**Vapor Space Characterization of Waste
Tank 241-BY-104 (*In Situ*): Results
from Samples Collected on 4/22/94**

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October 1995

Prepared for Westinghouse Hanford Company
under a Related Services Agreement
with the U.S. Department of Energy
Contract DE-AC06-76RLO 1830

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Summary

This report describes inorganic and organic analyses results from *in situ* samples obtained from the headspace of the Hanford waste storage Tank 241-BY-104 (referred to as Tank BY-104). The results described here were obtained to support safety and toxicological evaluations. A summary of the results for inorganic and organic analytes is listed in Table 1. Detailed descriptions of the results appear in the text.

Quantitative results were obtained for the inorganic compounds NH_3 , NO_2 , NO , HCN , and H_2O . Sampling for sulfur oxides was not requested. Several organic compounds were also quantitatively determined. Eighty-nine tentatively identified organic analytes were observed above the detection limit of (ca.) 10 ppbv, but standards for most of these were not available at the time of analysis, and the reported concentrations are semi-quantitative estimates. In addition, the 41 standard TO-14 analytes were sought. Of these, only a few were observed above the 2-ppbv detection limit. The 10 organic analytes with the highest estimated concentrations are listed in Table 1. These 10 analytes account for approximately 48% of the total organic components in the headspace of Tank BY-104. Detailed results appear in the text.

Table 1. Summary Results of Inorganic and Organic Samples Collected from the Headspace of Tank BY-104 on 4/22/94

Category	Analyte	Vapor ^(a) Concentration	Units
Inorganic	NH_3	285 ± 4	ppmv
	NO_2	≤ 0.07	ppmv
	NO	≤ 0.18	ppmv
	HCN	≤ 0.007	ppmv
	H_2O	17.4 ± 1.7	mg/L
Organic	Acetone	2.85	mg/m^3
	n-Tridecane	2.63	mg/m^3
	2-Methylpentane	2.08	mg/m^3
	n-Dodecane	2.08	mg/m^3
	Propene	1.68	mg/m^3
	n-Hexane	1.67	mg/m^3
	n-Pentane	1.54	mg/m^3
	2,6-Dimethylundecane	1.50	mg/m^3
	Trichlorofluoromethane	1.42	mg/m^3
	n-Tetradecane	1.18	mg/m^3

(a) Vapor concentrations were determined using sample-volume data provided by Westinghouse Hanford Company and are based on averaged data.

Acknowledgments

The authors gratefully acknowledge the support of other project staff at Pacific Northwest Laboratory who contributed to the successful completion of this sampling and analysis activity. Jeff Edwards served as the PNL single-point-of-contact and coordinated sample handling and communications with Westinghouse Hanford Company. Sally Slate and May-Lin Thomas analyzed inorganic samples, and Gary Dennis prepared the solid-sorbent sample trains. Georgia K. Ruebsamen provided word processing support.

Abbreviations

CAS	Chemical Abstracts Service
COC	chain of custody
C _v	concentration by volume
DIW	deionized water
emf	electromotive force
EPA	U.S. Environmental Protection Agency
GC/MS	gas chromatography/mass spectrometry
HP	Hewlett Packard
IC	ion chromatography
IL	impact level
IS	internal standard
ISS	<i>in situ</i> sampling system
MDL	minimum detection limit
NIST	National Institute for Standards and Technology
NPH	normal paraffin hydrocarbon
OSHA	Occupational Safety and Health Administration
PFA	perfluoroalkoxy
PNL	Pacific Northwest Laboratory
ppbv	part per billion by volume
ppmv	part per million by volume
QA	quality assurance
REL	recommended exposure limit
SCIC	suppressed-conductivity ion chromatography
SIE	selective ion electrode
SRM	standard reference material
STP	standard temperature and pressure
TEA	triethanolamine
TIC	tentatively identified compound
VSS	vapor sampling system
WHC	Westinghouse Hanford Company

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1.0 Introduction

This report describes results of the analyses of tank-headspace samples taken from the Hanford waste Tank 241-BY-104 (referred to as Tank BY-104). The Pacific Northwest Laboratory (PNL)^(a) contracted with Westinghouse Hanford Company (WHC) to provide sampling devices and to analyze inorganic and organic analytes collected from the tank headspace. The sample job was designated S4062, and samples were collected by WHC on April 22, 1994, using the *in situ* sampling system (ISS).

Sampling devices, including six sorbent trains (for inorganic analyses), were supplied to WHC on March 28, 1994. One SUMMATM canister (for organic analyses) was supplied to the WHC sampling staff on March 23. Samples were taken (by WHC) from the tank headspace on April 22 and were returned to PNL from the field on May 12. Inorganic (sorbent trap) samples were delivered to PNL on chain of custody (COC)/sample analysis request 006862 (see Figure 1.1a). One SUMMATM canister was delivered on COC/sample analysis request No. 006852 (see Figure 1.1b). The sampling date (4/28/94) on the COCs is incorrect as per internal communication from WHC dated March 23, 1995 (see Figure 1.1c).

The samples were inspected upon delivery to the 326/23B laboratory and logged into PNL record book 55408 before implementation of PNL Technical Procedure PNL-TVP-07^(b). Custody of the sorbent traps was transferred to PNL personnel performing the inorganic analysis and stored at refrigerated ($\leq 10^{\circ}\text{C}$) temperature until the time of analysis. The canister was stored in the 326/23B laboratory at ambient (25°C) temperature until time of analysis. Access to the 326/23B laboratory is limited to PNL personnel working on the waste-tank safety program. Analyses described in this report were performed at PNL in the 300 area of the Hanford Reservation. Analytical methods that were used are described in the text. In summary, sorbent traps for inorganic analyses containing sample materials were either weighed (for water analysis) or desorbed with the appropriate aqueous solutions (for NH_3 , NO_x , or HCN analyses). The aqueous extracts were analyzed either by selective ion electrode or by ion chromatography (IC). Organic analyses were performed using cryogenic preconcentration followed by gas chromatography/mass spectrometry (GC/MS).

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- (a) Pacific Northwest Laboratory is operated for the U. S. Department of Energy by Battelle Memorial Institute under Contract DE-AC06-76RLO 1930.
 - (b) PNL-TVP-07, Rev. 0, October 1994, *Sample Shipping and Receiving Procedure for PNL Waste Tank Samples*, PNL-Technical Procedure, Tank Vapor Project, Richland, Washington.

Figure 1.1a Chain-of-Custody for Inorganic Samples from Tank BY-104

Westinghouse Hanford Company		CHAIN OF CUSTODY/SAMPLE ANALYSIS REQUEST				Page <u>1</u> of <u>2</u>	
Collector: S.M. Brown		Company Contact: L.A. Pinnel		Telephone No. (509) 373-6051		Data Turnaround ☐ Priority ☒ Normal	
Project Designation: 241-BY-104		Sampling Location: 200 EAST TANK FARM; VAPOR SAMPLE		SAF No. 54011		Date: 6 May 94 COC 006862 (P)	
Ice Chest No. N/A		Field Logbook No. VHC-9-797		Method of Shipment: GOVERNMENT TRUCK		BB of Lading/Air BB No. N/A	
Shipped To: VHC-PNL		Offsite Property No. N/A		BB of Lading/Air BB No. N/A			
Possible Sample Hazards/Remarks: UNKNOWN AT TIME OF SAMPLING		Preservative: NONE		Type of Container: SORBANT SUMMA			
		No. of Container(s): 1					
Special Handling and/or Storage: 		Volume: --					
SAMPLE ANALYSIS		INORG ORG					
Sample No.	Matrix*	Date Sampled	Time Sampled				
⁶² 54011-162.X31 ✓	VAPOR	04/28/94	1345	X			
⁶² 54011-162.X32 ✓	VAPOR	04/28/94	1345	X			
⁶² 54011-162.X33 ✓	VAPOR	04/28/94	1345	X			
⁶² 54011-162.X34 ✓	VAPOR	04/28/94	1345	X			
⁶² 54011-162.X35 ✓	VAPOR	04/28/94	1345	X			
⁶² 54011-162.X36 ✓	VAPOR	04/28/94	1345	X			
SIGNATURE NAMES				SPECIAL INSTRUCTIONS			
Relinquished By: J.A. Edwards		Date/Time: 3/28/94 @ 1345		Received By: S.M. Brown		Date/Time: 3/28/94 @ 1345	
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cc:Mail for: Margaret McCulloch

Mike, Margaret, Jeff,

BY-104 was sampled using the ISS on 22 APR 94. The date of 28 APR 94 listed on the Chain of Custody is a mistake.

I'm sorry it took me so long to get back to you on this.

If you have any other questions or comments, please let me know.

Thanks,

Rick Mahon

RECEIVED

MAR 23 1995

M. McCULLOCH

Figure 1.1c Confirmation of WHC Sampling Date for Tank BY-104

2.0 Inorganic Analysis

Solid sorbent traps, prepared in sampling trains, were supplied to WHC for sampling the tank headspace. Blanks, spiked blanks (when requested), and exposed samples were returned to PNL for analysis. Analyses were performed to provide information on the tank-headspace concentration of the following analytes: ammonia (NH_3), nitrogen dioxide (NO_2), nitric oxide (NO), hydrogen cyanide (HCN), and water. Procedures were similar to those developed previously during sample jobs performed with the vapor sampling system (VSS) connected to the headspace of Tank C-103 (Ligotke et al. 1994). Analytical accuracy was estimated based on procedures used. Sample preparation and analyses were performed following PNL quality assurance (QA) impact level (IL) III requirements.

2.1 Standard Sampling Methodology

Standard sorbent traps consisting of glass tubes containing sorbent materials to trap the analytes of NH_3 , NO_2 , NO , HCN , and H_2O (SKC Inc., Eighty Four, Pennsylvania) were obtained, prepared, and submitted for use by WHC. The sorbent traps were selected based on their use by the Occupational Safety and Health Administration (OSHA) to perform workplace monitoring, and because of available procedures and verification results associated with that particular application. The typical sorbent traps used consisted of a glass tube containing a sorbent material specific to the compound of interest. In general, the tubes contained two sections; the first section was the primary trap, and the second section provided an indication of breakthrough. In the tubes, sorbent sections are generally held in packed layers separated by glass wool. The sorbent traps were connected end-to-end to prepare multi-trap sorbent trains for sampling.

The type and nominal quantity of sorbent material varied by application. Sorbent traps selected for the tank sample job included the following products. The NH_3 sorbent traps contained carbon beads impregnated with sulfuric acid; nominally, 500 mg were contained in the primary and 250 mg in the breakthrough sections. The NH_3 was chemisorbed as ammonium sulfate $\{(\text{NH}_4)_2\text{SO}_4\}$. The NO_2 traps contained a zeolite impregnated with triethanolamine (TEA), with 400 mg in the primary and 200 mg in the breakthrough sections. The NO_2 was absorbed and disproportionated to equi-molar quantities of nitrite ions (NO_2^-) and nitrate ions (NO_3^-). Glass tubes containing 800 mg of an oxidant such as chromate were used to convert NO to NO_2 . The converted NO was then collected as nitrite and nitrate in a NO_2 trap. The HCN traps contained soda lime, with 600 mg in the primary and 200 mg in the breakthrough sections. The water traps contained 300 mg of silica gel in the primary and 150 mg in the breakthrough sorbent sections.

Sample materials for inorganic analytes include some or all of the following: samples, spiked samples, spares, blanks, and spiked blanks. The samples of each type were prepared from same-lot batches, with the oxidizer sections of the NO_x sorbent trains having been stored previously in a freezer. After sample preparation, all samples, spiked samples, blanks, and spiked blanks were stored in a freezer, primarily because of handling recommendations for the oxidizer tubes attached to some samples. After receipt of exposed and radiologically cleared samples from WHC and disassembly of the sorbent trains, samples were provided to the analytical laboratory at ambient temperature, and selected oxidizer sections were returned to a freezer until completion of analyses.

The sorbent traps were prepared in multi-trap sorbent trains configured so sample flow passed in order through the traps, targeting specific analytes, and then through a desiccant trap. The specific

order of traps within the various sorbent trains is described in Section 2.4. The ends of the glass-tube traps were broken, and the traps were weighed and then connected to each other using uniform lengths of 3/8-in. perfluoroalkoxy (PFA)-grade Teflon® tubing. The tubing was heated in hot air and forced over the open ends of the traps to form a tight seal. Both the inlet and outlet ends of the sorbent trains (the downstream ends of the traps always contained silica gel) were sealed with red-plastic end caps provided by the manufacturer. The leading and trailing ends of the sorbent traps remained sealed other than during the actual sampling periods. C-Flex® tubing was provided by WHC to connect the downstream ends of the sorbent trains to the sampling exhaust-manifold connections.

2.1.1 Concentration Calculations. The concentrations of target compounds in the tank headspace were determined from sample results, assuming effective sample transport to the sorbent traps. Concentration, in parts per million by volume (ppmv), was determined by dividing the mass of the compound, in μmol , by the volume of the dried tank air sampled in mol. The micromolar compound mass was determined by dividing the compound mass, in μg , by the molecular weight of the compound, in g/mol. The molar sample volume was determined, excluding water vapor, by dividing the standard sample volume (at the conditions used by WHC, 21.1°C and 760 torr), in L, by 24.1 L/mol. For example, the concentration (C_v) of a 3.00-L sample containing 75.0 μg of ammonia equals

$$C_v = \frac{75.0 \mu\text{g}}{17 \text{ g/mol}} \left(\frac{3.00 \text{ L}}{24.1 \text{ L/mol}} \right)^{-1} = 35.4 \text{ ppmv} \quad (2.1)$$

This calculational method produces concentration results that are slightly conservative (greater than actual) because the volume of water vapor in the sample stream is neglected. The volume of water vapor is not included in the measured sampled volume because of its removal in desiccant traps upstream of the mass flowmeters. However, the bias is generally expected to be small. For a tank-headspace temperature of 35°C, the magnitude of the bias would be about 1 to 6%, assuming tank-headspace relative humidities of 20 to 100%, respectively. The concentration of mass (determined gravimetrically) was also per dry-gas volume at standard conditions.

2.2 Analytical Procedures

The compounds of interest were trapped using solid sorbents and chemisorption (adsorption of water vapor). Analytical results were based on extraction and analysis of selected ions. Analytical procedures used are specified herein and compiled in PNL-MA-599.

2.2.1 Ammonia Analysis. The sorbent material from the ammonia-selective sorbent traps was placed into labeled 20-mL glass scintillation vials. Vials containing front-, or primary-, section sorbent material were treated with 10.0 mL of deionized water (DIW), and vials containing back-up-section sorbent material were treated with 5.0 mL of DIW. After extraction, the NH_3 sorbent traps were analyzed using the selective ion electrode (SIE) procedure PNL-ALO-226 {Ammonia (Nitrogen) in Aqueous Samples}. Briefly, this method includes 1) preparing a 1000 $\mu\text{g/mL}$ (ppm) NH_3 stock standard solution from dried reagent-grade NH_4Cl and DIW on the day analyses are performed; 2) preparing 0.1-, 0.5-, 1.0-, 10-, and 100-ppm NH_3 working calibration standards by serial dilution of the freshly made stock standard; 3) generating an initial calibration curve from the measured electromotive force (emf) signal versus NH_3 concentration data obtained for the set of working standards; 4) performing a calibration-verification check, using one of the midrange standards, after analyzing every four or five samples; 5) continuing this sequence until all samples of

the batch have been measured, including duplicates and spiked samples; and 6) remeasuring the complete set of calibration standards at the end of the session. Emf signal measurements obtained for samples are compared to those for standards, either graphically or algebraically (using linear regression) to determine ammonia concentration in the samples.

2.2.2 Nitrite Analysis. The sorbent traps for NO_2 and NO were desorbed in an aqueous TEA and n-butanol solution and analyzed by suppressed-conductivity ion chromatography (SCIC) for nitrite according to PNL-ALO-212, Rev. 1 (*Determination of Inorganic Anions by Ion Chromatography*) and modified to obviate interferences by concentrations of non-target analytes. Specifically, the modifications used were 1) eluent 1.44 mM Na_2CO_3 + 1.8 mM NaHCO_3 at 2.0 mL/min, 2) one guard column (AG4A) and two separator columns (AS4A) in series instead of just one separator column, and 3) all standards, samples, and blanks injected into the IC sample loop through 0.45- μm syringe filters.

For the analysis, the sorbent materials were placed into labeled 20-mL glass scintillation vials. To each vial, 3.0 mL of desorbing solution (15 g TEA + 1 mL n-butanol in 1.0 L DIW) was added. Primary sorbent-tube sample materials and back-up (breakthrough) sorbent-tube materials were analyzed separately using identical procedures. Each analytical session was conducted as follows. Working nitrite standards (0, 0.1, 0.25, and 0.5 ppm) were prepared by diluting a stock nitrite standard with desorbing solution. An initial calibration curve was prepared from the instrument response (chromatographic peak height) versus nitrite standard concentration data for the set of working standards. A calibration verification check using one of the midrange standards was performed after the analysis of every six samples. If the instrument response indicated that sample nitrite concentration was outside the calibration range (> 0.5 ppm nitrite), the sample was diluted with desorbing solution and reanalyzed. After all samples of a batch were analyzed, the complete set of calibration standards was remeasured to verify consistent instrument response, and the analytical session was terminated. Instrument responses (peak height) observed for samples were compared to those for standards to determine the nitrite concentration of the samples. Because NO_2 and NO converted to NO_2^- were collected on the sorbent as equal quantities of nitrite and nitrate, and the analysis was specific for nitrite, the molar masses of NO_2 and NO were determined by doubling the analytically determined molar mass of nitrite.

2.2.3 Cyanide Analysis. The HCN samples were desorbed in 3.0 mL of 0.02 N sodium hydroxide and analyzed by amperometric detection IC according to PNL-ALO-271 (*Procedure for Analysis of Free cyanide in Water and Soil Sample Leachates*). Calibration standards, typically 0, 20, 50, and 100 ng/mL (ppb) CN^- , were prepared from a stock 1000-ppm CN^- standard on the day of sample analysis in 0.02 N NaOH matrix. The same analysis sequence described above in the "nitrite analysis" section was used. Instrument responses (peak height) observed for the samples were compared to those for standards to determine the CN^- concentration of the samples.

2.2.4 Mass (Water) Analysis. Sorbent traps used to make each sample train were weighed using a semi-micro mass balance, after labeling and breaking the glass tube ends, without plastic end caps. After receipt of exposed samples, the sorbent traps were again weighed to determine the change in mass. Records of the measurements were documented on sample-preparation data sheets. The mass concentration, generally roughly equal to the concentration of water, was determined by dividing the combined change in mass from all traps in a sorbent train by the actual volume of gas sampled. Blanks and spiked blanks were included to provide information on uncertainty.

2.3 Quality Assurance/Quality Control

Analytical work was performed according to quality levels identified in the project QA plan and several PNL documents. The samples were analyzed following PNL IL III. The PNL documents include some or all of the following: PNL-MA-70 (Part 2), PNL-MA-599, PNL-ALO-212, PNL-ALO-226, PNL-ALO-271, and MCS-033. A summary of the analysis procedures and limits for the target inorganic compounds is provided in Table 2.1. From the table, it can be seen that the minimum detection limit (MDL) required to resolve the analyte at one-tenth of the recommended exposure limit (REL) for each of the target analytes is achieved using current procedures and with a vapor-sample volume of 3 L and a desorption-solution volume of 3 mL (10 mL for ammonia).

Table 2.1. Analysis Procedures and Typical Detection Limits of Target Inorganic Analytes. Not all analytes are included in every sample job.

Analyte	Formula	Procedure	REL ^(a) (ppmv)	.01 x REL ^(a) (ppmv)	MDL ^(b) (ppmv)
Ammonia	NH ₃	PNL-ALO-226	25	2.5	0.5
Nitrogen dioxide	NO ₂	PNL-ALO-212	1	0.1	0.02
Nitric oxide	NO	PNL-ALO-212	25	2.5	0.02
Sulfur oxide	SO _x	PNL-ALO-212	2	0.2	0.02
Hydrogen cyanide	HCN	PNL-ALO-271	4.8	0.48	0.01
Mass (water) ^(c)	n/a	n/a	n/a	n/a	n/a

(a) Target analytical limits are equal to one-tenth of the REL.

(b) MDL is defined as the vapor concentration that can be detected with an uncertainty equal to about the magnitude of the measurement. The uncertainty is expected to reduce to about one-quarter of the magnitude of the measurement at a concentration of four times the MDL. The MDLs were based on the assumption that 3 L of vapor are sampled; if greater volumes of vapor are sampled, correspondingly smaller MDLs can be achieved. The MDLs were also based on desorbing-solution volumes of 10 mL for ammonia and 3 mL for the other analytes.

(c) The vapor-mass concentration, thought to be largely water vapor, is determined for estimates of humidity.

The accuracy of concentration measurements depends on errors associated with both sampling and analysis (see Section 2.4). Sampling information, including sample volumes, was provided by WHC; sample volume uncertainty was not provided. The accuracy of analytical results depends on the method used. For ammonia analyses, the accuracy of laboratory measurements by SIE was estimated to be $\pm 5\%$ relative, independent of concentration at 1 $\mu\text{g/mL}$ or greater levels. The uncertainty includes preparation of standards, purity of the ammonium salt used to prepare standards, potential operator bias, ambient temperature variations, etc. Unfortunately, no known National Institute for Standards and Technology (NIST)-traceable standard reference material (SRM) is available against which to compare working standards. As for ammonia, no known NIST SRM is available for nitrite analysis (for NO₂ and NO). Based on experience in comparing nitrite working standards prepared from several different sources and factors mentioned for ammonia above, the estimated maximum bias for samples derived from sampling for NO₂ is $\pm 10\%$, and for samples derived from sampling for NO, it is $\pm 5\%$ relative. For SO_x analysis, the IC determination of sulfate derived from SO_x trapped in sorbent traps is judged to be biased no more than 10% even at the very low concentrations of solutions actually analyzed. Comparison of working standards to a NIST SRM solution for sulfate exhibit no more than a 2 to 3% difference at concentrations of 2 ppm or greater.

For HCN analyses, a NIST SRM for uncomplexed cyanide is not available. The estimated bias (accuracy) of the free cyanide measurements is no more than 5% relative for the normal working range (which encompasses the concentration levels encountered in blanks and samples derived from sorbent-trap leachates). The accuracy of measurements of sample mass is ± 0.05 mg, or much less than 1% of the mass changes of samples, and roughly 5% or less of the mass change of blanks.

2.4 Inorganic Sample Results

Sorbent-trap trains and controls were prepared on 3/25/94 and submitted to WHC on 3/28/94. They were used by WHC to sample the headspace of Tank BY-104 on 4/28/94 using the ISS. The sample job designation number was S4062. The exposed samples were returned to PNL on 5/12/94 and subsequently analyzed on 6/8/94 (H_2O), 6/14/94 (NH_3 and HCN), and 7/1/94 (NO_x) to provide information on the tank-headspace concentrations of selected inorganic compounds. Sampling and analysis for sulfur oxides were not requested. Blank and spiked-blank samples associated with this sample job were those sent for the related ISS sample job in Tank BY-111 (S4028) or related sample jobs. The sample-volume information for the current sample job was received from WHC on 8/2/94.

A list of samples, sampling information, sample volumes, and gravimetric results is shown in Table 2.2. The types of sample trains used and the order of sorbent traps within each train are also shown in the table. For example, the sorbent train $\text{NH}_3/\text{NO}_x/\text{H}_2\text{O}$ contained an ammonia trap at the inlet end, a NO_x series in the middle (Section 2.4.2), and a desiccant trap at the outlet end. Analytical mass and concentration results are shown in Table 2.3. Sample volumes were provided by WHC; sample-volume uncertainty was not provided. Tank-headspace concentration results (Table 2.3) are based on this information, and the listed uncertainties equal plus-or-minus one standard deviation of the individual results from each set of samples. Where analytical results from samples were nearly indistinguishable from those of blanks, indicating very low tank-headspace concentrations of the analyte, the concentration results (Table 2.3) are listed as "less than or equal to" a probable maximum value determined by subtracting the average of the blanks less one standard deviation from the average of the samples plus one standard deviation. Results of control samples, such as spiked blanks, are discussed in this section. Spiked blanks, when used, were transported to the field but not opened. Spiked samples, when used, were opened in the field and used to collect tank-headspace gas. Sample results were not corrected for the percentage recoveries of spiked blanks.

2.4.1 Ammonia Results. The concentration of NH_3 was 285 ± 4 ppmv, based on all three samples. The blank-corrected NH_3 quantities in the sorbent traps ranged from 42 to 46 μmol with no indication of breakthrough. Blank corrections, ≤ 0.06 μmol in front and ≤ 0.03 μmol in back sorbent sections (Clauss et al. 1994), were not significant ($<2\%$). Although spiked blanks were not tested, the percentage recoveries of three sets of blanks spiked with 12.2, 22.3, and 46.4 μmol NH_3 were $101 \pm 4\%$, $109 \pm 2\%$, and $104 \pm 1\%$, respectively, during related sample jobs (Clauss et al. 1994; Ligothke et al. 1994). The analysis of one sample was duplicated and yielded a repeatability of $\pm 4\%$. No sample leachates were spiked after initial analysis to determine a percentage recovery. A 5-point calibration was performed over an NH_3 range of 0.1 to 1000 $\mu\text{g/mL}$. Minor differences between the individual results of the three NH_3 samples matched closely the differences in the quantities of dry air sampled during each sample.

2.4.2 Nitrogen Oxides Results. Measurements of NO_2 and NO were made using three 5-segment $\text{NH}_3/\text{NO}_x/\text{H}_2\text{O}$ sorbent-trap trains (the NO_x trains consisted of NO_2 trap, oxidizer, NO_2 trap). Related sample jobs, performed using the VSS in BY-104, -105, and -106 both with and without NO_x trains protected by a leading NH_3 trap (e.g., Clauss et al. 1994), indicated that the presence of the upstream

Table 2.2 List of PNL Inorganic Samples, Blanks, and Gravimetric Results From *In Situ* Sampling of the Headspace of Tank BY-104 on 4/22/94

Sample Number	Sorbent Train Type	Sample Flow Rate (mL/min)	Duration (min)	Sample Volume ^(a) (L)	Mass Gain (g)
S4062-162-X34	NH ₃ /NO _x /H ₂ O Sample	250	15.0	3.75	NA ^(b)
S4062-162-X35	NH ₃ /NO _x /H ₂ O Sample	263	15.0	3.94	NA
S4062-162-X36	NH ₃ /NO _x /H ₂ O Sample	237	15.0	3.55	NA
S4062-162-X31	HCN/H ₂ O Sample	300	15.0	4.50	0.0798
S4062-162-X32	HCN/H ₂ O Sample	363	15.0	5.45	0.0898
S4062-162-X33	HCN/H ₂ O Sample	240	15.0	3.60	0.0727
S4028-028-X66	HCN/H ₂ O Blank (BY-111)	n/a	n/a	n/a	0.0031
S4028-028-X67	HCN/H ₂ O Blank (BY-111)	n/a	n/a	n/a	0.0033
S4028-028-X68	HCN/H ₂ O Blank (BY-111)	n/a	n/a	n/a	0.0033

(a) Sampling information and dry-gas sample volumes, corrected to 21°C and 760 torr, were provided by WHC. Uncertainty values were not provided with sample volume results.

(b) NA = not analyzed. Only selected back sorbent sections were analyzed; n/a = not applicable.

NH₃ traps resulted in NO concentrations that were about 1.3- to 1.6-fold less than those from unprotected NO₂ traps. The NO₂ concentrations were also potentially less following an NH₃ trap.

The concentrations of NO₂ and NO were ≤ 0.07 and $\leq 0.18 \pm 0.05$ ppmv, respectively. Blank-corrected NO₂⁻ quantities in the sorbent traps averaged ≤ 0.0054 μmol (NO₂ samples) and 0.014 μmol (NO samples). Nitrite blank levels used to correct data were 0.016 ± 0.002 μmol in front and 0.0087 ± 0.0007 μmol in back sorbent sections, and were based on typical values such as those reported by Clauss et al. (1994). Although spiked blanks were not tested, blanks spiked with 0.0064, 0.047, 0.11, and 0.74 μmol NO₂⁻ during a related sample job yielded percentage recoveries of 153 ± 14 , $103 \pm 4\%$, 106 ± 8 , and $111 \pm 7\%$, respectively (Ligotke et al. 1994; Clauss et al. 1994). No samples were reanalyzed to check repeatability. No sample leachates were spiked after initial analysis with quantities of NO₂⁻ to test analytical percentage recoveries. A 4-point calibration was performed over a concentration range of 0 to 0.5 μg NO₂⁻/mL in the desorbing matrix.

2.4.3 Hydrogen Cyanide Results. The concentration of HCN was ≤ 0.007 ppmv, based on all three samples. Blank-corrected sample results averaged ≤ 0.0013 μmol . Blank corrections were based on the results of three blanks from a related sample job (Tank BY-111) that averaged 0.0128 ± 0.0005 μmol for the front and 0.0049 ± 0.0002 μmol for the back sorbent sections. Three spiked-blank sorbent traps were prepared by adding 50 μL of 5-ppm CN⁻ in water, confirmed gravimetrically to average 0.0099 ± 0.0001 μmol CN⁻ {e.g., $0.0520 \text{ g} \times 1.00 \text{ mL/g} \times 5.00 \mu\text{g CN}^-/\text{mL} \div 26 \text{ g/mol} = 0.0100 \mu\text{mol}$ }, and sent to WHC with samples for a related job in Tank BY-111. The spikes, sample numbers S4028-028-X66, -X67, and -X68, were applied to the center of front sorbent sections, and levels of CN⁻ in the back sections were equal to blank levels, which indicated that no migration of CN⁻ occurred. The spiked blanks were analyzed and yielded an average blank-corrected percentage recovery of $69 \pm 10\%$ after a hold time of 11 weeks. It is probable that the poor spike recovery was related to the relatively long hold time. No samples were reanalyzed to

Table 2.3 Inorganic Sample Results Obtained from *In Situ* Samples Inserted into the Headspace of Tank BY-104 on 4/22/94

Sample	Analytical Results (μmol)			Sample Volume (L)	Vapor (a) Concentration (ppmv)
	Front Section	Back Section	Total(b) Blank-Corrected		
<u>NH₃ Samples:</u>			<u>44.2 (c)</u>	<u>3.75(c)</u>	<u>285 $\pm$ 4(c)</u>
S4062-162-X34	44.7	≤ 0.03	44.6	3.75	287
S4062-162-X35	45.9	NA(d)	45.8	3.94	281
S4062-162-X36	42.4	N/A	42.3	3.55	288
<u>NO₂ Samples:</u>			<u>≤ 0.0054</u>	<u>3.75</u>	<u>≤ 0.07</u>
S4062-162-X34	0.0141	0.0084	n/a	3.75	n/a
S4062-162-X35	0.0171	0.0090	n/a	3.94	n/a
S4062-162-X36	0.0179	0.0088	n/a	3.55	n/a
<u>NO Samples:</u>			<u>0.014</u>	<u>3.75</u>	<u>0.18 $\pm$ 0.05</u>
S4062-162-X34	0.0284	0.0075	0.0124	3.75	0.15
S4062-162-X35	0.0285	0.0089	0.0125	3.94	0.15
S4062-162-X36	0.0330	0.0077	0.0170	3.55	0.23
<u>HCN Samples:</u>			<u>≤ 0.0013</u>	<u>4.52</u>	<u>≤ 0.007</u>
S4062-162-X31	0.0120	0.0052	n/a	4.50	n/a
S4062-162-X32	0.0124	0.0047	n/a	5.45	n/a
S4062-162-X33	0.0131	0.0049	n/a	3.60	n/a
<u>Gravimetric Samples (mg, mg/L):</u>			<u>77.6 mg</u>	<u>4.52</u>	<u>17.4 $\pm$ 1.7 mg/L</u>
S4062-162-X31	n/a	n/a	76.6	4.50	17.0
S4062-162-X32	n/a	n/a	86.6	5.45	15.9
S4062-162-X33	n/a	n/a	69.5	3.60	19.3

- (a) Blank-corrected vapor concentrations were calculated using WHC-reported dry-air sample volumes (corrected to 21°C and 760 torr). In the calculation for concentration, the nitrite values (listed) were doubled to account for unanalyzed nitrate. Sample results were not corrected for percentage recovery of spiked samples or spiked blanks.
- (b) Total blank-corrected analyte masses were determined, when significant, by subtracting the quantity of analyte found in blanks from that found in samples. The levels of analytes found in blanks are described in Section 2.4.
- (c) Underlined values represent the average of the set of samples. Concentration uncertainty equals ± 1 standard deviation (absolute) for each set of samples. The use of " $\leq$ " is defined in Section 2.0.
- (d) NA = not analyzed. Only selected back sorbent sections were analyzed; n/a = not applicable.

check repeatability. No sample leachates were spiked after initial analysis with quantities of CN⁻ to test analytical percentage recoveries.

2.4.4 Gravimetric Results. The mass concentration of material collected in the sorbent-trap trains, believed to be primarily water vapor, was 17.4 ± 1.7 mg/L. The result was based on an average mass gain of 0.078 ± 0.009 g from three of three sets of HCN/H₂O sample trains. The NH₃/NO_x/H₂O

sample trains were not weighed. Blank corrections for the weighed sample trains were 0.0032 ± 0.0001 g, based on the results of three blank sorbent trains sent to WHC with samples for Tank BY-111. Although no spiked blanks were tested, the percentage recovery of mass from three blank H₂O traps spiked with 51 mg water was $103 \pm 2\%$ during a related sample job (Clauss et al. 1994).

3.0 Organic Analysis

3.1 SUMMA™ Canister Preparation

Before sending SUMMA™ canisters out to the field for sampling, the canisters are cleaned and verified contaminant free according to PNL Technical Procedure PNL-TVP-02^(a). The cleaning procedure uses an EnTech 3000 cleaning system that controls 1) filling the canisters with purified humid air and 2) evacuating, for several cycles with applied heat, before allowing the canister to evacuate overnight. The canister is filled a final time with purified humid air for analysis by PNL Technical Procedure PNL-TVP-01^(b), which is a modification of U.S. Environmental Protection Agency (EPA) Compendium Method TO-14. If the canister is verified as clean, free of TO-14 contaminants to a level of 5 parts per billion by volume (ppbv), the canister is evacuated to 30 in. Hg, tagged, and stored for use in the field. Before sending the canisters out to the field for sampling, the canisters are prehumidified with 100 µL of distilled water and labeled with a field-sampling identification. Canisters stored more than 30 but less than 60 days are re-evacuated and rehumidified before use. If stored more than 60 days, the canisters are recleaned and validated before use.

3.2 Sample Analysis Method

The SUMMA™ canister sample was analyzed according to PNL Technical Procedure PNL-TVP-03, *Determination of TO-14 Volatile Organic Compounds in Hanford Waste Tank Headspace Samples Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometry Analysis*, which is a modified version of EPA compendium Method TO-14. The method uses an EnTech cryoconcentration system interfaced with a Hewlett Packard (HP) 5971 GC/MS. The EnTech concentrator is used to pull a metered volume of sample air from the SUMMA™ canister, cryogenically concentrate the air volume, then transfer the volume to the GC/MS for analysis. A 100-mL volume of sample is measured and analyzed from the tank headspace. The organic components in the sampled air are separated on an analytical column, J&W Scientific DB-1 phase, 60-m by 0.32-mm internal diameter with 3-µm film thickness. The GC oven is programmed to run a temperature gradient beginning at 40°C, holding for 5 min, and ramping at 4°C per min to a final temperature of 260°C, with a 5-min hold.

3.3 Quality Assurance/Quality Control

Before the tank sample was analyzed, a diagnostic check was performed on the GC/MS instrument by running an instrument "quick tune," as described in PNL-TVP-03. Upon satisfactory completion of the instrument diagnostic check, a blank volume of purified nitrogen was analyzed to check the cleanliness of the system. The instrument was then calibrated over 6 data points ranging from 2 ppbv to 100 ppbv, using a standard gas mixture containing 40 volatile organic compounds listed in EPA compendium Method TO-14. A gas mixture containing bromochloromethane, 1,4-difluorobenzene, and chlorobenzene-d₅ was used as an internal standard (IS) for all blank, calibration

(a) Pacific Northwest Laboratory. 8/94. *Cleaning SUMMA™ Canisters and the Validation of the Cleaning Process*, PNL-TVP-02 (Rev. 0), PNL Technical Procedure, Richland, Washington.

(b) Pacific Northwest Laboratory. 8/94. *Determination of TO-14 Volatile Organic Compounds in Ambient Air Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometric Analysis*, PNL-TVP-01 (Rev. 0). PNL Technical Procedure, Richland, Washington.

standard, and sample analyses. Analyte response from sample components, ISs, and standards were obtained from the extracted ion plot from their selected mass ion. The calibration curve was generated by calculating the relative response ratios of the IS to calibration standard responses and plotting the ratios against the ratio of the calibration-standard concentration (in ppbv) to the IS concentration. A least-squares linear-regression routine was applied to the data set to generate the best-fit line for each compound. The equation for that line was then used to quantify the TO-14 compounds found in the tank samples.

3.3.1 Quantitation of TO-14 Results. The quantitative-analysis results for the TO-14 volatile organic compounds were calculated directly from the calibration curve generated using the IS method described above and in PNL-TVP-03. The conversion from ppmv to mg/m^3 assumes standard temperature and pressure (STP) conditions of 760 torr and 273K and was calculated directly from the following equation:

$$\text{mg}/\text{m}^3 = \frac{\text{ppmv} \times \text{g mol wt of compound}}{22.4 \text{ L/mole}} \quad (3.1)$$

3.3.2 Identification and Quantitation of Tentatively Identified Compounds. The tentatively identified compounds (TICs) are determined by mass-spectral interpretation and comparison of the spectra with the EPA/NIST/WILEY Library, which is a part of the HP 5971 instrument operating system. Chromatographic peaks with an area count greater than, or equal to, one half of the total area count of the chlorobenzene- d_5 IS peak at the 20-ppbv calibration level are tentatively identified and quantitatively estimated. This standard was chosen to determine the integration cutoff as it is in the middle of the chromatographic range and not in a region typically affected by coelution of other compounds. The quality of the mass-spectral searches was then reviewed by the principal investigators before the identification was assigned to each chromatographic peak.

The concentration of each TIC was estimated using a relative response factor calculated using a corrected total peak area for the IS chlorobenzene- d_5 . Specifically, the total integrated area for the chlorobenzene- d_5 peak had to be corrected for possible coeluting compounds before calculating the response factor. The corrected total peak area for the IS was calculated by multiplying the IS quantitation ion by a correction factor based on the ratio of the total integrated peak area to the quantitation ion as measured in blank runs. The corrected peak area was then used to calculate a response factor using the IS concentration in mg/m^3 :

$$\text{Response Factor} = \frac{\text{IS conc. (mg}/\text{m}^3)}{\text{IS peak area}} \quad (3.2)$$

The calculated response factor was then multiplied by the TIC peak area to give an estimated concentration for that compound. For butane, the total peak area was multiplied by the response factor for chlorobenzene- d_5 to give an estimated concentration of $1.25 \text{ mg}/\text{m}^3$ (for PNL 077). Internal standards bromochloromethane and difluorobenzene were not used to quantitate the TICs because coeluting compounds appeared to have greatly altered the signal of the quantitation ions for those two ISs.

The ppmv concentrations are calculated from mg/m^3 and the molecular weight of the analyte.

$$\text{TIC in ppmv} = \frac{\text{TIC (mg/m}^3\text{)} \times 22.4 \text{ L/mole}}{\text{TIC g mol wt}} \quad (3.3)$$

The IS level added to all blank, standard, and sample injections was 18.3 ppbv for bromochloromethane, 20.3 ppbv for 1,4-difluorobenzene, and 18.2 ppbv for chlorobenzene-d₅. The IS concentrations were converted from ppbv to mg/m³ at STP using a molecular weight of 129.39 (g/mol) for bromochloromethane, 114.09 for 1,4-difluorobenzene, and 117.6 for chlorobenzene-d₅.

3.4 Analysis Results

The results from the GC/MS analysis of the tank-headspace samples are presented in Tables 3.1 and 3.2. A representative total ion chromatogram showing the identity of major constituents is given in Figure 3.1.

Table 3.1 lists the quantitative results for compounds listed in Method TO-14. The levels of TO-14 analytes observed in the samples collected from Tank BY-104 were significantly low, close to the quantitation limit (2 ppb).

Table 3.2 lists the semi-quantitative results for the TICs. The predominant species observed in this sample were acetone, 2-methylpentane, hexane, propene, heptane, and normal paraffin hydrocarbons (NPH). The NPH, defined as n-alkanes from C₁₁ to C₁₅, were present in a significant amount in the sample. The total concentration of the TICs was 41.1 mg/m³. A significant amount (2.85 mg/m³) of acetone was observed in the sample. In addition, the amount of NPH and alkanes present in the TIC tables was approximately 26% of the total analytes identified.

Table 3.1 TO-14 Analysis Results for *In Situ* Sample S4062-162-077^(a) Collected from the Headspace of Tank BY-104 in SUMMATM Canisters on 4/22/94

TO-14 Analytes	CAS No.	PNL 077 ^(b)	
		(ppbv)	(mg/m ³) ^(c)
Dichlorodifluoromethane (FREON-12)	75-71-8	< 8.96	< 0.05
Methylchloride (Chloromethane)	74-87-3	< 2	< .004
1,2-Dichloro-1,1,2,2,-tetrafluoroethane (FREON-114)	76-14-2	< 2	< .02
Chloroethene (Vinyl chloride)	75-01-4	< 2	< .006
Methylbromide (Bromomethane)	74-83-9	< 2	< .009
Ethylchloride	75-00-3	< 2	< .006
Trichlorofluoromethane (FREON-11)	75-69-4	551.89	3.38
1,1-Dichloroethene			
(1,1-Dichloroethylene)	75-35-4	< 2	< .009
Dichloromethane (Methylene chloride)	75-09-2	< 2	< .01
1,1,2-Trichloro-1,2,2-trifluoroethane (FREON-113)	76-13-1	< 2	< .02
1,1-Dichloroethane	75-34-3	< 2	< .009
cis-1,2-Dichloroethene	156-60-5	< 2	< .009
Trichloromethane (Chloroform)	67-66-3	< 2	< .01
cis 1,2-Dichloroethane	107-06-2	< 2	< .009
1,1,1-Trichloroethane	71-55-6	< 2	< .01
Benzene	71-43-2	14.84	0.09
Carbon Tetrachloride	56-23-5	< 2	< .01
1,2-Dichloropropane	78-87-5	< 2	< .01
Trichloroethylene	79-01-6	< 2	< .01
cis 1,3-Dichloropropene	10061-01-5	< 2	< .01
trans 1,3-Dichloropropene	10061-02-6	< 2	< .01
1,1,2-Trichloroethane	79-00-5	< 2	< .01
Methylbenzene (Toluene)	108-88-3	170.04	0.70
1,2-Dibromoethane	106-93-4	< 2	< .02
Tetrachloroethene	127-18-4	< 2	< .01
Chlorobenzene	108-90-7	< 2	< .02
Ethylbenzene	100-41-4	2.75	0.01
m-Xylene (1,3-Dimethylbenzene)	108-38-3	3.21	0.02
p-Xylene (1,4-Dimethylbenzene)	106-42-3		
Styrene	100-42-5	< 2	< .009
1,1,2,2-Tetrachloroethane	79-34-5	< 2	< .02
o-Xylene (1,2-Dimethylbenzene)	95-47-6	< 2	< .01
1,3,5-Trimethylbenzene	108-67-8	< 2	< .01
1,2,4-Trimethylbenzene	95-63-6	< 2	< .01
Chloromethylbenzene	100-44-7	< 2	< .01
m-Dichlorobenzene	541-73-1	< 2	< .01
p-Dichlorobenzene	106-46-7	< 2	< .01
o-Dichlorobenzene	95-50-1	< 2	< .01
1,2,4-Trichlorobenzene	120-82-1	< 2	< .02
Hexachloro-1,3-butadiene	87-68-3	< 2	< .02

(a) WHC ID number.

(b) PNL SUMMATM canister number.

(c) Calculated from ppbv using molecular weight of compound, 760 mm torr, and 0°C.

Table 3.2 Tentatively Identified Compounds and Estimated Concentrations in the Headspace of Tank BY-104 *In Situ* SUMMA™ Canister Sample S4062-162-077^(a) Collected on 4/22/94

Tentatively Identified Compounds ^(c)	Mol. Wt.	CAS No. ^(c)	Retention Time (min)	PNL 077 ^(b)	
				Estimated Concentration ^(d) mg/m ³	ppmv
Carbon dioxide	44	115-91-2	6.06	(e)	(e)
Propene ^(f)	42	115-90-5	6.57	1.68	0.90
n-Propane (coeluent)	44	74-98-6		(f)	(f)
Cyclopropane	42	75-19-4	7.25	0.14	0.07
2-Methylpropane	58	75-28-5	7.50	0.43	0.17
2-Butene	56	107-01-7	7.97	0.93	0.37
n-Butane	58	106-97-8	8.17	1.25	0.48
C4 Alkane ^(g)	58	(h)	8.70	0.23	0.09
C4 Alkene ^(g)	56	(h)	9.68	0.18	0.07
Acetone	58	67-64-1	10.24	2.85	1.10
Trichlorofluoromethane	136	75-69-4	10.56	1.42	0.23
2-Pentene	70	109-68-2	10.77	0.58	0.19
C5 Alkene ^(g)	70	(h)	11.01	0.07	0.02
Pentane	70	109-66-0	11.20	1.54	0.49
Unknown	(h)	(h)	11.82	(h)	(h)
1-Pentane	70	109-67-1	11.98	0.07	0.02
1-Propanol	60	71-23-8	13.17	0.06	0.02
4-Methyl-1-pentene	84	691-37-2	13.62	0.36	0.10
2-Methylpentane	86	107-83-5	14.23	2.08	0.54
Butanal	72	123-72-8	14.43	0.15	0.05
2-Butanone	72	78-93-3	14.60	0.70	0.22
3-Methylpentane	86	96-14-0	14.95	0.43	0.11
1-Hexene	84	592-41-6	15.20	0.40	0.11
Bromochloromethane (IS)		74-97-5	15.76		
Hexane (coeluent)	86	110-54-3		1.67	0.44
2-Hexene	84	592-41-6	16.48	0.05	0.01
Tetrahydrofuran	84	109-99-9	16.68	0.49	0.13
Methylcyclopentane	86	96-37-7	17.37	0.25	0.07
3-Methyl-2-butanone	86	563-80-4	17.94	0.07	0.02
1-Butanol	74	713-66-3	18.30	0.53	0.16
Benzene	78	71-43-2	18.71	0.07	0.02
1,4-Difluorobenzene (IS)		540-36-3	19.28		
2-Pentanone (coeluent)	86	107-87-9		0.80	0.21
3-Methylhexane	100	589-34-4	19.82	0.58	0.13
1-Heptene	98	592-76-7	20.45	0.22	0.05
n-Heptane	100	142-82-5	21.06	0.99	0.22
Unknown	(h)	(h)	22.26	0.07	(h)
Methylcyclohexane	100	10120-28-2	22.63	0.19	0.04
Toluene	92	108-88-3	24.41	0.53	0.13
2-Methylheptane	114	592-27-8	24.62	0.23	0.05
3-Methylheptane	114	(h)	25.05	0.06	0.01
C8 Alkane ^(g)	114	(h)	25.78	0.06	0.01
n-Octane	114	111-65-9	26.37	0.38	0.07
Siloxane (impurity)	(h)	(h)	27.08	0.12	(h)
Alkane	(h)	(h)	27.96	0.14	(h)
Chlorobenzene-d5 (IS)		3114-55-4	28.44		
C3 Cyclohexane ^(g)	128	(h)	28.87	0.09	0.02
3-Heptanone	114	(h)	29.81	0.21	0.04
2-Heptanone	114	(h)	29.93	0.19	0.04

Table 3.2 Contd

Tentatively Identified Compounds ^(c)	Mol. Wt.	CAS No.	Retention Time (min)	PNL 077 ^(b)	
				Estimated Concentration ^(d) mg/m ³	ppmv
C9 Alkane ^(g)	128	(h)	30.10	0.07	0.01
Unknown	(h)	(h)	30.51	0.07	(h)
2-Butoxyethanol	118	111-76-2	30.95	0.32	0.06
n-Nonane	128	111-84-2	31.38	0.23	0.04
6-Methyl-2-heptanone	128	(h)	33.14	0.36	0.06
Phenol	94	108-95-2	33.80	0.08	0.02
2-Octanone	128	111-13-7	34.80	0.07	0.01
Siloxane (impurity)	(h)	(h)	35.39	0.05	(h)
n-Decane	142	124-18-5	36.06	0.21	0.03
C11 Alkane ^(g)	156	(h)	37.18	0.14	0.02
Unknown	(h)	(h)	38.07	0.12	(h)
Unknown	(h)	(h)	38.63	0.05	(h)
C11 Alkane ^(g)	156	(h)	39.90	0.07	0.01
n-Undecane	156	1120-21-4	40.37	0.45	0.06
Unknown	(h)	(h)	40.95	0.10	(h)
C12 Alkane ^(g)	170	(h)	41.21	0.08	0.01
Alkane ^(g)	(h)	(h)	41.65	0.12	(h)
Unknown	(h)	(h)	41.89	0.07	(h)
Methyldecahydronaphthalene ^(g)	170	(h)	42.09	0.12	0.02
C5 Cyclohexane ^(g)	156	(h)	42.44	0.16	0.02
Methyldecahydronaphthalene ^(g)	170	(h)	42.65	0.18	0.02
Alkane	(h)	(h)	42.85	0.21	(h)
Alkane	(h)	(h)	43.02	0.20	(h)
Alkane	(h)	(h)	43.33	0.13	(h)
Alkene	(h)	(h)	44.16	0.11	(h)
n-Dodecane	170	(h)	44.39	2.08	0.27
2,6-Dimethylundecane	184	(h)	45.03	1.50	0.18
C13 Alkene ^(g)	182	(h)	46.45	0.83	0.10
C2 Decahydronaphthalene ^(f,g)	184	(h)	46.70	0.22	(f)
Alkane ^(e) (coeluent)	(h)	(h)		(f)	(f)
Alkane	(h)	(h)	46.86	0.33	(h)
Alkane	(h)	(h)	47.16	0.19	(h)
Alkane	(h)	(h)	47.28	1.82	(h)
n-Tridecane	184	629-50-5	48.14	2.63	0.32
C14 Alkene ^(g)	196	(h)	48.60	0.37	0.04
Alkane	(h)	(h)	48.91	0.37	(h)
Alkene	(h)	(h)	49.48	0.08	(h)
Alkane	(h)	(h)	50.72	0.12	(h)
Alkane	(h)	(h)	51.01	1.37	(h)
n-Tetradecane	198	629-59-4	51.64	1.18	0.13
C8 Cyclohexane ^(g)	198	(h)	53.63	0.06	0.01
Alkane	(h)	(h)	53.84	0.48	(h)
n-Pentadecane	212	(h)	54.93	0.13	0.01

(a) WHC sample number.

(b) PNL canister identification number.

(c) Obtained by mass spectral interpretation and comparison with the EPA/NIST and WILEY Libraries.

(d) Semi-quantitative estimate calculated using concentration of the chlorobenzene-d₅ IS.

(e) Concentration information for carbon dioxide is not possible by this analytical method.

(f) Percent contribution from each coeluting peak cannot be determined.

(g) Other structural isomers should be considered.

(h) Information for this TIC cannot be determined.

4.0 Conclusions

The concentrations of selected inorganic and organic compounds were determined from *in situ* samples of the headspace of Tank BY-104 on 4/28/94. The ammonia concentration was found to be 285 ± 4 ppmv. The concentration of NO_2 was ≤ 0.07 ppmv. The concentration of NO was ≤ 0.18 ppmv. The concentration of HCN was ≤ 0.007 ppmv. The mass concentration was 17.4 ± 1.7 mg/mL and was expected to consist largely of water vapor. Minor differences between the individual results of the three NH_3 samples matched closely the differences in the quantities of dry air sampled during each sample.

Acetone was found to make up about 18% of the total concentration of all the organic compounds identified. Samples from Tank BY-104 had a significant amount of NPH and decane present (25% of the total concentration of all the identified analytes). The estimated concentration of all the analytes identified in the ISS sample of Tank BY-104 was found to be below the 1-ppmv level.

5.0 References

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6.0 Further Reading

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Figure 3.1 GC/MS Chromatogram of Hanford Waste Tank BY-104 *In Situ* SUMMA™ Canister Sample S4062-162-077 Collected on 4/22/94

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