**Vapor Space Characterization of Waste
Tank 241-C-108: Results from Samples
Collected Through the Vapor Sampling
System on 8/5/94**

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Summary

This report describes inorganic and organic analyses results from samples obtained from the headspace of the Hanford waste storage Tank 241-C-108 (referred to as Tank C-108). The results described here were obtained to support safety and toxicological evaluations. A summary of the results for inorganic and organic analytes is listed in Table 1. Detailed descriptions of the results appear in the text.

Quantitative results were obtained for the inorganic compounds ammonia (NH_3), nitrogen dioxide (NO_2), nitric oxide (NO), and water (H_2O). Sampling for hydrogen cyanide (HCN) and sulfur oxides (SO_x) was not requested. Organic compounds were also quantitatively determined. Two organic tentatively identified compounds (TICs) were observed above the detection limit of (ca.) 10 ppbv, but standards for most of these were not available at the time of analysis, and the reported concentrations are semiquantitative estimates. In addition, we looked for the 41 standard TO-14 analytes. Of these, only a few were observed above the 2-ppbv detection limit. The five organic analytes with the highest estimated concentrations are listed in Table 1. The five analytes account for approximately 85% of the total organic components in Tank C-108.

Table 1. Summary Results of Inorganic and Organic Samples Collected from the Headspace of Tank C-108 on 8/5/94

Category	Analyte	Vapor ^(a) Concentration	Units
Inorganic	NH_3	2.7 ± 0.3	ppmv
	NO_2	≤ 0.04	ppmv
	NO	0.24 ± 0.01	ppmv
	H_2O	20 ± 2	mg/L
Organic	Acetaldehyde	0.1 ± 0.02	mg/m^3
	Acetone	0.09 ± 0.03	mg/m^3
	Trichlorofluoromethane	0.06 ± 0.001	mg/m^3
	Ethyl chloride	0.05 ± 0.001	mg/m^3
	Methylene chloride	0.05 ± 0.001	mg/m^3

(a) Vapor concentrations were determined using sample-volume data provided by Westinghouse Hanford Company and are based on averaged data.

Acknowledgments

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Abbreviations

CAS	Chemical Abstracts Service
COC	chain of custody
C _v	concentration by volume
DIW	deionized water
emf	electromotive force
EPA	U.S. Environmental Protection Agency
GC/MS	gas chromatography/mass spectrometry
HP	Hewlett Packard
IC	ion chromatography
IL	impact level
IS	internal standard
MDL	minimum detection limit
NIST	National Institute for Standards and Technology
NPH	normal paraffin hydrocarbon
OSHA	Occupational Safety and Health Administration
PFA	perfluoroalkoxy
PNL	Pacific Northwest Laboratory
ppbv	part per billion by volume
ppmv	part per million by volume
QA	quality assurance
REL	recommended exposure limit
SCIC	suppressed-conductivity ion chromatography
SIE	selective ion electrode
SRM	standard reference material
STP	standard temperature and pressure
TEA	triethanolamine
TIC	tentatively identified compound
VSS	vapor sampling system
WHC	Westinghouse Hanford Company

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1.0 Introduction

This report describes results of the analyses of tank-headspace samples taken from the Hanford waste Tank 241-C-108 (referred to as Tank C-108) using the vapor sampling system (VSS). Pacific Northwest Laboratory (PNL)^(a) contracted with Westinghouse Hanford Company (WHC) to provide sampling devices and to analyze inorganic and organic analytes collected from the tank headspace. The sample job was designated S4052, and samples were collected by WHC on August 5, 1994, using the WHC VSS.

Sampling devices, including six sorbent trains (for inorganic analyses), and five SUMMA^{TM} canisters (for organic analyses) were supplied to WHC on July 26, 1994. Samples were taken (by WHC) from the tank headspace on August 5 and were returned to PNL from the field on August 8. Inorganic (sorbent trap) samples were delivered to PNL on chain-of-custody (COC) 007486 (see Figure 1.1a). Five SUMMA^{TM} canisters were delivered on COC No. 007487 (see Figure 1.1b).

The samples were inspected upon delivery to the 326/23B laboratory and logged into PNL record book 55408 before implementation of PNL Technical Procedure PNL-TVP-07^(b). Custody of the sorbent traps was transferred to PNL personnel performing the inorganic analysis and stored at refrigerated ($\leq 10^{\circ}\text{C}$) temperature until the time of analysis. The canister was stored in the 326/23B laboratory at ambient (25°C) temperature until time of analysis. Access to the 326/23B laboratory is limited to PNL personnel working on the waste-tank safety program. Analyses described in this report were performed at PNL in the 300 area of the Hanford Reservation. Analytical methods that were used are described in the text. In summary, sorbent traps for inorganic analyses containing sample materials were either weighed (for water analysis) or desorbed with the appropriate aqueous solutions (for ammonia, nitrogen oxides, and nitric oxide analyses). The aqueous extracts were analyzed either by selective electrode or by ion chromatography (IC). Organic analyses were performed using cryogenic preconcentration followed by gas chromatography/mass spectrometry (GC/MS).

(a) Pacific Northwest Laboratory is operated for the U. S. Department of Energy by Battelle Memorial Institute under Contract DE-AC06-76RLO 1930.

(b) PNL-TVP-07, Rev. 0, October 1994, *Sample Shipping and Receiving Procedure for PNL Waste Tank Samples*, PNL-Technical Procedure, Tank Vapor Project, Richland, Washington.

Westinghouse Hanford Company	CHAIN OF CUSTODY	WHC 007486

Custody Form Initiator	J. A. Edwards	Telephone	(509) 373-0141
		Pager	85-3009

Company Contact	R. A. Westberg	Telephone	(509) 373-5734

Project Designation/Sampling Locations 200 East Tank Farm	Collection Date
241-C-108 Tank Vapor Sample SAF S4052	07-25-94 @ 08/0/94
Ice Chest No. (VSS Truck)	Preparation Date
	07-25-94
	Field Logbook No. WHC-____

Bill of Lading/Airbill No. N/A	Offsite Property No. N/A

Method of Shipment Government Truck	Sample Job _____

Shipped to PNL

Possible Sample Hazards/Remarks Unknown at time of sampling

Sample Identification

S4052 - A23 .07W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 1) Line # 8
S4052 - A24 .08W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 2) Line #10
S4052 - A25 .09W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 3) Line # 9
S4052 - A26 .10W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 4) Line # 10
S4052 - A27 .11W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 5) Line # 8
S4052 - A28 .12W ✓ SAP Ref _____	NH ₃ /NO _x /H ₂ O (Trap # 6) Line #10
S4052 - A29 .13W ✓ SAP Ref _____	NH ₃ /NO ₂ /H ₂ O (Trip Blank# 1)
S4052 - A30 .14W ✓ SAP Ref _____	NH ₃ /NO ₂ /H ₂ O (Trip Blank# 2)
S4052 - A31 .15W ✓ SAP Ref _____	NH ₃ /NO ₂ /H ₂ O (Trip Blank# 3)

[ ] Field Transfer of Custody			[X] Chain of Possession			(Sign and Print Names)		
Relinquished By	Date	Time	Received By	Date	Time			
J. A. Edwards	07-26-94	0940	T. Utecht	07-26-94	0940			
T. R. Utecht	08-02-94	0930	R. A. Westberg	08-02-94	0945			
T. R. Utecht	08-05-94	0900	T. R. Utecht	08-05-94	0900			
T. R. Utecht	08-09-94	1125	J. A. Edwards	08-09-94	1125			

(Revised 02/28/94)

Final Sample Disposition

Disposal Method:

Disposed by:

Date/Time:

A-60001-407 (12/92) WEF061

Figure 1.1a Chain-of-Custody for Inorganic Samples from Tank C-108

Westinghouse Hanford Company	CHAIN OF CUSTODY	WHC 007487

Custody Form Initiator	J. A. Edwards	Telephone (509) 373 -0141 Pager 85-3009
Company Contact	R. A. Westberg	Telephone (509) 373-5734
Project Designation/Sampling Locations	200 East Tank Farm	Collection Date <u>07-05-94</u> ⁰⁸⁻⁰⁵ ^{10/04}
241-C-108 Tank	Vapor Sample SAF S4052	Preparation Date 07- 2 5 -94
Ice Chest No.	(VSS Truck)	Field Logbook No. WHC- - - -
Bill of Lading/Airbill No.	N/A	Offsite Property No. N/A
Method of Shipment	Government Truck	Sample Job _____
Shipped to	PNL	

Possible Sample Hazards/Remarks Unknown at time of sampling

Sample Identification

S4052 - A01 - 014	SAP Ref _____	Ambient air SUMMA #1, <u>Upwind VSS</u> (PNL)
S4052 - A02 - 015	SAP Ref _____	Ambient air SUMMA #2, <u>Through VSS</u> (PNL)
S4052 - A04 - 016	SAP Ref _____	SUMMA #4 (PNL)
S4052 - A06 - 017	SAP Ref _____	SUMMA #6 (PNL)
S4052 - A08 - 018	SAP Ref _____	SUMMA #8 (PNL)

[X] Field Transfer of Custody			Chain of Possession (Sign and Print Names)		
Relinquished By	Date	Time	Received By	Date	Time
J. A. Edwards <i>J.A. Edwards</i>	07-26-94	0940	T. Utecht <i>T.B. Utecht</i>	07-26-94	0540
<i>T.B. Utecht</i> T. B. Utecht	08-05-94	0845	<i>R. Westberg</i> R. Westberg	08-05-94	0845
<i>T.A. Westberg</i> T. A. Westberg	08-05-94	0900	<i>T.B. Utecht</i> T. B. Utecht	08-05-94	0900
<i>T.B. Utecht</i> T. B. Utecht	08-05-94	1135	<i>J.A. Edwards</i> J. A. Edwards	08-05-94	1135

(Revised 02/28/94)

Final Sample Disposition

Disposal Method:

Disposed by:

Date/Time:

A-6000-407 (12/92) WEF061

Figure 1.1b Chain-of-Custody for Organic Samples from Tank C-108

2.0 Inorganic Task

Solid sorbent traps, prepared in multi-trap sampling trains, were supplied to WHC for sampling the tank headspace using the VSS. Blanks, spiked blanks (when requested), and exposed samples were returned to PNL for analysis. Analyses were performed to provide information on the tank-headspace concentration of the following analytes: ammonia (NH_3), nitrogen dioxide (NO_2), nitric oxide (NO), and water (H_2O). Procedures were similar to those developed previously during sample jobs performed with the VSS connected to the headspace of Tank C-103 (Ligotke et al. 1994). During those sample jobs, control samples provided validation that samples were effectively trapping NH_3 and mass. Analytical accuracy was estimated based on procedures used. Sample preparation and analyses were performed following PNL quality assurance (QA) impact level (IL) III requirements.

2.1 Standard Sampling Methodology

Standard glass tubes containing sorbent materials to trap vapors of selected analytes (NH_3 , NO_x , and H_2O) (SKC Inc., Eighty Four, Pennsylvania) were obtained, prepared, and submitted for use by WHC. The sorbent traps were selected based on their use by the Occupational Safety and Health Administration (OSHA) to perform workplace monitoring and because of available procedures and verification results associated with that particular application. The typical sorbent traps used consisted of a glass tube containing a sorbent material specific to the compound of interest. In general, the tubes contained two sorbent layers, or sections; the first layer was the primary trap, and the second layer provided an indication of breakthrough. In the tubes, sorbent layers are generally held in packed layers separated by glass wool. The sorbent traps, having glass-sealed ends, were received from the vendor.

The type and nominal quantity of sorbent material varied by application. Sorbent traps were selected for the tank sample job and included the following products. The NH_3 sorbent traps contained carbon beads impregnated with sulfuric acid; nominally, 500 mg were contained in the primary and 250 mg in the breakthrough sections. The NH_3 was chemisorbed as ammonium sulfate ($\{\text{NH}_4\}_2\text{SO}_4$). The NO_2 traps contained a zeolite impregnated with triethanolamine (TEA), with 400 mg in the primary and 200 mg in the breakthrough sections. The NO_2 was adsorbed and disproportionated to equi-molar quantities of nitrite ions (NO_2^-) and nitrate ions (NO_3^-). Glass tubes containing 800 mg of an oxidant such as chromate were used to convert NO to NO_2 . The converted NO was then collected as nitrite and nitrate in an NO_2 trap. The water traps contained 300 mg of silica gel in the primary and 150 mg in the breakthrough sorbent sections.

Samples provided by PNL to trap inorganic compounds include all or some of the following: samples, spiked samples, spares, single-trap blanks, and spiked blanks. The samples of each type were prepared from same-lot batches, with the oxidizer sections of the NO_x sorbent trains having been stored previously in a freezer. After sample preparation, all samples, spiked samples, blanks, and spiked blanks were stored in a freezer, primarily because of handling recommendations for the oxidizer tubes attached to some samples. After receipt of exposed and radiologically cleared samples from WHC and disassembly of the sorbent trains, samples were provided to the analytical laboratory at ambient temperature, and selected oxidizer sections were returned to a freezer until completion of analysis.

The sorbent traps were prepared in multi-trap sorbent trains configured so sample flow passed in order through the traps, targeting specific analytes, and then through a desiccant trap. The specific order of traps within the various sorbent trains is described in Section 2.4. The ends of the glass-tube traps were broken, and the traps were weighed and then connected to each other using uniform lengths of 3/8-in. perfluoroalkoxy (PFA)-grade Teflon® tubing. The tubing was heated in hot air and forced over the open ends of the traps to form a tight seal. The inlets of the sorbent trains each consisted of a short section of tubing having a 3/8-in. stainless steel Swagelok® nut, sealed using a Swagelok® cap. The trailing ends of the sorbent trains (the downstream end of the traps containing silica gel) were each sealed with red-plastic end caps provided by the manufacturer. The sorbent-tube trains remained sealed other than during the actual sampling periods. C-Flex® tubing was provided by WHC to connect the downstream ends of the sorbent trains to the sampling exhaust-manifold connections.

2.1.1 Concentration Calculations. The concentrations of target compounds in the tank headspace were determined from sample results, assuming effective sample transport to the sorbent traps. Concentration, in parts per million by volume (ppmv), was determined by dividing the mass of the compound, in μmol , by the volume of the dried tank air sampled, in mol. The micromolar sample mass was determined by dividing the compound mass, in μg , by the molecular weight of the compound, in g/mol. The molar sample volume was determined, excluding water vapor, by dividing the standard sample volume (at 0°C and 760 torr), in L, by 22.4 L/mol. For example, the concentration (C_v) of a 3.00-L sample containing 75.0 μg of ammonia equals

$$C_v = \frac{75.0 \mu\text{g}}{17 \text{ g/mol}} \left(\frac{3.00 \text{ L}}{22.4 \text{ L/mol}} \right)^{-1} = 32.9 \text{ ppmv} \quad (2.1)$$

This calculational method produces concentration results that are slightly conservative (greater than actual) because the volume of water vapor in the sample stream is neglected. The volume of water vapor is not included in the measured sampled volume because of its removal in desiccant traps upstream of the mass flowmeters. However, the bias is generally expected to be small. For a tank-headspace temperature of 35°C, the magnitude of the bias would be about 1 to 6%, assuming tank-headspace relative humidities of 20 to 100%, respectively. The concentration of mass (determined gravimetrically) was also per dry-gas volume at standard conditions.

2.2 Analytical Procedures

The compounds of interest were trapped using solid sorbents and chemisorption adsorption of water vapor. Analytical results were based on extraction and analysis of selected ions. Analytical procedures used are specified in the text. All were compiled in PNL-MA-599.

2.2.1 Ammonia Analysis. The sorbent material from the ammonia-selective sorbent traps was placed into labeled 20-mL glass scintillation vials. Vials containing front, or primary, section sorbent material were treated with 10.0 mL deionized water (DIW), and vials containing back-up section sorbent material were treated with 5.0 mL of DIW. After extraction, the NH_3 sorbent traps were analyzed using the selective ion electrode (SIE) procedure PNL-ALO-226 {Ammonia (Nitrogen) in Aqueous Samples}. Briefly, this method includes 1) preparing a 1000- $\mu\text{g/mL}$ (ppm) NH_3 stock standard solution from dried reagent-grade NH_4Cl and DIW on the day analyses are performed; 2) preparing 0.1, 0.5, 1.0, 10, and 100 ppm NH_3 working calibration standards by serial dilution of the freshly made stock standard; 3) generating an initial calibration curve from the measured emf signal versus NH_2 concentration data obtained for the set of working standards; 4) performing a

calibration-verification check, using one of the midrange standards, after analyzing every four or five samples; 5) continuing this sequence until all samples of the batch have been measured, including duplicates and spiked samples; and 6) remeasuring the complete set of calibration standards (at the end of the session). Emf signal measurements obtained for samples are compared to those for standards, either graphically or algebraically (using linear regression), to determine ammonia concentration in the samples.

2.2.2 Nitrite Analysis. The sorbent traps for NO_2 and NO were desorbed in an aqueous triethanolamine (TEA) and n-butanol solution and analyzed by suppressed-conductivity ion chromatography (SCIC) for nitrite according to PNL-ALO-212, Rev. 1 (*Determination of Inorganic Anions by Ion Chromatography*) modified to obviate interferences by concentrations of non-target analytes. Specifically, the modifications used were 1) eluent 1.44 mM Na_2CO_3 + 1.8 mM NaHCO_3 at 2.0 mL/min; 2) one guard column (AG4A) and two separator columns (AS4A) in series instead of just one separator column; 3) all standards, samples, and blanks injected into the IC sample loop through 0.45- μm syringe filters.

For the analysis, the sorbent materials were placed into labeled 20-mL glass scintillation vials. To each vial, 3.0 mL of desorbing solution (15 g TEA + 1 mL N-butanol in 1.0 L DIW) was added. Primary sorbent-trap sample materials and back-up (breakthrough) sorbent-trap materials were analyzed separately using identical procedures. Each analytical session was conducted as follows. Working nitrite standards (0, 0.1, 0.25, and 0.5 ppm) were prepared by diluting a stock nitrite standard with desorbing solution. An initial calibration curve was prepared from the instrument response (chromatographic peak height) versus nitrite standard concentration data for the set of working standards. A calibration verification check using one of the midrange standards was performed after analyzing every six samples. If the instrument response indicated that sample nitrite concentration was outside the calibration range (> 0.5 ppm nitrite), the sample was diluted with desorbing solution and reanalyzed. After all samples of a batch were analyzed, the complete set of calibration standards was remeasured to verify consistent instrument response, and the analytical session was terminated.

Instrument responses (peak height) observed for samples were compared to those for standards to determine the nitrite concentration of the samples. Because NO_2 and NO converted to NO_2 were collected on the sorbent as equal quantities of nitrite and nitrate, and the analysis was specific for nitrite, the molar masses of NO_2 and NO were determined by doubling the analytically determined molar mass of nitrite.

2.2.3 Mass (Water) Analysis. Sorbent traps used to make each sample train were weighed using a semi-micro mass balance after labeling and breaking the glass tube ends, without plastic end caps. After receipt of exposed samples, the sorbent traps were again weighed to determine the change in mass. Records of the measurements were documented on sample-preparation data sheets. The mass concentration, generally roughly equal to the concentration of water, was determined by dividing the combined change in mass from all traps in a sorbent train by the actual volume of gas sampled. Blanks and spiked blanks were included to provide information on uncertainty.

2.3 Quality Assurance/Quality Control

Analytical work was performed according to quality levels identified in the project QA plan and several PNL documents. The samples were analyzed following PNL QA impact level (IL) III. The

PNL documents include PNL-MA-70 (Part 2), PNL-MA-599, PNL-ALO-212, PNL-ALO-226, PNL-ALO-271, and MCS-033. A summary of the analysis procedures and limits for the target inorganic compounds is provided in Table 2.1. From the table, it can be seen that the minimum detection limit (MDL) required to resolve the analyte at one-tenth of the recommended exposure limit (REL) for each of the target analytes is achieved using current procedures and with a vapor-sample volume of 3 L and a desorption-solution volume of 3 mL (10 mL for ammonia).

The accuracy of concentration measurements depends on errors associated with both sampling and analysis (see Section 2.4). Sampling information, including sample volumes, was provided by WHC; sample volume uncertainty was not provided. The accuracy of analytical results depends on the method used. For ammonia analyses, the accuracy of laboratory measurements by SIE was estimated to be $\pm 5\%$ relative, independent of concentration at $1 \mu\text{g/mL}$ or greater levels. The uncertainty includes preparation of standards, purity of the ammonium salt used to prepare standards, potential operator bias, ambient temperature variations, etc. Unfortunately, no known National Institute for Standards and Technology (NIST)-traceable standard reference material (SRM) is available against which to compare working standards. As for ammonia, no known NIST SRM is available for nitrite analysis (for NO_2 and NO). Based on experience in comparing nitrite working standards prepared from several different sources and factors mentioned for ammonia above, the estimated maximum bias for samples derived from sampling for NO_2 is $\pm 10\%$, and for samples derived from sampling for NO , it is $\pm 5\%$ relative. The accuracy of measurements of sample mass is $\pm 0.05 \text{ mg}$, or much less than 1% of the mass changes of most samples, and roughly 5% or less of the mass change of most blanks.

Table 2.1 Analysis Procedures and Detection Limits of Target Inorganic Analytes

Analyte	Formula	Procedure	REL ^(a) (ppmv)	0.1 x REL ^(a) (ppmv)	MDL ^(b) (ppmv)
Ammonia	NH_3	PNL-ALO-226	25	2.5	0.5
Nitrogen dioxide	NO_2	PNL-ALO-212	1	0.1	0.02
Nitric oxide	NO	PNL-ALO-212	25	2.5	0.02
Mass (water) ^(c)	$\text{n/a}^{(d)}$	n/a	n/a	n/a	n/a

(a) Target analytical limits are equal to one-tenth of the REL.

(b) MDL is defined as the vapor concentration that can be detected with an uncertainty equal to about the magnitude of the measurement. The uncertainty is expected to reduce to about one-quarter of the magnitude of the measurement at a concentration of four times the MDL. The MDLs were based on the assumption that 3 L of vapor are sampled; if greater volumes of vapor are sampled, correspondingly smaller MDLs can be achieved. The MDLs were also based on desorbing-solution volumes of 10 mL for ammonia and 3 mL for the other analytes.

(c) The vapor-mass concentration, thought to be largely water vapor, is determined for estimates of humidity.

(d) n/a = not applicable

2.4 Inorganic Sample Results

Samples were obtained by WHC from the headspace of Tank C-108 on 8/5/94 using the VSS. The sample job designation number was S4052. Samples were prepared, submitted to WHC, and then analyzed to provide information on the concentrations of NH_3 , NO_2 , NO , and H_2O . Sampling and analysis for hydrogen cyanide (HCN) and sulfur oxides (SO_x) was not requested. The inorganic samples were received from WHC on 8/8/94; the sample-volume information was received on 9/12/94.

A list of samples, sampling information, sample volumes, and gravimetric results is shown in Table 2.2. The types of sample trains used, and the order of sorbent traps within each train, are also shown in the table. For example, the sorbent train $\text{NH}_3/\text{NO}_x/\text{H}_2\text{O}$ contained an ammonia trap at the inlet end, an NO_x series in the middle (Section 2.4.2), and a desiccant trap at the outlet end. Analytical mass and concentration results are shown in Table 2.3. Sample volumes were provided by WHC; sample-volume uncertainty was not provided. Tank-headspace concentration results (Table 2.3) are based on this information, and the listed uncertainties equal plus-or-minus one standard deviation of the individual results from each set of samples. Where analytical results from samples were nearly indistinguishable from those of blanks, indicating very low vapor concentrations of the analyte, the concentration results (Table 2.3) are listed as "less than or equal to" a probable maximum value determined by subtracting the average of the blanks less one standard deviation from the average of the samples plus one standard deviation. Results of control samples, such as spiked blanks, are discussed in this section. Spiked blanks, when used, were transported to the field but not opened. Spiked samples, when used, were opened in the field and used to collect tank vapors. Sample results were not corrected for the percentage recoveries of spiked blanks.

Table 2.2 List of PNL Inorganic Samples, Controls, and Gravimetric Results Obtained From a Heated Tube Inserted into the Headspace of Tank C-108 on 8/5/94

Sample Number	Sorbent Train Type	Sample Port and Volume Information(a)				Mass Gain (g)
		Sample Port	Flow Rate (mL/min)	Duration (min)	Volume (L)	
<u>Samples:</u>						
S4052-A23-07W	NH ₃ /NO _x /H ₂ O Train	8	200	15.0	3.00	0.0514
S4052-A24-08W	NH ₃ /NO _x /H ₂ O Train	10	200	15.0	3.00	0.0593
S4052-A25-09W	NH ₃ /NO _x /H ₂ O Train	9	200	15.0	3.00	0.0613
S4052-A26-10W	NH ₃ /NO _x /H ₂ O Train	10	200	15.0	3.00	0.0606
S4052-A27-11W	NH ₃ /NO _x /H ₂ O Train	8	200	15.0	3.00	0.0645
S4052-A28-12W	NH ₃ /NO _x /H ₂ O Train	10	200	15.0	3.00	0.0516
<u>Controls:</u>						
S4052-A29-13W	NH ₃ /NO ₂ /H ₂ O Blanks	n/a(b)	n/a	n/a	n/a	-0.0008
S4052-A30-14W	NH ₃ /NO ₂ /H ₂ O Blanks	n/a	n/a	n/a	n/a	-0.0016
S4052-A31-15W	NH ₃ /NO ₂ /H ₂ O Blanks	n/a	n/a	n/a	n/a	-0.0014

(a) Sampling information and dry-gas sample volumes, corrected to 0°C and 760 torr, were provided by WHC. Uncertainty values were not provided with sample-volume results.

(b) n/a = not applicable.

2.4.1 Ammonia Results. The concentration of NH_3 was 2.7 ± 0.3 ppmv, based on all six samples. The blank-corrected NH_3 quantities in the sorbent traps ranged from 0.32 to 0.44 μmol in the front sorbent sections with no indication of breakthrough. Blank corrections, ≤ 0.06 μmol in front and ≤ 0.03 μmol in back sorbent sections, were based on three blanks and were only significant because of the very low quantities of NH_3 found in the samples. Although spiked blanks were not tested, the percentage recoveries of three sets of blanks spiked with 12.2, 22.3, and 46.4 μmol NH_3 were $101 \pm 4\%$, $109 \pm 2\%$, and $104 \pm 1\%$, respectively, during related sample jobs (Clauss et al. 1994; Ligothke et al. 1994). The analyses of two samples were duplicated and yielded repeatabilities of $\pm 2\%$ and $\pm 10\%$. One sample leachate was spiked after initial analysis with roughly the quantity of NH_3 in the sample and yielded a percentage recovery of 84%. The relatively poor percentage recovery was a

Table 2.3. Inorganic Vapor-Sample Results Obtained From a Heated Tube Inserted into the Headspace of Tank C-108 on 8/5/94

Sample	Analytical Results (μmol)			Sample Volume (L)	Vapor ^(a) Concentration (ppmv)
	Front Section	Back Section	Total ^(b) Blank-Corrected		
<u>NH₃ Samples:</u>			<u>0.36^(c)</u>	<u>3.00^(c)</u>	<u>2.7 $\pm$ 0.3^(c)</u>
S4052-A23-07W	0.50	0.03	0.44	3.00	3.3
S4052-A24-08W	0.44	NA ^(d)	0.38	3.00	2.8
S4052-A25-09W	0.40	NA	0.34	3.00	2.5
S4052-A26-10W	0.40	0.03	0.34	3.00	2.5
S4052-A27-11W	0.41	NA	0.35	3.00	2.6
S4052-A28-12W	0.38	NA	0.32	3.00	2.4
<u>NO₂ Samples:</u>			<u>≤ 0.0029</u>	<u>3.00</u>	<u>≤ 0.04</u>
S4052-A23-07W	0.0133	0.0078	n/a	3.00	n/a
S4052-A24-08W	0.0147	0.0080	n/a	3.00	n/a
S4052-A25-09W	0.0152	0.0078	n/a	3.00	n/a
S4052-A26-10W	0.0148	0.0068	n/a	3.00	n/a
S4052-A27-11W	0.0145	0.0103	n/a	3.00	n/a
S4052-A28-12W	0.0139	0.0078	n/a	3.00	n/a
<u>NO Samples:</u>			<u>0.0164</u>	<u>3.00</u>	<u>0.24 $\pm$ 0.01</u>
S4052-A23-07W	0.0306	0.0078	0.0161	3.00	0.24
S4052-A24-08W	0.0308	0.0080	0.0163	3.00	0.24
S4052-A25-09W	0.0310	0.0074	0.0165	3.00	0.25
S4052-A26-10W	0.0306	0.0082	0.0161	3.00	0.24
S4052-A27-11W	0.0316	0.0080	0.0171	3.00	0.26
S4052-A28-12W	0.0308	0.0073	0.0163	3.00	0.24
<u>Gravimetric Samples (mg, mg/L):</u>			<u>60 mg</u>	<u>3.00</u>	<u>20 $\pm$ 2 mg/L</u>
S4052-A23-07W	n/a	n/a	53	3.00	18
S4052-A24-08W	n/a	n/a	61	3.00	20
S4052-A25-09W	n/a	n/a	63	3.00	21
S4052-A26-10W	n/a	n/a	63	3.00	21
S4052-A27-11W	n/a	n/a	66	3.00	22
S4052-A28-12W	n/a	n/a	54	3.00	18

(a) Blank-corrected vapor concentrations were calculated using WHC-reported dry-air sample volumes (corrected to 0°C and 760 torr). In the calculation for concentration, the nitrite values (listed) were doubled to account for unanalyzed nitrate. Sample results were not corrected for percentage recovery of spiked samples or spiked blanks.

(b) Total blank-corrected analyte masses (nitrite for NO₂ and NO) were determined, when significant, by subtracting the quantity of analyte found in blanks from that found in samples. The levels of analytes found in blanks are described in the subsections of Section 2.4.

(c) Underlined values represent the average of the set of samples. Concentration uncertainty equals ± 1 standard deviation (absolute) for each set of samples. The use of " $\leq$ " is defined in Section 2.4.

(d) NA = not analyzed; n/a = not applicable. Only selected back sorbent sections were analyzed.

result of the very small quantities of NH₃ in the samples. A 5-point calibration was performed over an NH₃ range of 0.1 to 1000 $\mu\text{g/mL}$.

2.4.2 Nitrogen Oxides Results. Measurements of NO_2 and NO were made using six 5-segment $\text{NH}_3/\text{NO}_x/\text{H}_2\text{O}$ sorbent-trap trains (the NO_x trains consisted of NO_2 trap, oxidizer, NO_2 trap). Related sample jobs, performed using the VSS in Tanks BY-104, -105, and -106 both with and without NO_x trains protected by a leading NH_3 trap (e.g., Clauss et al. 1994), indicated that the presence of the upstream NH_3 traps resulted in NO concentrations that were about 1.3- to 1.6-fold less than those from unprotected NO_2 traps. The NO_2 concentrations were also potentially less following an NH_3 trap.

The concentrations of NO_2 and NO were ≤ 0.04 and 0.24 ± 0.01 ppmv, respectively. Blank-corrected NO_2^- quantities in the sorbent traps averaged ≤ 0.0029 μmol (NO_2 samples) and 0.0164 μmol (NO samples). Nitrite blank levels used to correct data were 0.0145 ± 0.0002 μmol in front and 0.0076 ± 0.0004 μmol in back sorbent sections, and were based on three blanks. Although spiked blanks were not tested, blanks spiked with 0.0064 , 0.047 , 0.11 , and 0.74 μmol NO_2^- during related sample jobs yielded percentage recoveries of $153 \pm 14\%$, $103 \pm 4\%$, $106 \pm 8\%$, and $111 \pm 7\%$, respectively (Clauss et al. 1994; Ligothe et al. 1994). No samples were reanalyzed to check repeatability. No sample leachates were spiked after initial analysis with quantities of NO_2^- to test analytical percentage recoveries. A 4-point calibration was performed over a concentration range of 0 to 0.5 μg NO_2^- per mL in the desorbing matrix.

2.4.3 Gravimetric Results. The mass concentration of material collected in the sorbent-trap trains, believed to be primarily water vapor, was 20 ± 2 mg/L. The result was based on an average mass gain of 58 mg from all six $\text{NH}_3/\text{NO}_2/\text{H}_2\text{O}$ sample trains. The blank correction applied to the results was 2 ± 2 mg per sample train, based on an average per-trap mass loss of 0.4 ± 0.4 mg from nine blanks. Three traps each of NH_3 , NO_2 , and H_2O were prepared and analyzed as trip blanks. Although no spiked blanks were tested, the percentage recovery of mass from three blank H_2O traps spiked with 51 mg of water was $103 \pm 2\%$ during a related sample job (Clauss et al. 1994).

3.0 Organic Task

3.1 SUMMA™ Canister Preparation

Before sending SUMMA™ canisters out to the field for sampling, the canisters are cleaned and verified contaminant free according to PNL Technical Procedure PNL-TVP-02^(a). The cleaning procedure uses an EnTech 3000 cleaning system that controls 1) filling the canisters with purified humid air and 2) evacuating, for several cycles with applied heat, before allowing the canister to evacuate overnight. The canister is filled a final time with purified humid air for analysis by PNL Technical Procedure PNL-TVP-01^(b), which is a modification of U.S. Environmental Protection Agency (EPA) compendium Method TO-14. If the canister is verified as clean, free of TO-14 contaminants to a level of 5 parts per billion by volume (ppbv), the canister is evacuated to 30 in. Hg, tagged, and stored for use in the field. Before sending the canisters out to the field for sampling, the canisters are prehumidified with 100 µL of distilled water and labeled with a field-sampling identification. Canisters stored more than 30 but less than 60 days are re-evacuated and rehumidified before use. If stored more than 60 days, the canisters are recleaned and validated before use.

3.2 Sample Analysis Method

The SUMMA™ canister samples were analyzed according to PNL Technical Procedure PNL-TVP-03, *Determination of TO-14 Volatile Organic Compounds in Hanford Waste Tank Headspace Samples Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometry Analysis*, which is a modified version of EPA compendium Method TO-14. The method uses an EnTech cryoconcentration system interfaced with a Hewlett Packard (HP) 5971 GC/MS. The EnTech concentrator is used to pull a metered volume of sample air from the SUMMA™ canister, cryogenically concentrate the air volume, then transfer the volume to the GC/MS for analysis. A 100-mL volume of sample is measured and analyzed from the tank headspace. The organic components in the sampled air are separated on an analytical column, J&W Scientific DB-1 phase, 60-m by 0.32-mm internal diameter with 3-µm film thickness. The GC oven is programmed to run a temperature gradient beginning at 40°C, holding for 5 min, and ramping at 4°C per min to a final temperature of 260°C, with a 5-min hold.

At least twenty-four hours before analysis, the SUMMA™ canister samples were pressurized with purified air (Aadco zero air). The starting pressure was first measured using a calibrated diaphragm gauge (Cole Parmer), then pressurized to a level exactly twice the original pressure. For example, PNL canister number 014 (ambient sample without the VSS) had a start pressure of 740 torr. It was pressured to 1480 torr. This dilution was an effort to improve the reproducibility of sample aliquot pulled through the EnTech system and consequently the precision of the analysis. The sample dilution was taken into account when calculating the analysis results.

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- (a) Pacific Northwest Laboratory. 8/94. *Cleaning SUMMA™ Canisters and the Validation of the Cleaning Process*, PNL-TVP-02 (Rev. 0), PNL Technical Procedure, Richland, Washington.
  - (b) Pacific Northwest Laboratory. 8/94. *Determination of TO-14 Volatile Organic Compounds in Ambient Air Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometric Analysis*, PNL-TVP-01 (Rev. 0). PNL Technical Procedure, Richland, Washington.

3.3 Quality Assurance/Quality Control

Before the tank sample was analyzed, a diagnostic check was performed on the GC/MS instrument by running an instrument "auto tune," as described in PNL-TVP-03. Upon satisfactory completion of the instrument diagnostic check, a blank volume of purified nitrogen was analyzed to check the cleanliness of the system. The instrument was then calibrated over 6 data points ranging from 2 ppbv to 100 ppbv, using a standard gas mixture containing 40 volatile organic compounds listed in EPA compendium Method TO-14. A gas mixture containing bromochloromethane, 1,4-difluorobenzene, and chlorobenzene- d_5 was used as an internal standard (IS) for all blank, calibration standard, and sample analyses. Analyte response from sample components, ISs, and standards were obtained from the extracted ion plot from their selected mass ion. The calibration curve was generated by calculating the relative response ratios of the IS to calibration standard responses and plotting the ratios against the ratio of the calibration-standard concentration (in ppbv) to the IS concentration. A least-squares linear-regression routine was applied to the data set to generate the best-fit line for each compound. The equation for that line was then used to quantify the TO-14 compounds found in the tank samples.

3.3.1 Quantitation of TO-14 Results. The quantitative-analysis results for the TO-14 volatile organic compounds were calculated directly from the calibration curve generated using the IS method described above and in PNL-TVP-03. The conversion from ppmv to mg/m^3 assumes standard temperature and pressure (STP) conditions of 760 torr and 273K and was calculated directly from the following equation:

$$\text{mg}/\text{m}^3 = \frac{\text{ppmv} \times \text{g mol wt of compound}}{22.4 \text{ L/mole}} \quad (3.1)$$

3.3.2 Identification and Quantitation of Tentatively Identified Compounds. The tentatively identified compounds (TICs) are determined by mass-spectral interpretation and comparison of the spectra with the EPA/NIST/WILEY Library, which is a part of the GC/MS instrument operating system. Chromatographic peaks with an area count greater than, or equal to, one half of the total area count of the chlorobenzene- d_5 IS peak at the 20-ppbv calibration level are tentatively identified and quantitatively estimated. This standard was chosen to determine the integration cutoff as it is in the middle of the chromatographic range and not in a region typically affected by coelution of other compounds. The quality of the mass-spectral searches was then reviewed by the principal investigators before the identification was assigned to each chromatographic peak.

The concentration of each TIC was estimated using a relative response factor calculated using a corrected total peak area for the IS chlorobenzene- d_5 . Specifically, the total integrated area for the chlorobenzene- d_5 peak had to be corrected for possible coeluting compounds and matrix effects before calculating the response factor. The corrected total peak area for the IS was calculated by multiplying the IS quantitation ion by a correction factor based on the ratio of the total integrated peak area to the quantitation ion as measured in blank runs. The corrected peak area was then used to calculate a response factor using the IS concentration in mg/m^3 :

$$\text{Response Factor} = \frac{\text{IS conc. } (\text{mg}/\text{m}^3)}{\text{IS peak area}} \quad (3.2)$$

The calculated response factor was then multiplied by the TIC peak area to give an estimated concentration for that compound. For acetaldehyde, the total peak area was multiplied by the response factor for chlorobenzene- d_5 to give an estimated concentration of 0.10 mg/m^3 (average of three samples). Internal standards bromochloromethane and difluorobenzene were not used to quantitate the TICs because previous sample matrices appeared to have greatly altered the signal of the quantitation ions for these two ISs. By pressurizing the samples as described in Section 3.2, and increasing the relative amount of IS used in sample analysis, the quantitation ions of these two ISs are less suppressed.

The ppmv concentrations are calculated from mg/m^3 and the molecular weight of the analyte.

$$\text{TIC in ppmv} = \frac{\text{TIC} \left(\text{mg/m}^3 \right) \times 22.4 \text{ L/mole}}{\text{TIC g mol wt}} \quad (3.3)$$

The IS level added to all blank, standard, and sample injections was 91.5 ppbv for bromochloromethane, 101.5 ppbv for 1,4-difluorobenzene, and 91.0 ppbv for chlorobenzene- d_5 . The IS concentrations were converted from ppbv to mg/m^3 at STP using a molecular weight of 129.39 (g/mol) for bromochloromethane, 114.09 for 1,4-difluorobenzene, and 117.6 for chlorobenzene- d_5 . All sample concentrations were multiplied by a factor of two to account for the sample dilution step described in Section 3.2.

3.4 Analysis Results

The results from the GC/MS analysis of the tank-headspace samples are presented in Tables 3.1, 3.2, and 3.3. A representative total ion chromatogram showing the identity of major constituents is given in Figure 3.1.

Table 3.1 lists the quantitative results for compounds listed in Method TO-14. The levels of TO-14 analytes observed in the samples collected from Tank C-108 were low, close to the quantitation limit (2 ppb).

Table 3.2 lists the semi-quantitative results for the TICs observed in the samples, and Table 3.3 lists TICs observed in ambient samples. The only species observed in these samples were acetaldehyde and acetone. The normal paraffin hydrocarbons (NPH), defined as n-alkanes from C_{11} to C_{15} , were not present in the sample. The total concentration of the TICs was 93.5 mg/m^3 for all canisters analyzed. Acetaldehyde and acetone were observed in both ambient samples, as well as in all three sample canisters. These two compounds were not observed in blank runs through the GC/MS system.

4.0 Conclusions

The concentrations of selected inorganic and organic compounds were determined from samples of the headspace of Tank C-108. Sampling and analysis methods followed those described by Ligothe et al. (1994) for samples obtained from Tank C-103, a tank containing a relatively complex headspace composition. Method-validation measurements during that study did appear to validate the trapping and analysis of NH_3 , but did not eliminate the possibility of interferences that could affect NO_x results. It is recommended that additional control samples be obtained if a tank is discovered in the future to contain significant quantities of NO_x . In the current sample job, NO_x samples were obtained after first passing the sample flow through an NH_3 trap. The NH_3 concentration was found to be 2.7 ± 0.3 ppmv. The concentration of NO_2 was ≤ 0.04 ppmv. The concentration of NO was 0.24 ± 0.01 ppmv. The mass concentration was ≥ 2 mg/mL, and was expected to consist largely of water vapor.

Acetaldehyde and acetone made up about 50% of the total concentration of all the organic compounds detected. These compounds were seen in both tank and ambient samples. They were not observed in the background blank runs of the EnTech/GC/MS instrument system. The compound 1,1,2-trichloro-1,2,2-trifluoroethane (also known as FREON 113) was observed as a contaminant in the samples and the instrument blanks. This compound may not be a true constituent of Tank C-108. Generally, the organic content of these vapor samples was low when compared to all other BY and C-farm tanks reported to date. Pressurizing the canisters greatly improved the analytical reproducibility between samples, as observed in the relative standard deviations for the compounds detected.

5.0 References

Clauss, T. W., M. W. Ligothe, B. D. McVeety, K. H. Pool, R. B. Lucke, J. S. Fruchter, and S. C. Goheen. 1994. *Vapor Space Characterization of Waste Tank 241-BY-104: Results from Samples Collected on 6/24/94*. PNL-10208. Pacific Northwest Laboratory, Richland, Washington.

Ligothe, M. W., K. H. Pool, and B. D. Lerner. 1994. *Vapor Space Characterization of Waste Tank 241-C-103: Inorganic Results from Sample Job 7B (5/12/94 - 5/25/94)*. PNL-10172, Pacific Northwest Laboratory, Richland, Washington.

6.0 Further Reading

Pacific Northwest Laboratory. Analytical Laboratory Procedure Compendium. Procedures PNL-ALO-212, -226, -271. PNL-MA-599, Richland, Washington.

Pacific Northwest Laboratory. Quality Assurance Manual, Part 2: Good Practices Standard. PNL-MA-70, Part 2, Richland, Washington.

Pacific Northwest Laboratory. Quality Assurance Plan for Activities Conducted by the Analytical Chemistry Laboratory (ACL). MCS-033, Analytical Chemistry Laboratory, Richland, Washington.

Pacific Northwest Laboratory. 1994. *Determination of TO-14 Volatile Organic Compounds in Hanford Waste Tank Headspace Samples Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometry Analysis*, PNL-TVP-03 (Rev. 0), PNL Technical Procedure, Richland, Washington.

Pacific Northwest Laboratory. 1994. *Sample Shipping and Receiving Procedure - DRAFT for PNL Waste Tank Samples*. PNL-TVP-07 (Rev. 0), PNL Technical Procedure, Richland, Washington.

Table 3.1

TO-14 Analysis Results for Samples Collected from the Headspace of Tank C-108 in SUMMA™ Canisters on 8/5/94

TO-14 Analyte	Mol Wt	CAS #	S4052-A04-016 ^(a)		S4052-A06-017 ^(a)		S4052-A08-018 ^(a)		S4052-A01-014 ^(a)		S4052-A02-015 ^(a)	
			PNL 016 ^(b)		PNL 017 ^(b)		PNL 018 ^(b)		PNL 014 ^(b)		PNL 015 ^(b)	
			Concentration		Concentration		Concentration		Concentration		Concentration	
			ppbv	mg/m ³	ppbv	mg/m ³	ppbv	mg/m ³	ppbv	mg/m ³	ppbv	mg/m ³
dichlorodifluoromethane (FREON-12)	120.92	75-71-8	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011
methyl chloride (chloromethane)	50.49	74-87-3	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011	< 2	< 0.011
1,2-dichloro-1,1,2,2-tetrafluoroethane (FREON-114)	170.93	76-14-2	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02
chloroethene (vinyl chloride)	62.50	75-01-4	< 2	< 0.006	< 2	< 0.006	< 2	< 0.006	< 2	< 0.006	< 2	< 0.006
methyl bromide (bromomethane)	94.95	74-83-9	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
ethyl chloride	64.52	75-00-3	8.40	0.050	8.47	< 0.006	8.40	0.050	8.43	0.050	8.40	0.020
trichlorofluoromethane (FREON-11)	137.38	75-69-4	9.55	0.060	9.50	< 0.006	9.29	0.060	6.35	0.040	6.39	0.040
1,1-dichloroethene (1,1-dichloroethylene)	96.95	75-35-4	3.53	0.020	< 2	< 0.009	3.53	0.020	3.52	0.020	3.55	0.020
dichloromethane (methylene chloride)	84.94	75-09-2	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
1,1,2-trichloro-1,2,2-trifluoroethane (FREON-113) ^(c)	187.40	76-13-1	8.00	0.050	9.17	0.060	8.79	< 0.02	< 2	< 0.02	5.05	0.040
1,1-dichloroethane	99.00	75-34-3	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
cis-1,2-dichloroethene (cis-1,2-dichloroethylene)	96.90	156-60-5	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
trichloromethane (chloroform)	119.40	67-66-3	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
1,2-dichloroethane	99.00	107-06-2	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
1,1,1-trichloroethane	133.40	71-55-8	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
benzene	78.10	71-43-2	< 2	< 0.009	< 2	< 0.009	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02
carbon tetrachloride	153.80	56-23-5	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
1,2-dichloropropane	113.00	78-87-5	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
trichloroethylene	131.40	79-01-6	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
cis-1,3-dichloropropene	111.00	10081-01-6	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
trans-1,3-dichloropropene	111.00	10061-02-5	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
1,1,2-trichloroethane	133.40	79-00-5	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
methyl benzene (toluene)	92.14	108-88-3	< 2	< 0.008	< 2	< 0.008	< 2	< 0.008	< 2	< 0.008	< 2	< 0.008
1,2-dibromoethane	187.90	106-93-4	2.20	0.010	2.18	0.010	2.18	0.020	2.23	0.020	2.20	0.020
tetrachloroethene (tetrachloroethylene)	165.83	127-18-4	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02
chlorobenzene	112.60	108-90-7	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
ethylbenzene	106.20	100-41-4	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
p/m-xylene (1,3-dimethylbenzene)	106.17	108-38-3	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
styrene	104.20	106-42-3	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
1,1,2,2-tetrachloroethane	167.90	106-42-3	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02
o-xylene (1,2-dimethylbenzene)	106.17	100-42-5	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009	< 2	< 0.009
1,3,5-trimethylbenzene	120.20	79-34-5	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
1,2,4-trimethylbenzene	120.20	95-47-6	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
chloromethylbenzene, alpha (benzyl chloride)	126.60	108-67-8	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
m-dichlorobenzene (1,3-dichlorobenzene)	147.00	95-63-6	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
p-dichlorobenzene (1,4-dichlorobenzene)	147.00	100-44-7	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
o-dichlorobenzene (1,2-dichlorobenzene)	147.00	541-73-1	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01	< 2	< 0.01
1,2,4-trichlorobenzene	181.45	106-46-7	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	2.21	0.020	2.01	0.020
hexachloro-1,3-butadiene	260.80	95-50-1	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02	< 2	< 0.02

(a) WHC sample identification number.

(b) PNL canister number.

(c) Analytical system contaminant

Table 3.2 Table of Tentatively Identified Compounds and Estimated Concentrations^(a) in Hanford C-108 SUMMA^{TM} Canister Samples Collected on 8/5/94

Tentatively Identified Compounds ^(d)	CAS # ^(d)	Mol Wt	Ret Time (min)	S4052-A04-016 ^(b) PNL 016 ^(c) Concentration		S4052-A06-017 ^(b) PNL 017 ^(c) Concentration		S4052-A04-018 ^(b) PNL 018 ^(c) Concentration		Means and Standard Deviation (mg/m ³)	
				mg/m ³	ppmv	mg/m ³	ppmv	mg/m ³	ppmv	Mean	stdev
carbon dioxide	124-38-9	44	3.2	e	e	e	e	e	e	e	e
acetaldehyde	75-07-0	55	5.2	0.090	0.050	0.120	0.060	0.080	0.040	0.10	0.02
acetone	110-62-3	58	8.5	0.080	0.040	0.120	0.060	0.060	0.030	0.09	0.03

(a) Semi-quantitative estimate calculated using concentration of closest eluting internal standard.

(b) WHC sample number.

(c) PNL SUMMA^{TM} canister number.

(d) Obtained by mass spectral interpretation and comparison with the EPA/NIST and WILEY Libraries.

(e) Carbon Dioxide cannot be determined by the analytical method used.

Table 3.3 Tentatively Identified Compounds and Estimated Concentrations^(a) in Ambient Air Samples Collected
Near Hanford Tank C-108 on 8/5/94

Tentatively Identified Compounds ^(d)	CAS # ^(d)	Mol Wt	Ret Time (min)	S4052-A01-014 ^(b) PNL 014 ^(c) Upwind sample		S4052-A02-015 ^(b) PNL 015 ^(c) Through VSS	
				mg/m ³	ppmv	mg/m ³	ppmv
carbon dioxide	124-38-9	44	3.1	e	e	e	e
acetaldehyde	75-07-0	55	5.2	0.06	0.02	0.07	0.03
acetone	110-62-3	58	8.5	0.07	0.03	0.48	0.19
acetophenone	98-86-2	128	9.1	0.08	0.01	0.06	0.01

(a) Semi-quantitative estimate calculated using concentration of closest eluting internal standard.

(b) WHC sample number.

(c) PNL SUMMA^{TM} canister number.

(d) Obtained by mass spectral interpretation and comparison with the EPA/NIST/WILEY Library.

(e) Carbon Dioxide cannot be determined by the analytical method used.

(f) Concentration information for internal standards are determined by direct calibration.

Figure 3.1 Total Ion Chromatogram of SUMMA™ Canister Sample S4052-A04-016 Collected from the Headspace of Tank C-108 on 8/5/94