil samples for the 31 measurement of strontium isotopes by gas flow proportional counting. The need for rapid 32 analyses in the event of a Radiological Dispersive Device (RDD) or Improvised Nuclear 33 Device (IND) event is well-known. In addition, the recent accident at Fukushima Nuclear 34 Power Plant in March, 2011 reinforces the need to have rapid analyses for radionuclides 35 in environmental samples in the event of a nuclear accident. The method employs a novel 36 pre-concentration step that utilizes an iron hydroxide precipitation (enhanced with calcium 37 phosphate) followed by a final calcium fluoride precipitation to remove silicates and other 38 matrix components. The pre-concentration steps, in combination with a rapid Sr Resin 39 separation using vacuum box technology, allow very large soil samples to be analyzed for 40 ^{89,90}Sr using gas flow proportional counting with a lower method detection limit. The 41 calcium fluoride precipitation eliminates column flow problems typically associated with 42 large amounts of silicates in large soil samples. 43

44

45 Introduction

There is an increasing need to develop faster analytical methods for emergency response, including emergency environmental samples [1-2]. There are a number of analytical methods reported that use ion exchange/extraction chromatography to determine radiostrontium in soil. Vajda and Kim provide a very good overview of recent radiostrontium separation and analytical measurement techniques. [3]

This review also included more classical methods using fuming nitric 51 52 precipitation as reported by Bojanowski et al [4]. Fuming nitric acid presents handling difficulties and can be very tedious and time-consuming. Wang et al. [5] reported a 53 sequential method to determine actinides and strontium in soil samples. A large anion 54 resin column (Dowex 1x8) was used to collect and separate Pu and Th. The rinse 55 fractions from the anion resin were treated further and processed individually for Am, U 56 57 and Sr. Several sequential precipitations were carried out. An oxalate precipitation was performed at pH 4.2 on the anion resin rinse solution followed by a Sr Resin separation. 58 Strontium was counted using Čerenkov counting, while all actinide fractions were 59 electrodeposited for counting by alpha spectrometry. The chemical recoveries using this 60 method on NRIP (National Institute of Standards and Technology [NIST] Radiochemistry 61 Intercomparison Program) soil for strontium were 63-77%. A large number of sequential 62 steps were required, but the accuracy of the results versus the NIST reference values was 63 64 very good. Cerenkov counting, while it offers benefits regarding selectivity against low energy beta emitters and allows for rapid measurement of ⁸⁹Sr, has a much higher 65 detection limit than gas flow proportional counting. 66

Ageyev et al. [6] reported a sequential method for soil samples. After ashing the 67 samples at 550°C the samples were leached with 8M nitric acid, followed by calcium 68 oxalate precipitation, furnace heating of oxalates, redissolution in hydrochloric acid, iron 69 70 hydroxide precipitation, and a lanthanum precipitation of plutonium, americium and 71 curium. Carbonate, fuming nitric acid, chromate and iron hydroxide precipitations were 72 performed to prepare strontium. Plutonium was separated using Dowex 1 anion resin loaded under reduced atmosphere. Am and Cm were precipitated as LaOH₃, redissolved 73 in dilute hydrochloric acid, separated on Dowex 50 cation resin loaded under reduced 74 pressure. A gradient elution separation of Am and Cm with rare earths was performed 75

 α using α -hydroxy-iso-butyric acid. Chemical yields for Sr were 50-70%. The method is,

⁷⁷ however, relatively complex and would not be considered a rapid method.

Tavčar et al. [7] reported a method to determine actinides in soil. Soil and 78 sediment samples up to 10g were leached using strong nitric acid, filtration, evaporation, 79 and the residue was redissolved in 1M HNO₃. The acid concentration was increased to 80 8M HNO₃ and the samples were loaded onto Dowex 1x8 resin. Sr was collected from the 81 anion exchange eluent solution using calcium oxalate and the Sr was then separated using 82 Sr Resin. Very large elution volumes were used in this method, including 100 ml of water 83 to elute Sr from Sr Resin. The Sr was precipitated as an oxalate, redissolved and 84 measured using liquid scintillation counting. The average chemical yield for Sr was 67%. 85 Based on this survey of the literature, a more rapid method to determine 86 radiostrontium in large soil samples is needed. The method would need simple, effective 87 pre-concentration steps and good chemical yields. 88 Recently, the Savannah River Environmental Laboratory performed analyses on 89 samples received from Japan in April, 2011 as part of a U.S. Department of Energy effort to 90 provide assistance to the government of Japan. Of particular concern was whether it was safe 91 to plant rice in certain areas (prefectures) near Fukushima. Two different analytical 92 93 techniques were employed in the Savannah River Environmental Laboratory to determine ^{89,90}Sr in Japanese soil samples very rapidly. 94 A rapid fusion technique using 1.5 gram soil aliquots to enable a MDA (Minimum 95 Detection Limit) of <1 pCi^{89,90} Sr /g of soil was employed. This sequential technique has 96 been published recently by this laboratory for actinides and radiostrontium in soil and 97 vegetation [8, 9]. It consists of a rapid sodium hydroxide fusion, pre-concentration steps 98 using iron hydroxide and calcium fluoride precipitations, followed by Sr-Resin separation 99

and gas flow proportional counting. To achieve a lower detection limit for analysis of

101	certain soil samples, a 10 gram aliquot of soil was taken, acid-leached and processed with
102	similar preconcetration chemistry. The MDA using this approach was ~0.03 pCi/g (1.1
103	mBq/g)/, which is less than the 0.05-0.10 pCi/g 90 Sr levels found in soil as a result of
104	global fallout. The large amounts of iron in Japanese soil limited the size of soil sample
105	aliquots that could be easily handled to $\sim 10-15$ g due to very large iron hydroxide
106	precipitates. The chemical yields observed for the Japanese soil samples was typically 75-
107	80% and the laboratory control sample (LCS) and matrix spike (MS) results looked very
108	good for this work [10].
109	A newer version of this approach was developed for application to even larger

sample aliquots (50 g) from soil samples taken at the Savannah River Site so that very low detection limits can be achieved. The method uses an acid leach, a rapid sample preconcentration approach and vacuum-assisted column flow rates. The sample analyses required <16 hours to complete, with a significant portion (4 hours or more) of that time being the evaporation of the relatively large leachate solutions.

115

116 **Experimental**

117 Reagents

Sr Resin (4, 4', (5') di-t-butylcyclohexane-18-crown-6), was obtained from
Eichrom Technologies, Inc., (Lyle, Illinois, USA). Nitric, hydrochloric and hydrofluoric
acids were prepared from reagent-grade acids (Fisher Scientific, Inc., Pittsburgh, PA,
USA). All water was obtained from a Milli-Q2TM water purification system. All other
materials were ACS reagent grade and were used as received. Radiochemical isotopes
⁹⁰Sr were obtained from Eckert & Ziegler Analytics, Inc. (Atlanta, GA, USA) and diluted
to the appropriate level.

125

126

128	<i>Column preparation</i> . Sr Resin was obtained as cartridges containing 2 mL of each
129	resin from Eichrom Technologies, Inc Small particle size (50-100 micron) resin was
130	employed, along with a vacuum extraction system (Eichrom Technologies).
131	Sample Preparation. Soil samples obtained from a location near the Savannah
132	River Site (Aiken, SC, USA) were dried at 110 °C and blended prior to taking sample
133	aliquots. The soil was heated in a furnace at 550 °C to remove any organics present.
134	Seven 50 g soil samples were aliquoted into 600 ml glass beakers. Sr carrier (6.09 mg Sr)
135	was added to each sample to allow determination of chemical yield.
136	Known amounts of ⁹⁰ Sr were pipetted into each set of 50 g soil samples to
137	demonstrate performance using this method. ⁹⁰ Sr was added to each set of soil samples
138	to test at the following levels: 59.2 mBq/g, 11.84 mBq/g and 5.92 mBq/g respectively.
139	The uncertainty associated with the known value of 90 Sr standard added was ~3 % at the
140	95% confidence level. Unspiked soil aliquots was analyzed to determine the native
141	amount of ⁹⁰ Sr present in the soil. The stable strontium in the soil was found to be very
142	low relative to the amount of stable strontium added so no correction had to be made for
143	the native stable Sr content.

Figure 1 provides a flow chart of the soil sample preparation method. For 50 g soil 144 145 samples, 50 ml concentrated nitric acid and 25 ml hydrochloric acid volumes were added. The samples were digested on medium heat to prevent splattering and taken to dryness on 146 a hot plate. The solids were rinsed with 50 ml concentrated nitric acid, warmed on a hot 147 plate and the leachate plus additional solids were transferred to a centrifuge tube. The 148 centrifuge tubes were centrifuged at 3500 rpm for 10 minutes. The leachate was 149 150 transferred to a 600 ml beaker and heated on a hot plate to evaporate the leachate to 151 dryness.

The residual solids were rinsed two more times. For each rinse, twenty-five 152 milliliters of concentrated nitric acid were added to each beaker, and the leachate plus 153 154 additional solids were transferred to the centrifuge tube. The centrifuge tubes were centrifuged at 3500 rpm for 10 minutes. The leachate was transferred to a 600 ml beaker, 155 heated on a hot plate to evaporate the leachate to dryness. The leachate solutions were 156 157 evaporated to dryness on a hot plate on low to medium heat. If there were small solids still left in the leachate, the leachate was centrifuged again in a 50 ml tube to remove 158 solids after the leachate volume reached ~40 ml. The residual solids were rinsed with ~10 159 ml concentrated nitric acid, centrifuged, and the rinse was added to the remaining 160 leachate, which was taken to dryness. 161

To each evaporated leachate, 15 to 20 ml 1M hydrochloric acid were added and warmed on a hot plate to easily dissolve the residue. This solution was transferred to 225 mlcentrifuge tube and 1 ml 1.25M calcium nitrate and 2 ml 3.2M ammonium hydrogen phosphate were added to each tube. Twenty-five milliliters of concentrated ammonium hydroxide was added to each tube and the strongly alkaline solution was diluted to 180 ml with water.

168 The tubes were centrifuged at 3500 rpm for 6 minutes and the supernatant was

discarded. The remaining precipitate was dissolved in 40 ml 1.5M HCl This solution was 169 170 diluted to ~170 ml with 0.01M HCl and 25 ml 28M HF were added to each sample. More of the undissolved precipitate resulting from the iron hydroxide/calcium phosphate 171 precipitate will redissolve at this point and a new, much smaller calcium fluoride 172 precipitate will form upon addition of HF. The samples were centrifuged for 10 minutes 173 at 3500 rpm. The CaF₂ precipitate is small and can be dissolved easily in a small volume 174 column load solution. For laboratories that have restrictions regarding the use of 175 hydrofluoric acid, the substitution of sodium fluoride may be possible. 176

The supernatant was removed and the residual solids containing the strontium were dissolved in 7 ml of concentrated nitric acid, 7 ml of 3M HNO₃-0.25M boric acid, and 7 ml of 2 M aluminum nitrate. If any residual solids remained, these were rinsed with 5 ml 3M HNO₃, centrifuged to remove the solids and this rinse was added to the load solution. Additional 3M HNO₃ may be needed to redissolve the precipitate with very high Ca soils.

Column separation. Figure 2 provides a flow chart of the rapid column 183 separation method using two 2 ml Sr cartridges (~1.4 g Sr Resin total). Sr Resin columns 184 were conditioned with 10 ml 8M HNO₃. The sample solution was loaded onto the Sr 185 Resin column at approximately ~ 1 drop per second. After the sample was loaded, a tube 186 rinse of $\sim 5 \text{ mL 8M HNO}_3$ was transferred to the Sr Resin column and allowed to pass 187 188 through the resin at ~ 2 drops per second. The following column rinses were performed at 189 ~2-3 drops per second: 15 ml 8M HNO₃, 10 ml 3M HNO₃- 0.05M oxalic acid, and 10 ml 190 8M HNO₃. Sr was eluted from the resin with 18 ml 0.05M HNO₃ at ~1 drop per second. This solution was transferred to preweighed planchets and evaporated on a hot 191 plate with medium heat to dryness. Two milliliters 0.05M HNO₃ were used to rinse each 192 tube and then was transferred to each planchet, and evaporated to dryness on a hot plate. 193

The dried planchets were allowed to cool and then were weighed to determine gravimetric carrier recovery. The planchets were counted by simultaneous gas flow proportional counting (Tennelec LB 4100) for 90 minutes. The detectors were calibrated using NIST Traceable 90 Sr/ 90 Y sources matching the sample geometry. Detector backgrounds are determined and subtracted from the sample counts. A mass attenuation correction factor was determined experimentally using prepared mounts containing 90 Sr/ 90 Y (>167 Bq) and a nominal amount of Sr carrier.

201

202 <u>Apparatus</u>

Polycarbonate vacuum boxes with 24 positions and a rack to hold 50 ml plastic tubes were used. Two boxes were connected to a single vacuum source by using a Tconnector and individual valves on the tubing to each box.

Planchets were annealed for ~4 hours in a furnace at 550 °C prior to use. This provides chemical resistance to the planchets so that iron oxide does not form during evaporation of the nitric acid, which would cause error in the gravimetric weights.

209

210 Results and Discussion

Table 1 shows the measured values for ⁹⁰Sr in a set of seven 50g soil samples spiked

at the 59.2 mBq/g level. The average 90 Sr result was 57.8 mBq/g ± 1.7 mBq (1SD,

standard deviation) with an average bias of -2.36%. The average stable Sr carrier

recovery was 89.3% (SD =4.7%), indicating very good chemical yield for the method

even for 50g sample aliquots. The measured values were corrected for native content in

the soil of 1.35 mBq 90 Sr /g.

Table 2 shows the measured values for 90 Sr in a set of seven 50 g soil samples spiked at the 11.84 mBq/g level. The average 90 Sr result was 11.5 mBq/g ± 0.7 mBq/g (SD). The average stable Sr carrier recovery was 89.6% (SD =2.7%) with an average bias of – 2.51%.

Table 3 shows the measured values for 90 Sr in a set of seven 50g soil samples spiked at the 5.92 mBq/g level (0.16 pCi/g), which is not much greater than global fallout levels. The average 90 Sr result was 5.95 mBq/g ± 0.22 mBq/g (SD), with an average bias of 0.43%. The average stable Sr carrier recovery was 94.0% (SD =2.6%), indicating very good chemical yield.

The tests indicate that for three different spiked levels that ⁹⁰Sr can be measured very 226 well. The sample pre-concentration steps to remove the soil matrix worked very well. 227 Chemical yields were very good and no column flow issues were observed. The use of 228 iron hydroxide precipitation enhanced with a slight amount of calcium phosphate was 229 very effective, and the calcium fluoride removal of silicates ensured ease of column flow. 230 It has been the experience in this laboratory that blending combinations of different co-231 precipitation steps can be a very effective way to maximize chemical yields from difficult 232 sample matrices. 233

The MDA (Minimum Detectable Activity) for the ⁹⁰Sr using this method with gas flow proportional counting were calculated according to equations prescribed by Currie: [11]

- 237
- 238 MDA = $[3+4.65\sqrt{B}]/(CT*R*V*Eff*0.060)$
- Where B = Total Background counts, = BKG (rate) * BKG Count time
 CT = sample count time (min)
 R = Chemical Recovery
 V = Sample aliquot (g)
 EFF = Detector Efficiency

The MDA for the results can be adjusted as needed, depending on the sample aliquot and count time. For a 50 g sample aliquot, the method MDA for 90 Sr with a 90 minute count time is 0.41 mBq g⁻¹, typically below global fallout levels.

It should be noted that soil aliquots may have to be adjusted if very high levels of calcium are present in the soil. If very high levels of Ca are found, it may be necessary to reduce the soil sample aliquot or not add any additional Ca in the pre-concentration steps. A larger column load solution could also be tolerated with 4ml Sr Resin. Combining purified solutions from smaller replicate aliquots may also be employed to maintain a low MDA even with a more difficult sample matrix.

The test data in Tables 1-3 were obtained with nitric acid/ hydrochloric acid leaching, 256 without aggressively attacking the soil silicate matrix. It is also possible to add 257 hydrofluoric (HF) acid to the acid digestion step if smaller soil aliquots (25g) and large 258 Teflon beakers are used. The HF can be removed via evaporation in the presence of 259 concentrated nitric acid and boric acid. The same sample preconcentration steps may be 260 applied but the chemical yields are not as high. Table 4 shows results using the digestion 261 of 25g samples with hydrofluoric acid. The average 90 Sr result was 12.6 mBq/g ± 1.2 262 mBq/g (SD), with an average bias of 6.1%. The chemical yields for the stable strontium 263 264 carrier were reduced to an average yield of 73%. If acid digestion and leaching without 265 HF provides sufficient ruggedeness (often the case), a larger soil aliquot can be analyzed with a higher chemical yield. If individual ⁸⁹Sr and ⁹⁰Sr results are required, a second 266 count measured after 7-10 days can be used to differentiate ⁸⁹Sr and ⁹⁰Sr. There are also 267

268	Čerenkov counting techniques for more rapid determination of ⁸⁹ Sr and ⁹⁰ Sr, however the
269	MDA levels will be significantly higher using this approach. [12]

Conclusions

272	A new method to determine ⁹⁰ Sr has been developed that allows the rapid
273	separation of radiostrontium in large soil samples with high chemical yields and
274	effective removal of interferences. The simple matrix removal steps and rapid column
275	separation steps resulted in reliable measurements of 90 Sr at very low levels from 50 g
276	soil aliquots. This approach allows large soil samples to be analyzed with a significant
277	reduction in the detection limit.
278	

279 Acknowledgment

280 This work was performed under the auspices of the Department of Energy, DOE
281 Contract No. DE-AC09-96SR18500.

References

- [1] D. Larivière, T. Cumming, S.Kiser, C. Li, and R. Cornett, Automated flow injection system using extraction chromatography for the determination of plutonium in urine by inductively coupled plasma mass spectrometry, J. Anal. At. Spectrom., 23 (2008), 352
- [2] D.L. Stricklin, A. Tjarnhage, and U. Nygren, Application of low energy gamma-spectrometry in rapid actinide analysis for emergency preparedness, J. Radioanal. Nucl.Chem. 251 (2002) (1), 69
- [3] N. Vajda and C.K. Kim, Determination of radiostrontium isotopes: A review of analytical methodology, Applied Radiation and Isotopes 67 (2010), 2306
- [4] R. Bojanowski and D. Knapinska- Skiba, Determination of low-level ⁹⁰Sr in environmental samples: a novel approach to the classical method, J. Radioanal. Nucl.Chem, VoL 138, No. 2 (1990), 207
- [5] J. Wang, I. Chen, and J. Chiu, Sequential isotopic determination of plutonium, thorium, americium, strontium and uranium in environmental and bioassay samples, Applied Radiation and Isotopes, 61 (2004.), 299
- [6] Ageyev, V.A., Odintsov, O.O. and Sajeniouk, A.D., Routine radiochemical method for the determination of ⁹⁰Sr, ²³⁸Pu, ²³⁹⁺²⁴⁰Pu, ²⁴¹Am and ²⁴⁴Cm in environmental samples, J. Radioanal. Nucl.Chem. 264 (2), (2005), 337
- [7] P. Tavčar, R. Jakopič, and L. Benedik, Sequential Determination of ²⁴¹Am, ²³⁷Np,
 Pu Radioisotopes and ⁹⁰Sr in Soil and Sediment Samples, Acta. Chim.Slov. 52 (2005), 60
- [8] S. Maxwell, B. Culligan, and G. Noyes, Rapid method for actinides in emergency soil samples, Radiochmica Acta, Vol. 98, No. 11, (2010), 793

- [9] S. Maxwell, B. Culligan, and G. Noyes, Rapid separation method for actinides and radiostrontium in vegetation samples, J. Radioanal. Nucl.Chem. Vol. 286 (2010), No. 1, 273
- [10] Rapid Radiochemical Methods in Support of Fukushima presentation at Radiobioassay and Radiochemical measurements Conference Eichrom User's group meeting, <u>http://www.eichrom.com/radiochem/meetings/2011/rrmc/pdf/4-</u> Maxwell-Eichromworkshop2011.pdf, November 2, 2011
- [11] L.A., Currie, Limits for qualitative and quantitative determination, Anal. Chem.40 (1968) 586
- J.P. Martin, and K.J. Odell, "The Development of Emergency Radioanalytical Techniques for the Determination of Radiostrontium and Transuranic Radioisotopes in Environmental Materials", Radioactivity and Radiochemisty, 1998, Vol. 9, No 3, 49-60.

Table Captions

- Table 1 ⁹⁰Sr in Soil Results (59.2 mBq/g level) 50 g samples
- Table 2⁹⁰Sr in Soil Results (11.8 mBq/g level) 50 g samples
- Table 3 ⁹⁰Sr in Soil Results (5.92 mBq/g level) 50 g samples
- Table 4 ⁹⁰Sr in Soil Results (11.8 mBq/g) 25g samples with HF Digest

Sample	Sr carrier	⁹⁰ Sr Reference Value	⁹⁰ Sr Reference Value	⁹⁰ Sr Measured Value	Difference
ID	(%)	(pCi g⁻¹)	(mBq g ⁻¹)	(mBq g ⁻¹)	(%)
1	86.4	1.60	59.2	60.6	2.36
2	93.9	1.60	59.2	54.9	-7.26
3	81.0	1.60	59.2	58.3	-1.52
4	92.5	1.60	59.2	57.7	-2.53
5	87.8	1.60	59.2	57.6	-2.70
6	93.9	1.60	59.2	58.3	-1.52
7	89.8	1.60	59.2	57.2	-3.38
Avg	89.3			57.8	-2.36
SD	4.7			1.7	
% RSD	5.3			2.9	
	Measured values corrected for 1.35 mBq ⁹⁰ Sr/g found in unpiked soil				

Table 1⁹⁰Sr in Soil Results (59.2 mBq/g level) - 50 g samples

Sample	Sr carrier	⁹⁰ Sr Reference Value	⁹⁰ Sr Reference Value	⁹⁰ Sr Measured Value	Difference
ID	(%)	(pCi g ⁻¹)	(mBq g ⁻¹)	(mBq g ⁻¹)	(%)
1	87.8	0.32	11.84	11.2	-5.41
2	88.4	0.32	11.84	11.9	0.51
3	87.1	0.32	11.84	12.2	3.04
4	93.9	0.32	11.84	12.7	7.26
5	92.5	0.32	11.84	11.2	-5.41
6	87.1	0.32	11.84	10.9	-7.94
7	90.5	0.32	11.84	10.7	-9.63
Avg	89.6			11.5	-2.51
SD	2.7			0.7	
% RSD	3.1			6.4	
	Measured values corrected for 1.35 mBq ⁹⁰ Sr/g found in unpiked soil				

Table 2⁹⁰Sr in Soil Results (11.8 mBq/g level) - 50 g samples

Sample	Sr carrier	⁹⁰ Sr Reference Value	⁹⁰ Sr Reference Value	⁹⁰ Sr Measured Value	Difference
ID	(%)	(pCi g⁻¹)	(mBq g ⁻¹)	(mBq g ⁻¹)	(%)
1	95.9	0.160	5.92	6.05	2.20
2	98.6	0.160	5.92	6.02	1.69
3	94.6	0.160	5.92	5.82	-1.69
4	91.8	0.160	5.92	6.32	6.76
5	93.2	0.160	5.92	5.96	0.68
6	92.5	0.160	5.92	5.60	-5.41
7	91.2	0.160	5.92	5.85	-1.18
Avg	94.0			5.95	0.43
SD	2.6			0.22	
% RSD	2.8			3.77	
	Measured values corrected for 1.35 mBq ⁹⁰ Sr/g found in unpiked soil				

Table 3 ⁹⁰Sr in Soil Results (5.92 mBq/g level) - 50 g samples

Sample	Sr carrier	⁹⁰ Sr Reference Value	⁹⁰ Sr Reference Value	⁹⁰ Sr Measured Value	Difference
ID	(%)	(pCi g⁻¹)	(mBq g ⁻¹)	(mBq g⁻¹)	(%)
1	78.9	0.32	11.84	13.9	17.40
2	70.1	0.32	11.84	12.4	4.73
3	70.1	0.32	11.84	11.4	-3.72
Avg	73.0			12.6	6.14
SD	5.1			1.3	
% RSD	7.0			10.0	
	Measured value	es corrected for 1.35 mBq ⁶	⁹⁰ Sr/g found in unpiked soil		

Table 4 ⁹⁰Sr in Soil Results (11.8 mBq/g) - 25g samples with HF Digest





Figure 2 Rapid Column Separation for Sr-89/90 in Large Soil Samples