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# Initial characterization of processes of soil carbon stabilization using forest stand-level radiocarbon enrichment

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1 **Initial characterization of processes of soil carbon stabilization using forest stand-level**  
2 **radiocarbon enrichment**

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**1 Abstract**

2 Although the rates and mechanisms of soil organic matter (SOM) stabilization are difficult to  
3 observe directly, radiocarbon has proven an effective tracer of soil C dynamics, particularly  
4 when coupled with practical fractionation schemes. To explore the rates of C cycling in  
5 temperate forest soils, we took advantage of a unique opportunity in the form of an inadvertent  
6 stand-level  $^{14}\text{C}$ -labeling originating from a local industrial release. A simple density  
7 fractionation scheme separated SOM into inter-aggregate particulate organic matter (free light  
8 fraction, free LF), particulate organic matter occluded within aggregates (occluded LF), and  
9 organic matter that is complexed with minerals to form a dense fraction (dense fraction, DF).  
10 Minimal agitation and density separation was used to isolate the free LF. The remaining dense  
11 sediment was subjected to physical disruption and sonication followed by density separation to  
12 separate it into occluded LF and DF. The occluded LF had higher C concentrations and C:N  
13 ratios than the free LF, and the C concentration in both light fractions was ten times that of the  
14 DF. As a result, the light fractions together accounted for less than 4% of the soil by weight, but  
15 contained 40% of the soil C in the 0-15 cm soil increment. Likewise, the light fractions were  
16 less than 1% weight of the 15-30 cm increment, but contained more than 35% of the soil C. The  
17 degree of SOM protection in the fractions, as indicated by  $\Delta^{14}\text{C}$ , was different. In all cases the  
18 free LF had the shortest mean residence times. A significant depth by fraction interaction for  $^{14}\text{C}$   
19 indicates that the relative importance of aggregation versus organo-mineral interactions for  
20 overall C stabilization changes with depth. The rapid incorporation of  $^{14}\text{C}$  label into the  
21 otherwise depleted DF shows that this organo-mineral fraction comprises highly stable material  
22 as well as more recent inputs.

23

## 1 **1. Introduction**

2 Carbon stabilization in forest soil organic matter (SOM) occurs concurrently with  
3 destabilization (Sollins et al., 1996), and large changes in the resident C in SOM may appear as  
4 only small net changes in bulk C values. Soil organic matter is protected by a variety of  
5 mechanisms, both physical and chemical, which results in SOM pools with different residence  
6 times (Oades, 1988; Torn et al., 1997). Although it is difficult to directly observe the processes  
7 of stabilization, carbon isotopes can give insight into rates of C cycling. Radiocarbon has  
8 proven an effective tracer of soil C at annual to decadal timescales because of the global spike in  
9 atmospheric  $^{14}\text{CO}_2$  caused by thermonuclear weapons testing during the early 1960's (Goh et al.,  
10 1977; Trumbore, 1993; Trumbore and Zheng, 1996). However, the sensitivity of the bomb  
11 spike as a tracer is lower each year as the annual rate of atmospheric change declines (Levin and  
12 Hesshaimer, 2000). In addition, its application to some questions is limited because the same  
13 enrichment level applies to all ecosystems, making it hard to find an experimental control.

14 Recently, an unplanned  $^{14}\text{C}$  release created a pulse label that can be used to study a range of  
15 questions in C cycling in a temperate forest region of the United States. In the summer of 1999,  
16 a large pulse of  $^{14}\text{CO}_2$  was released near Oak Ridge, Tennessee, presumably from a local  
17 incinerator (Trumbore et al., 2002). The photosynthetic uptake of the  $^{14}\text{CO}_2$  created a pulse label  
18 of  $^{14}\text{C}$  in plant biomass of the local forest. There was a gradient in enrichment, with sites near  
19 the source experiencing about three times the level of the bomb spike (enriched sites), and those  
20 further away receiving much less (near-background sites). This stand-level isotopic label  
21 presents unique opportunities for studying different processes in belowground carbon cycling in  
22 some detail.

1 Fractionating soils allows isotopic and elemental analysis to be applied to specific portions of  
2 organic matter, increasing both the sensitivity to detect change and the ability to ascribe results to  
3 a particular process or category of the organic matter (Trumbore, 1993; Hungate et al., 1996;  
4 Trumbore and Zheng, 1996). A challenge in SOM fractionation however is to develop  
5 procedures that produce fractions that represent functionally different soil C pools while being  
6 simple enough to link to ecosystem models. In the basic procedure, the material that floats to the  
7 surface of a dense liquid is called the light fraction, and the material that sinks to the bottom is  
8 called the heavy or dense fraction (Greenland and Ford, 1964; Sollins et al., 1999). The density  
9 of the liquid is initially adjusted to maximize the recovery of particulate organic matter with high  
10 C concentration in the light fraction. In comparison to the dense fraction, the light fraction is  
11 characterized as chemically and visually more plant- or litter-like (Spycher et al., 1983; Golchin  
12 et al., 1994a; Amalfitano et al., 1995; Gregorich et al., 1996), and has been shown to be more  
13 sensitive to management (Dalal and Mayer, 1986; Compton and Boone, 2000). Organic C in the  
14 dense fraction (DF) is more closely associated with minerals and typically more stable *in situ*  
15 (Trumbore and Zheng, 1996; Gaudinski et al., 2000). Golchin et al. (1994a) adapted the method  
16 to better represent the complexities of soil structure by separating a 'free' light fraction (free LF)  
17 from between aggregates and an 'occluded' light fraction (occluded LF) from within aggregates.  
18 Carbon-13 NMR analyses of the free LF and occluded LF indicated that the occluded LF was  
19 somewhat more humified and presumably older. Baisden et al. (2002b) used  $^{14}\text{C}$  to estimate the  
20 residence times of the free LF and three mineral-associated light fractions (Golchin et al., 1994b)  
21 from grassland soils. They concluded that there was no appreciable difference among the three  
22 mineral-associated fractions, but found that the free light fraction had a shorter residence time  
23 than the occluded or mineral-associated carbon.

1 Our objectives were to 1) use the gradient of  $^{14}\text{C}$  enrichment in the lands surrounding Oak  
2 Ridge National Laboratory, known as the Oak Ridge Reservation (ORR) to assess the  
3 effectiveness of the three-pool density fractionation procedure; and 2) to investigate C cycling  
4 and stabilization in those soils. Sites were selected with two levels of labeling: near-background  
5 sites and enriched sites. We considered the soils at the near-background site to be indicative of  
6 the system prior to the 1999  $^{14}\text{C}$  pulse. We used the results from the enriched site to gauge the  
7 progression of the recent  $^{14}\text{C}$  pulse through soil organic matter. We judged a successful method  
8 to be one that would adequately discriminate meaningful C pools in the absence of a pulse label,  
9 as well as show sensitivity to changes in those pools as elevated levels of  $^{14}\text{C}$  moved through the  
10 system.

11

## 12 **2. Materials and Methods**

### 13 *2.1. Site characteristics*

14 The Enriched Background Isotope Study (EBIS) was established in the autumn of 2000 on  
15 the Oak Ridge Reservation (ORR), in the U.S. Department of Energy's National Environmental  
16 Research Park near Oak Ridge, Tennessee (Trumbore et al., 2002). The mean annual  
17 precipitation at ORR is 1358 mm and mean annual temperature is  $14^\circ\text{C}$  (Johnson and Van Hook  
18 1989). All EBIS research plots are located on upper slope, ridge positions in the upland oak  
19 forest type (*Quercus* spp.; *Acer* spp.; *Carya* spp.) with scattered pine (*Pinus echinata* and *P.*  
20 *virginiana*) and mesophytic hardwoods (*Liriodendron tulipifera*, *Fagus grandifolia*). The ages  
21 of the over-story trees cover a broad range from about 40 to 150 years, and the maximum canopy  
22 height is approximately 26 meters. Leaf area index is typically about  $6\text{ m}^2\text{ m}^{-2}$ .

1       The whole-system background  $^{14}\text{C}$ -enrichment, described previously by Trumbore et al.  
2 (2002), was not experimentally applied, but rather originated from an industrial source, such as  
3 hazardous waste incinerators, in the vicinity of ORR. There appear to have been numerous small  
4 releases beginning in 1995, punctuated by a large release between June and August of 1999 (Fig.  
5 1). According to the record of  $^{14}\text{C}$  in tree-ring cellulose, the releases prior to 1999 affected only  
6 the west end of the ORR (TVA site). The large 1999 release raised atmospheric levels at both  
7 sites but had a larger effect on the west end than the east (Walker Branch site). Labeled  $^{14}\text{CO}_2$   
8 was fixed by vegetation and incorporated into roots, leaves, and non-structural storage pools  
9 (Gaudinski, 2001; Trumbore et al., 2002). Measurements in the summer of 2000 revealed a  
10 gradient of  $^{14}\text{C}$ -enrichment, highest in the western ORR and dropping to near-background in the  
11 eastern ORR.

12       Permanent plots were established during Winter 2000 in the western and eastern ORR. Eight  
13 7×7 m plots were located at each of four sites. The sites were located on Inceptisols derived from  
14 shale or Ultisols derived from dolomitic parent materials. In the present work we only consider  
15 the Ultisols. These are deep, highly weathered soils with a 10-15-cm thick A horizon above a  
16 well-developed EB horizon and multiple Bt subsoil horizons. The clay fraction is dominated by  
17 kaolinite with lesser quantities of hydroxy-Al interlayered vermiculite.

18       In the present study, we analyze samples from 0-15 and 15-30-cm soils at an eastern  
19 (Walker Branch) and western site (TVA) of the ORR. Eight samples were taken in March 2001  
20 from each depth increment at each site, for a total of 32 bulk soil samples. Soil was collected  
21 with a 28-cm and a 10-cm diameter core sampler, respectively, and stored at  $-20^\circ\text{C}$  until further  
22 processing. Upon thawing, soils were sieved to  $<2\text{mm}$ , sorted by hand to remove roots, and  
23 finally oven-dried at  $105^\circ\text{C}$ .



1 Subsequent to collection of these samples, a litter manipulation study was initiated by  
2 applying highly enriched ( $971 \pm 12\%$ ) or near-background ( $222 \pm 1\%$ ) litter, both collected in  
3 summer 2000, to the plots in a random design. In future studies, the litter treatments can be used  
4 to investigate the influence of litter versus root input of carbon (and  $^{14}\text{C}$ ) into soil organic carbon  
5 pools.

## 6 *2.2. Density separation*

7 Three density fractions were separated from bulk soil following the method of Golchin et al.  
8 (1994a): free light fraction, occluded light fraction, and dense fraction. About 100 mL of  
9 sodium polytungstate (NaPT,  $\text{Na}_6[\text{H}_2\text{W}_{12}\text{O}_{40}]$ , Sometu-US, Van Nuys, California) of  $1.70 \text{ g mL}^{-1}$   
10 was added to 20 g oven-dried soil in a centrifuge bottle, and the bottle was inverted gently until  
11 the soil was wetted. After sitting for 45 min, the mixture was centrifuged in a bucket rotor for 45  
12 min at 3600 rpm. Floating materials (free LF) were aspirated from the centrifuge bottle, then  
13 rinsed with double de-ionized  $\text{H}_2\text{O}$  (di  $\text{H}_2\text{O}$ ) on a  $0.8 \mu\text{m}$  polycarbonate filter (Whatman  
14 Nucleopore Track Etch Membrane). Sediment and NaPT remaining in the centrifuge bottle were  
15 mixed for 1 min using a benchtop mixer (G3U05R, Lightnin, New York, NY) at 1700 rpm. The  
16 blade was rinsed over the bottle, and the mixture sonicated in an ice bath for 3 min at 70% pulse  
17 for a total input of  $200 \text{ J mL}^{-1}$  (Branson 450 Sonifier, Danbury, CT). Debris were rinsed with  
18 NaPT from the sonicator probe into the bottle, and the mixture was centrifuged as before for 45  
19 min. Floating materials (occluded LF) were very fine, and prone to clouding the supernatant  
20 with even small perturbations. If the supernatant was clouded, the bottle was allowed to sit up to  
21 12 h before the occluded LF was aspirated. The separated occluded LF was also rinsed with di  
22  $\text{H}_2\text{O}$  on a  $0.8 \mu\text{m}$  polycarbonate filter. The sediment remaining in the centrifuge bottle (dense  
23 fraction, DF) was rinsed by repeatedly aspirating the supernatant, adding di  $\text{H}_2\text{O}$ , shaking, and

1 centrifuging at 7500 rpm for 25 min. Fractions were dried for 24 h at 105°C. Sample splits of  
2 the bulk soil were re-dried at 105°C to correct for moisture accumulation during storage. We  
3 present all values on an oven-dry basis. Weights were measured and tracked to the nearest  
4 0.0001 g.

### 5 *2.3. Elemental analysis*

6 Samples were analyzed for total C, total N, and fraction modern  $^{14}\text{C}$ . The total C and N  
7 values for the density fractions were obtained from a Carlo Erba 2100 C/N elemental analyzer.  
8 Total C and N values for the bulk soil were obtained from a LECO CN-2000 elemental analyzer.  
9 Both were calibrated against standards traceable to the National Institute of Standards and  
10 Technology (Gaithersburg, MD). Radiocarbon values were measured on the Van de Graaff FN  
11 accelerator mass spectrometer (AMS) at the Center for Accelerator Mass Spectrometry,  
12 Lawrence Livermore National Laboratory. In preparation for AMS analysis, samples were  
13 combusted in evacuated, sealed tubes in the presence of CuO and Ag, then reduced onto iron  
14 powder in the presence of  $\text{H}_2$  (Vogel et al., 1984). Splits of combusted sample were taken for  $^{13}\text{C}$   
15 analysis from each type of fraction from both sites and depths for correction of the AMS values,  
16 and reporting in  $\Delta^{14}\text{C}$  notation (Stuiver and Polach, 1977).

### 17 *2.4. Data analysis*

18 The Walker Branch and TVA sites were considered separately, each as a split plot design.  
19 Soil increment was the main plot, and density fraction the subplot. Analyses of variance  
20 (ANOVA's) were carried out in SAS (SAS Institute Inc., Version 8.2) using the Mixed  
21 procedure. Residuals were examined for normality and constant variance, and if necessary  
22 transformed using natural logarithm or square-root transformations. Specifically, for Walker  
23 Branch,  $^{14}\text{C}$ , C:N ratios, C and N concentrations, and soil C distribution in the density fractions

1 were transformed with natural logarithm prior to accepting the ANOVA results. Likewise, for  
2 TVA, C:N ratios and C concentrations were transformed with square root, and N concentrations  
3 were transformed with natural logarithm. If ANOVA results were significant, pre-planned  
4 comparisons were made using least-squares differences within the Mixed model. All references  
5 to statistical differences between treatments are based upon a significance level of 0.05, although  
6 p-values between 0.05 and 0.1 are presented and discussed.

### 7 **3. Results**

#### 8 *3.1. Walker Branch (Near-background site)*

9 Specific mass recoveries, C and N concentrations, and soil C distribution are located in Table  
10 1. Carbon was at least ten times more concentrated in the light fractions than in the dense  
11 fraction. As a result, the light fractions contained a disproportionate amount of the total soil C.  
12 In the upper soil, the free LF was only 2% of the soil mass, but contained 25% of the soil C.  
13 Likewise, the occluded LF was only 1% of the soil mass, but contained over 16% of the soil C.  
14 In contrast, the DF made up nearly 97% of the soil by mass, but contained only 50% of the soil  
15 C. The distribution of soil C in the fractions did not change significantly with depth (Table 2),  
16 although the relative mass of the light fractions four times greater in the upper soil. The  
17 remaining soil C was not recovered, and was probably lost as dissolved organic C (NaPT- and  
18 water-extractable) during rinsing of the fractions. Comparable C loss from some soils was also  
19 reported by Golchin et al. (Golchin et al., 1994a; Golchin et al., 1994b).

20 The C concentrations of the fractions changed differently with depth, resulting in a statistical  
21 interaction (Table 2). The C concentration of the DF decreased by nearly five times with depth,  
22 the free LF increased slightly, and the occluded LF showed no change. Thus, although the  
23 occluded LF had higher C concentrations than the free LF in the upper soil ( $t_{25}=4.48$ ,  $P=0.0001$ ),

1 the evidence for a difference was weaker in the lower soil increment ( $t_{25}=1.94$ ,  $P=0.06$ ). The N  
2 concentrations of the fractions also changed differently with depth, resulting in a statistical  
3 interaction (Table 2). There was no statistical evidence of a difference between the N content of  
4 the free and occluded light fractions, and the LF N concentrations were at least ten times higher  
5 than those of the DF. The N by depth interaction appeared to result from a steeper decrease in N  
6 concentration in the DF with depth than in the light fractions.

7 The C:N ratios of the light fractions and the DF showed opposite trends with increasing  
8 depth, resulting in a significant interaction (Table 2). The C:N of the DF decreased with depth  
9 whereas the C:N ratios of both light fractions increased with depth.

10 The  $\Delta^{14}\text{C}$  of the density fractions interacted significantly with soil increment (Table 2), in  
11 that the  $\Delta^{14}\text{C}$  of the protected-C fractions (occluded LF and DF) decreased more with depth than  
12 the free LF (Figure 1). Within this trend, the free LF had greater  $\Delta^{14}\text{C}$  than the occluded LF,  
13 which in turn was higher than the DF.

### 14 3.2. TVA (*Enriched site*)

15 Specific mass recoveries, C and N concentrations, and soil C distribution, are located in  
16 Table 3. The C and N values at TVA followed the same patterns as those at Walker Branch,  
17 including disproportionately high soil C in the light fractions and the highest concentration of C  
18 and widest C:N ratio in the occluded LF. However, there were some exceptions. Specifically,  
19 there was a significant interaction between the distribution of soil C in the fractions and soil  
20 increment (Table 4), which reflected lower C content in the 15-30-cm free LF as compared to the  
21 surface soil, and weak evidence for a higher C content in the occluded LF ( $t_{26}=1.72$ ,  $P=0.09$ ) and  
22 DF ( $t_{26}=1.83$ ,  $P=0.08$ ) at depth. The free LF contained higher N concentrations than the

1 occluded LF in both depths, although the DF had a much lower N concentration that decreased  
2 more with depth than either light fraction.

3 The  $\Delta^{14}\text{C}$  of the density fractions interacted significantly with soil increment (Table 4), but  
4 apparently not in the same manner as the  $\Delta^{14}\text{C}$  in the fractions from Walker Branch (Figures 2  
5 and 3). As with the Walker Branch fractions, the free LF exhibited the highest  $^{14}\text{C}$  activity of the  
6 three fractions. However, at 0-15 cm the  $\Delta^{14}\text{C}$  in the DF was higher than in the occluded LF,  
7 and not different in the 15-30-cm soil increment.

8

#### 9 **4. Discussion**

10 Our results show that the three-pool density fractionation procedure (Golchin et al., 1994a)  
11 separates SOM in these soils into three fractions with different physical and chemical  
12 characteristics, activity, and, presumably, C stabilization mechanisms. These fractions are the  
13 particulate organic matter (POM) found outside of aggregates (free LF), POM found within  
14 aggregates (occluded LF), and a mineral-associated carbon fraction (DF). One quarter of the  
15 soil C in each depth was free LF, and about 65% was stabilized by occlusion or organo-mineral  
16 interactions. There are several lines of evidence that there are different processes of  
17 stabilization, or cycling, among these fractions, including %C, C:N,  $^{14}\text{C}$ , and links to  $^{13}\text{C}$  NMR  
18 from other studies.

19 At the most basic level, the C concentrations and C:N ratios of the fractions indicate that they  
20 are distinctly different. The DF was 1% C, whereas the occluded and free LF were 40 and 25%,  
21 respectively. These values are comparable to those obtained by Golchin et al., (1994a) and are  
22 typical of the disproportionate soil C in density fractions (Dalal and Mayer, 1986; Christensen,  
23 1992; Cromack et al., 1999; Parker et al., 2002). Analysis of the light fractions from Walker

1 Branch and TVA soils support the characterization of the occluded LF as a more protected, less  
2 active fraction than the free LF (Golchin et al., 1994a; Baldock et al., 1997). The observation  
3 that C concentration and C:N ratios were higher in the occluded LF than the free LF is consistent  
4 with the findings of Golchin et al. (1994a), who found that the occluded LF contained higher C  
5 concentrations and comparable or higher C:N ratios than the free LF from several forest and  
6 grassland soils in Australia. Parker et al. (2002) also reported higher C concentration and C:N  
7 ratios in the occluded LF from several forest soils in New England. In extensive  $^{13}\text{C}$  NMR  
8 characterizations, the occluded LF contained lower proportions of O-alkyl-C, higher proportions  
9 of alkyl-C, and higher aromaticity (Golchin et al., 1994a; Sohi et al., 2001) compared to free LF,  
10 which indicates that the occluded fraction is more degraded and recalcitrant (Baldock et al.,  
11 1992; Baldock et al., 1997) or contains black C (Baisden et al. 2002b).

12 Trends in  $\Delta^{14}\text{C}$  between fractions and depths support the basic conclusions from the C  
13 concentrations and C:N ratios. We can use the  $^{14}\text{C}$  signatures to compare relative residence  
14 times among the fractions and between depths. Translating the  $\Delta^{14}\text{C}$  to precise turnover times is  
15 problematic because of the combination of bomb spike and local pulse. The two clearest trends  
16 were the consistently high  $\Delta^{14}\text{C}$  of the free LF, and the consistent decrease in  $\Delta^{14}\text{C}$  with depth  
17 (Figures 2 and 3). The high  $\Delta^{14}\text{C}$  of the free LF's confirms that the free LF is the active fraction,  
18 most responsive to changes in C input (Dalal and Mayer, 1986; Janzen et al., 1992; Alvarez and  
19 Alvarez, 2000; Compton and Boone, 2000). In the Walker Branch fractions,  $\Delta^{14}\text{C}$  decreased  
20 across fractions with increasing mineral association. The decrease in SOM  $\Delta^{14}\text{C}$  with depth has  
21 been reported by numerous studies (Trumbore et al., 1989; Torn et al., 1997; Gaudinski et al.,  
22 2000; Paul et al., 2001; Baisden et al., 2002a; Kaiser et al., 2002; Rumpel et al., 2002; Certini et  
23 al., 2003), and is typically associated with C stabilization. Kaiser et al. (2002) measured

1 increasing proportions of HF-acid-soluble organic C, in addition to decreasing  $\Delta^{14}\text{C}$  with depth,  
2 and concluded that SOM at greater depths has increased mineral association and protection.  
3 Rumpel et al. (2002) found that soil C age and alkyl-C increased with depth, which they  
4 attributed to chemical and physical stabilization. We found that the difference in  $\Delta^{14}\text{C}$  between  
5 the unprotected (free LF) and protected (occluded LF and DF) fractions increased with depth,  
6 indicating increased efficiency of mineral protection with depth.

7 The fractions from TVA require a more detailed interpretation, as the ecosystem had  
8 prolonged exposure to elevated atmospheric  $^{14}\text{C}$  (Figures 1 and 3). In the 10 y prior to the large  
9  $^{14}\text{C}$  pulse in 1999, ambient  $^{14}\text{C}$  levels decreased at Walker Branch, from about 150 ‰ in 1989 to  
10 about 100 ‰ in 1999 (in tree ring cellulose). In contrast, however, tree core  $^{14}\text{C}$  was higher each  
11 year at TVA after 1995, and in 1999 was nearly 400 ‰. The elevated  $^{14}\text{C}$  was incorporated into  
12 the free LF at TVA, but was also apparent in the DF. The response of the free LF was expected,  
13 because it quickly responds to C inputs, and the photosynthetic products at TVA were elevated in  
14  $^{14}\text{C}$  for several years. However, the large incorporation of the label into the DF at TVA does not  
15 fit the conceptual model of the DF as a stable, passive C pool. The values of the DF at TVA  
16 sharply contrast those at the near-background Walker Branch, which had a depleted  $^{14}\text{C}$  signature  
17 relative to the free and occluded LF (indicating slower turnover). Rapid response by the DF to  
18 inputs has been observed in other studies, though. For example, increases in litter inputs resulted  
19 in increases in the amount of DF in a temperate forest soil (Boone, 1994). Using acid hydrolysis,  
20 Trumbore et al. (1989) and Trumbore and Zheng (1996) separated the DF into hydrolysable  
21 (rapid) and non-hydrolysable (passive) pools that had different  $^{14}\text{C}$  values.

22 The DF in these soils appears to contain at least two different C pools: an older, more stable  
23 pool of C, and a more recent, fast-cycling C pool. To the extent that the DF acts as a sink for

1 microbial byproducts and a medium for microbial biomass (Chotte et al., 1998), it must include a  
2 C component that can cycle rapidly (Trumbore et al., 1989; Boone, 1994; Golchin et al., 1996;  
3 Trumbore and Zheng, 1996). Swanston et al. (2002) incubated the LF and DF from forest soils  
4 separately, and found no difference in the rate of respiration for the first 120 d. Therefore there  
5 must be some labile material associated with the dense fraction. However, different patterns of  
6 net N mineralization, indicated that the chemistry of the labile C was not the same in the two  
7 fractions (Swanston et al., 2004). It seems likely that the  $^{14}\text{C}$  of the DF at TVA was enriched by  
8 recent plant inputs and associated microbial byproducts. This fast-cycling pool must be closely  
9 associated with the DF, considering that it withstood mixing and sonication in NaPT and  
10 repeated rinsing in water.

11 Golchin et al. (1994b), based on adding two intermediate densities to their earlier method  
12 (Golchin et al., 1994a), proposed a conceptual model that could explain the incorporation of  
13 recent SOM into the DF. Stable aggregates form as free LF is colonized by microbiota and  
14 encrusted with mineral particles, which are stabilized by microbial exudates. High-quality  
15 organics such as proteins and carbohydrates are degraded and converted to microbial biomass  
16 and exudates, selectively preserving more recalcitrant alkyl and lignaceous compounds in the  
17 organic core of the aggregate. As the organic substrate becomes increasingly recalcitrant, the  
18 microbial community declines and the aggregate becomes unstable. In the later stages of the  
19 cohesive aggregate, the conservation of recalcitrant molecules and decrease of microbial biomass  
20 and byproducts results in an increase in the C concentration and C:N ratio of the remaining  
21 organic core. At these stages, the biomass and exudates are more associated with the mineral  
22 matrix of the aggregate (i.e., the DF), decreasing its C:N ratio. A key aspect of this conceptual  
23 model is that more recently-formed aggregates with carbohydrate-rich cores are resistant to



1 ultrasonic disruption and ultimately separate into one of the intermediate densities (1.80-2.0 g  
2 mL<sup>-1</sup>), which corresponds to part of the DF in the present study. Thus, if the combination of  
3 physical and ultrasonic disruption we applied to the soil did not completely disperse recently-  
4 formed micro-aggregates, some of these micro-aggregates may have contained elevated <sup>14</sup>C and  
5 ultimately separated into the DF. This is essentially what we observed: high incorporation of the  
6 recent pulse-<sup>14</sup>C into the free LF, concurrent with or followed closely by the elevated <sup>14</sup>C in the  
7 DF. The occluded LF appeared to incorporate little of the <sup>14</sup>C pulse. If the model continues to  
8 accurately predict processes in these soils in subsequent years, the elevated soils at TVA should  
9 show a dilution of the elevated  $\Delta^{14}\text{C}$  in the free LF and DF, and a concomitant increase in the  
10  $\Delta^{14}\text{C}$  in the occluded LF.

11 Baisden et al. (2002b) proposed an alternate conceptual model to explain natural abundance  
12 <sup>15</sup>N and <sup>14</sup>C concentrations in density fractions separated similar to those of Golchin et al.  
13 (1994b) from several grassland soils in the Central Valley of California. Namely, that the free  
14 LF is humified and gradually converted to occluded LF as carboxylic groups both increase and  
15 bind with clay. To the extent that microbial byproducts are stabilized during the gradual process  
16 of occlusion and mineral association, the C:N ratio of the protected fractions could be expected  
17 to be lower than the initial plant-like value. Continued degradation of the occluded fraction is  
18 negligible until the aggregate is physically disturbed. Considering the results from TVA within  
19 the context of the Baisden model, as the elevated <sup>14</sup>C is removed from the free LF through  
20 degradation or occlusion within aggregates, the <sup>14</sup>C concentrations in the occluded LF and the  
21 DF should rise concurrently. Indeed, both fractions appeared to incorporate some of the elevated  
22 <sup>14</sup>C, but the incorporation was heavily weighted to the DF. These initial results appear to be  
23 more clearly explained and predicted by the Golchin conceptual model.

## 1 **5. Concluding Remarks**

2 We have shown that the three-pool density fractionation introduced by Golchin et al.  
3 (1994a), using density and increasing aggregate disruption to fractionate soil, has been successful  
4 in isolating meaningful soil organic and organo-mineral fractions from upland Ultisols at Oak  
5 Ridge Reservation, Tennessee. These fractions (free LF, occluded LF, and DF) differ in  
6 fundamental qualities such as C concentration, C:N ratio,  $\Delta^{14}\text{C}$ , change with soil depth, and the  
7 timing and magnitude of response to inputs of elevated  $^{14}\text{C}$ . In these soils the free LF (inter-  
8 aggregate POM) is clearly the most responsive and least protected organic fraction. The  
9 occluded LF (intra-aggregate POM) is more isolated from recent plant inputs than the DF  
10 (mineral-associated C), although this does not mean that occlusion within these aggregates  
11 provides more C stability than organo-mineral interactions. On the contrary, the  $^{14}\text{C}$  values from  
12 Walker Branch and TVA suggest that whereas the DF quickly incorporates plant inputs, it also  
13 contains a much older and more stable C pool than does the occluded fraction. The increase in  
14 the difference in  $\Delta^{14}\text{C}$  between unprotected and protected SOM with depth suggests that the  
15 importance of physical and organo-mineral C stabilization processes, compared to the  
16 importance of rates of C input from plants, increases from the surface to deep soil.

17 In subsequent EBIS work we will use these methods to trace the movement and stabilization  
18 of  $^{14}\text{C}$  from roots and litter using reciprocal litter transplants across the  $^{14}\text{C}$ -enrichment gradient.  
19 The combination of the ecosystem  $^{14}\text{C}$  enrichment gradient, the litter transplants, and the density  
20 fractionation, should give us insight into the specific roles of root versus litter inputs into these  
21 stabilization process.

22

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15

**16 References**

17 Alvarez, R. and Alvarez, C.R., 2000. Soil organic matter pools and their associations with carbon  
18 mineralization kinetics. *Soil Sci. Soc. Am. J.*, 64: 184-189.

19 Amalfitano, C., Quezada, R.A., Wilson, M.A. and Hanna, J.V., 1995. Chemical-Composition of  
20 Humic Acids - a Comparison with Precursor Light Fraction Litter from Different  
21 Vegetations Using Spectroscopic Techniques. *Soil Sci.*, 159: 391-401.

22 Baisden, W.T., Amundson, R., Brenner, D.L., Cook, A.C., Kendall, C. and Harden, J.W., 2002a.  
23 A multiisotope C and N modeling analysis of soil organic matter turnover and transport

- 1 as a function of soil depth in a California annual grassland soil chronosequence. *Glob.*  
2 *Biogeochem. Cycle*, 16: 1135.
- 3 Baisden, W.T., Amundson, R., Cook, A.C. and Brenner, D.L., 2002b. Turnover and storage of C  
4 and N in five density fractions from California annual grassland surface soils. *Glob.*  
5 *Biogeochem. Cycle*, 16: 1117.
- 6 Baldock, J.A., Oades, J.M., Nelson, P.N., Skene, T.M., Golchin, A. and Clarke, P., 1997.  
7 Assessing the extent of decomposition of natural organic materials using solid-state C-13  
8 NMR spectroscopy. *Aust. J. Soil Res.*, 35: 1061-1083.
- 9 Baldock, J.A., Oades, J.M., Waters, A.G., Peng, X., Vassallo, A.M. and Wilson, M.A., 1992.  
10 Aspects of the Chemical-Structure of Soil Organic Materials as Revealed by Solid-State  
11 C-13 NMR-Spectroscopy. *Biogeochemistry*, 16: 1-42.
- 12 Boone, R.D., 1994. Light-fraction soil organic matter: origin and contribution to net nitrogen  
13 mineralization. *Soil Biol. Biochem.*, 26: 1459-1468.
- 14 Certini, G., Corti, G., Agnelli, A. and Sanesi, G., 2003. Carbon dioxide efflux and concentrations  
15 in two soils under temperate forests. *Biol. Fertil. Soils*, 37: 39-46.
- 16 Chotte, J.L., Ladd, J.N. and Amato, M., 1998. Sites of microbial assimilation, and turnover of  
17 soluble and particulate <sup>14</sup>C-labelled substrates decomposing in a clay soil. *Soil Biol.*  
18 *Biochem.*, 30: 205-218.
- 19 Christensen, B.T., 1992. Physical fractionation of soil and organic matter in primary particle size  
20 and density separates. *Advances in Soil Science*, 20: 1-90.
- 21 Compton, J.E. and Boone, R.D., 2000. Long-term impacts of agriculture on soil carbon and  
22 nitrogen in New England forests. *Ecology*, 81: 2314-2330.

- 1 Cromack, K., Miller, R.E., Helgerson, O.T., Smith, R.B. and Anderson, H.W., 1999. Soil carbon  
2 and nutrients in a coastal Oregon Douglas-fir plantation with red alder. *Soil Sci. Soc. Am.*  
3 *J.*, 63: 232-239.
- 4 Dalal, R.C. and Mayer, R.J., 1986. Long-term trends in fertility of soils under continuous  
5 cultivation and cereal cropping in southern Queensland. IV. Loss of organic carbon for  
6 different density functions. *Aust. J. Soil Res.*, 24: 301-309.
- 7 Gaudinski, J.B., 2001. Belowground carbon cycling in three temperate forests of the Eastern  
8 United States. Ph.D. Thesis, University of California-Irvine, 285 pp.
- 9 Gaudinski, J.B., Trumbore, S.E., Davidson, E.A. and Zheng, S.H., 2000. Soil carbon cycling in a  
10 temperate forest: radiocarbon-based estimates of residence times, sequestration rates and  
11 partitioning of fluxes. *Biogeochemistry*, 51: 33-69.
- 12 Goh, K.M., Stout, J.D. and Rafter, T.A., 1977. Radiocarbon Enrichment of Soil Organic-Matter  
13 Fractions in New Zealand Soils. *Soil Sci.*, 123: 385-391.
- 14 Golchin, A., Clarke, P. and Oades, J.M., 1996. The heterogeneous nature of microbial products  
15 as shown by solid-state C-13 CP/MAS NMR spectroscopy. *Biogeochemistry*, 34: 71-97.
- 16 Golchin, A., Oades, J.M., Skjemstad, J.O. and Clarke, P., 1994a. Study of Free and Occluded  
17 Particulate Organic-Matter in Soils by Solid-State C-13 CP/MAS NMR-Spectroscopy  
18 and Scanning Electron-Microscopy. *Aust. J. Soil Res.*, 32: 285-309.
- 19 Golchin, A., Oades, J.M., Skjemstad, J.O. and Clarke, P., 1994b. Soil-Structure and Carbon  
20 Cycling. *Aust. J. Soil Res.*, 32: 1043-1068.
- 21 Greenland, D.J. and Ford, G.W., 1964. Separation of partially humified organic materials by  
22 ultrasonic dispersion. Eighth International Congress of Soil Science, Transactions, 3:  
23 137-148.

- 1 Gregorich, E.G., Monreal, C.M., Schnitzer, M. and Schulten, H.-R., 1996. Transformation of  
2 plant residues into soil organic matter: chemical characterization of plant tissue, isolated  
3 soil fractions, and whole soils. *Soil Science*, 161: 680-693.
- 4 Hungate, B.A., Jackson, R.B., Field, C.B. and Chapin, F.S., 1996. Detecting changes in soil  
5 carbon in CO<sub>2</sub> enrichment experiments. *Plant and Soil*, 187: 135-145.
- 6 Janzen, H.H., Campbell, C.A., Brandt, S.A., Lafond, G.P. and Townley-Smith, L., 1992. Light-  
7 fraction organic matter in soils from long-term crop rotations. *Soil Sci. Soc. Am. J.*, 56:  
8 1799-1806.
- 9 Kaiser, K., Eusterhues, K., Rumpel, C., Guggenberger, G. and Kogel-Knabner, I., 2002.  
10 Stabilization of organic matter by soil minerals - investigations of density and particle-  
11 size fractions from two acid forest soils. *J. Plant Nutr. Soil Sci.-Z. Pflanzenernahr.*  
12 *Bodenkd.*, 165: 451-459.
- 13 Levin, I. and Hesshaimer, V., 2000. Radiocarbon - A unique tracer of global carbon cycle  
14 dynamics. *Radiocarbon*, 42: 69-80.
- 15 Oades, J.M., 1988. The Retention of Organic-Matter in Soils. *Biogeochemistry*, 5: 35-70.
- 16 Parker, J.L., Fernandez, I.J., Rustad, L.E. and Norton, S.A., 2002. Soil organic matter fractions in  
17 experimental forested watersheds. *Water Air Soil Pollut.*, 138: 101-121.
- 18 Paul, E.A., Collins, H.P. and Leavitt, S.W., 2001. Dynamics of resistant soil carbon of  
19 Midwestern agricultural soils measured by naturally occurring <sup>14</sup>C abundance.  
20 *Geoderma*, 104: 239-256.
- 21 Rumpel, C., Kogel-Knabner, I. and Bruhn, F., 2002. Vertical distribution, age, and chemical  
22 composition of organic carbon in two forest soils of different pedogenesis. *Organic*  
23 *Geochemistry*, 33: 1131-1142.

- 1 Sohi, S.P., Mahieu, N., Arah, J.R.M., Powlson, D.S., Madari, B. and Gaunt, J.L., 2001. A  
2 Procedure for Isolating Soil Organic Matter Fractions Suitable for Modeling. *Soil Sci.*  
3 *Soc. Am. J.*, 65: 1121-1128.
- 4 Sollins, P., Glassman, C.A., Paul, E.A., Swanston, C., Lajtha, K., Heil, J.W. and Elliott, E.T.,  
5 1999. Soil Carbon and Nitrogen: Pools and Fractions. In: G.P. Robertson, C.S. Bledsoe,  
6 D.C. Coleman and P. Sollins (Editors), *Standard Soil Methods for Long-Term Ecological*  
7 *Research*. Oxford University Press, New York, pp. 89-105.
- 8 Sollins, P., Homann, P. and Caldwell, B.A., 1996. Stabilization and destabilization of soil  
9 organic matter: Mechanisms and controls. *Geoderma*, 74: 65-105.
- 10 Spycher, G., Sollins, P. and Rose, S., 1983. Carbon and Nitrogen in the Light Fraction of a  
11 Forest Soil - Vertical-Distribution and Seasonal Patterns. *Soil Sci.*, 135: 79-87.
- 12 Stuiver, M. and Polach, H.A., 1977. Reporting of C-14 Data. *Radiocarbon*, 19: 355-363.
- 13 Swanston, C., Homann, P.S., Caldwell, B.A., Myrold, D.D., Ganio, L. and Sollins, P., 2004.  
14 Long-term effects of elevated nitrogen on forest soil organic matter stability.  
15 *Biogeochemistry*, In press.
- 16 Swanston, C.W., Caldwell, B.A., Homann, P.S., Ganio, L. and Sollins, P., 2002. Carbon  
17 dynamics during a long-term incubation of separate and recombined density fractions  
18 from seven forest soils. *Soil Biology and Biochemistry*, 34: 1121-1130.
- 19 Torn, M.S., Trumbore, S.E., Chadwick, O.A., Vitousek, P.M. and Hendricks, D.M., 1997.  
20 Mineral control of soil organic carbon storage and turnover. *Nature*, 389: 170-173.
- 21 Trumbore, S., Gaudinski, J.B., Hanson, P.J. and Southon, J.R., 2002. Quantifying ecosystem-  
22 atmosphere carbon exchange with a <sup>14</sup>C label. *Eos, Transactions, AGU*, 83: 265, 267-268.

- 1 Trumbore, S.E., 1993. Comparison of Carbon Dynamics in Tropical and Temperate Soils Using  
2 Radiocarbon Measurements. *Glob. Biogeochem. Cycle*, 7: 275-290.
- 3 Trumbore, S.E., Vogel, J.S. and Southon, J.R., 1989. AMS C-14 Measurements of Fractionated  
4 Soil Organic-Matter - an Approach to Deciphering the Soil Carbon-Cycle. *Radiocarbon*,  
5 31: 644-654.
- 6 Trumbore, S.E. and Zheng, S.H., 1996. Comparison of fractionation methods for soil organic  
7 matter C- 14 analysis. *Radiocarbon*, 38: 219-229.
- 8 Vogel, J.S., Southon, J.R., Nelson, D.E. and Brown, T.A., 1984. Performance of Catalytically  
9 Condensed Carbon for Use in Accelerator Mass-Spectrometry. *Nucl. Instrum. Methods*  
10 *Phys. Res. Sect. B-Beam Interact. Mater. Atoms*, 233: 289-293.

11

12



1 Table 1. Mass recovery, C and N concentrations, and distribution of soil C in density fractions in  
 2 the 0-15-cm and 15-30-cm soil increments of Walker Branch. Values are  $\pm 1$  SE.

3

4

|                | Mass recovery    | C <sup>a</sup>        | N <sup>a</sup>        | C :N           | Distribution of C             |
|----------------|------------------|-----------------------|-----------------------|----------------|-------------------------------|
|                | (% of bulk soil) | -g kg <sup>-1</sup> - | -g kg <sup>-1</sup> - |                | (% of bulk soil) <sup>b</sup> |
| 0-15 cm        |                  |                       |                       |                |                               |
| Bulk soil      | NA               | 24.89 $\pm$ 2.20      | 0.97 $\pm$ 0.04       | 25.5 $\pm$ 1.5 | NA                            |
| Free LF        | 2.17 $\pm$ 0.37  | 299.79 $\pm$ 7.94     | 8.85 $\pm$ 0.20       | 34.1 $\pm$ 1.7 | 25.52 $\pm$ 2.44              |
| Occl LF        | 1.08 $\pm$ 0.11  | 388.57 $\pm$ 9.31     | 9.31 $\pm$ 0.40       | 42.3 $\pm$ 2.0 | 16.74 $\pm$ 0.80              |
| DF             | 96.74 $\pm$ 0.47 | 12.46 $\pm$ 0.54      | 0.72 $\pm$ 0.03       | 17.3 $\pm$ 0.6 | 49.69 $\pm$ 2.47              |
| Total recovery | 99.95 $\pm$ 0.20 | NA                    | NA                    | NA             | 91.83 $\pm$ 0.82              |
| 15-30 cm       |                  |                       |                       |                |                               |
| Bulk soil      | NA               | 5.89 $\pm$ 0.51       | 0.17 $\pm$ 0.02       | 35.6 $\pm$ 2.6 | NA                            |
| Free LF        | 0.44 $\pm$ 0.05  | 326.31 $\pm$ 8.57     | 6.91 $\pm$ 0.42       | 45.5 $\pm$ 2.1 | 23.84 $\pm$ 1.86              |
| Occl LF        | 0.24 $\pm$ 0.02  | 360.93 $\pm$ 2.38     | 6.53 $\pm$ 0.63       | 57.2 $\pm$ 3.7 | 14.93 $\pm$ 1.19              |
| DF             | 99.31 $\pm$ 0.08 | 2.89 $\pm$ 0.25       | 0.24 $\pm$ 0.02       | 12.4 $\pm$ 0.8 | 51.93 $\pm$ 4.93              |
| Total recovery | 99.91 $\pm$ 0.54 | NA                    | NA                    | NA             | 85.92 $\pm$ 2.47              |

5

6 NA=not applicable.

7 <sup>a</sup>g kg<sup>-1</sup> of fraction.

8 <sup>b</sup>percentage of total soil C found in a given fraction.

9

10

1 Table 2. Analysis of variance (ANOVA) results for main effects and interactions of soil  
 2 variables at Walker Branch arranged and tested according to split-plot designs.

3

| Variable                         | Depth x Fraction |     |         | Depth           |     |      | Fraction        |      |         |
|----------------------------------|------------------|-----|---------|-----------------|-----|------|-----------------|------|---------|
|                                  | DF <sup>1</sup>  | F   | P       | DF <sup>1</sup> | F   | P    | DF <sup>a</sup> | F    | P       |
| $\Delta^{14}\text{C}$            | 2,28             | 6.5 | 0.0005  | 1,14            | 33  | NA   | 2,28            | 57   | NA      |
| C                                | 2,25             | 185 | <0.0001 | 1,14            | 157 | NA   | 2,25            | 4368 | NA      |
| N                                | 2,26             | 50  | <0.0001 | 1,14            | 159 | NA   | 2,26            | 2295 | NA      |
| C:N                              | 2,25             | 33  | <0.0001 | 1,14            | 1.3 | NA   | 2,25            | 456  | NA      |
| Soil C <sub>F</sub> <sup>b</sup> | 2,25             | 0.1 | 0.95    | 1,14            | 2.0 | 0.18 | 2,25            | 173  | <0.0001 |

4

5 <sup>a</sup>DF = degrees of freedom.

6 <sup>b</sup>Soil C<sub>F</sub> = the percentage of total soil C found in fractions.

7 NA=not applicable; *P*-values are excluded due to significant interaction with other variable.

8

1 Table 3. Mass recovery, C and N concentrations, and distribution of soil C in density fractions in  
 2 the 0-15-cm and 15-30-cm soil increments of TVA. Values are  $\pm 1$  SE.

3

|                | Mass<br>(% of bulk soil) | C <sup>a</sup><br>-g kg <sup>-1</sup> - | N <sup>a</sup><br>-g kg <sup>-1</sup> - | C :N           | Distribution of C<br>(% of bulk soil) <sup>b</sup> |
|----------------|--------------------------|---|---|----------------|--|
| 0-15 cm        |                          |   |   |                |  |
| Bulk soil      | NA                       | 24.87 $\pm$ 0.72                        | 1.18 $\pm$ 0.05                         | 21.2 $\pm$ 0.6 | NA   |
| Free LF        | 2.16 $\pm$ 0.10          | 295.64 $\pm$ 5.03                       | 10.29 $\pm$ 0.36                        | 28.9 $\pm$ 0.8 | 25.58 $\pm$ 0.82                                   |
| Occl LF        | 0.88 $\pm$ 0.06          | 376.18 $\pm$ 11.14                      | 8.54 $\pm$ 0.44                         | 44.6 $\pm$ 2.0 | 13.33 $\pm$ 1.03                                   |
| DF             | 96.96 $\pm$ 0.08         | 13.33 $\pm$ 0.45                        | 0.88 $\pm$ 0.04                         | 15.2 $\pm$ 0.3 | 51.92 $\pm$ 1.10                                   |
| Total recovery | 99.84 $\pm$ 0.14         | NA                                      | NA                                      | NA             | 90.46 $\pm$ 1.33                                   |
| 15-30 cm       |                          |   |   |                |  |
| Bulk soil      | NA                       | 8.50 $\pm$ 0.55                         | 0.42 $\pm$ 0.03                         | 20.4 $\pm$ 1.1 | NA   |
| Free LF        | 0.50 $\pm$ 0.04          | 318.27 $\pm$ 2.68                       | 8.63 $\pm$ 0.30                         | 37.1 $\pm$ 1.3 | 19.27 $\pm$ 0.87                                   |
| Occl LF        | 0.32 $\pm$ 0.03          | 410.42 $\pm$ 12.85                      | 7.60 $\pm$ 0.35                         | 54.4 $\pm$ 1.8 | 15.59 $\pm$ 0.76                                   |
| DF             | 99.18 $\pm$ 0.06         | 4.58 $\pm$ 0.35                         | 0.40 $\pm$ 0.02                         | 11.4 $\pm$ 0.5 | 54.71 $\pm$ 1.03                                   |
| Total recovery | 99.36 $\pm$ 0.20         | NA                                      | NA                                      | NA             | 89.57 $\pm$ 1.39                                   |

4

5 NA=not applicable

6 <sup>a</sup>g kg<sup>-1</sup> of fraction

7 <sup>b</sup> percentage of total soil C found in a given fraction

8

1 Table 4. Analysis of variance (ANOVA) results for main effects and interactions of soil  
 2 variables at TVA arranged and tested according to split-plot designs.

3

| Variable                         | Depth x Fraction |      |         | Depth <sup>a</sup> |      | Fraction <sup>a</sup> |       |
|----------------------------------|------------------|------|---------|--------------------|------|-----------------------|-------|
|                                  | DF <sup>b</sup>  | F    | P       | DF <sup>b</sup>    | F    | DF <sup>b</sup>       | F     |
| $\Delta^{14}\text{C}$            | 2,26             | 4.3  | 0.0245  | 1,13               | 287  | 2,26                  | 307   |
| C                                | 2,26             | 61.4 | <0.0001 | 1,13               | 0.01 | 2,26                  | 11492 |
| N                                | 2,26             | 30.3 | <0.0001 | 1,13               | 73   | 2,26                  | 2170  |
| C:N                              | 2,26             | 52.9 | <0.0001 | 1,13               | 6.4  | 2,26                  | 53    |
| Soil C <sub>F</sub> <sup>c</sup> | 2,26             | 10.5 | 0.0006  | 1,13               | 0.52 | 2,26                  | 895   |

4

5 <sup>a</sup>P-values are excluded due to significant interaction with other variable.

6 <sup>b</sup>DF = degrees of freedom.

7 <sup>c</sup>Soil C<sub>F</sub> = the percentage of total soil C found in fractions.

8

1 Figure 1.  $\Delta^{14}\text{C}$  from tree ring cellulose from in or near the eastern end of the Oak Ridge  
2 Reservation (open symbols) and the western end of the Reservation (closed symbols). Adapted  
3 from Trumbore et al. (2002).

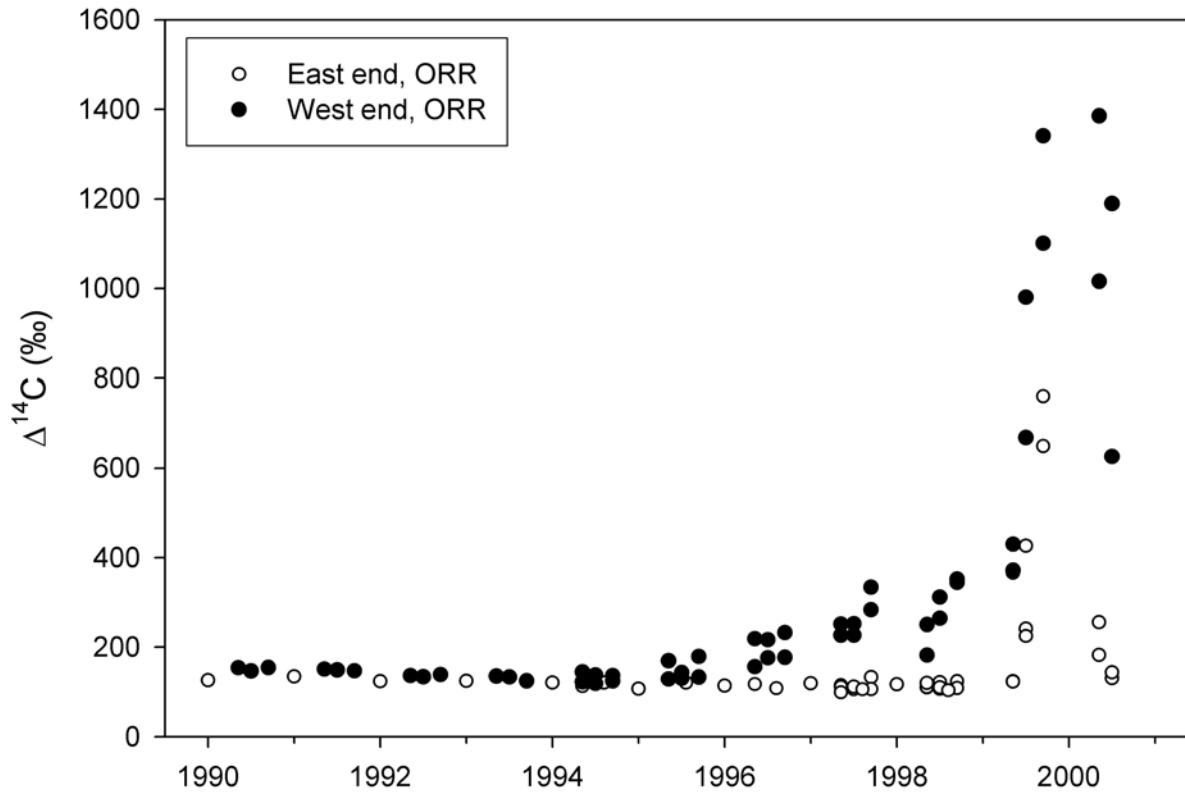
4

5 Figure 2.  $\Delta^{14}\text{C}$  (‰) of density fractions and bulk soil from 0-15 cm and 15-30 cm at Walker  
6 Branch. Error bars are 1 SE.

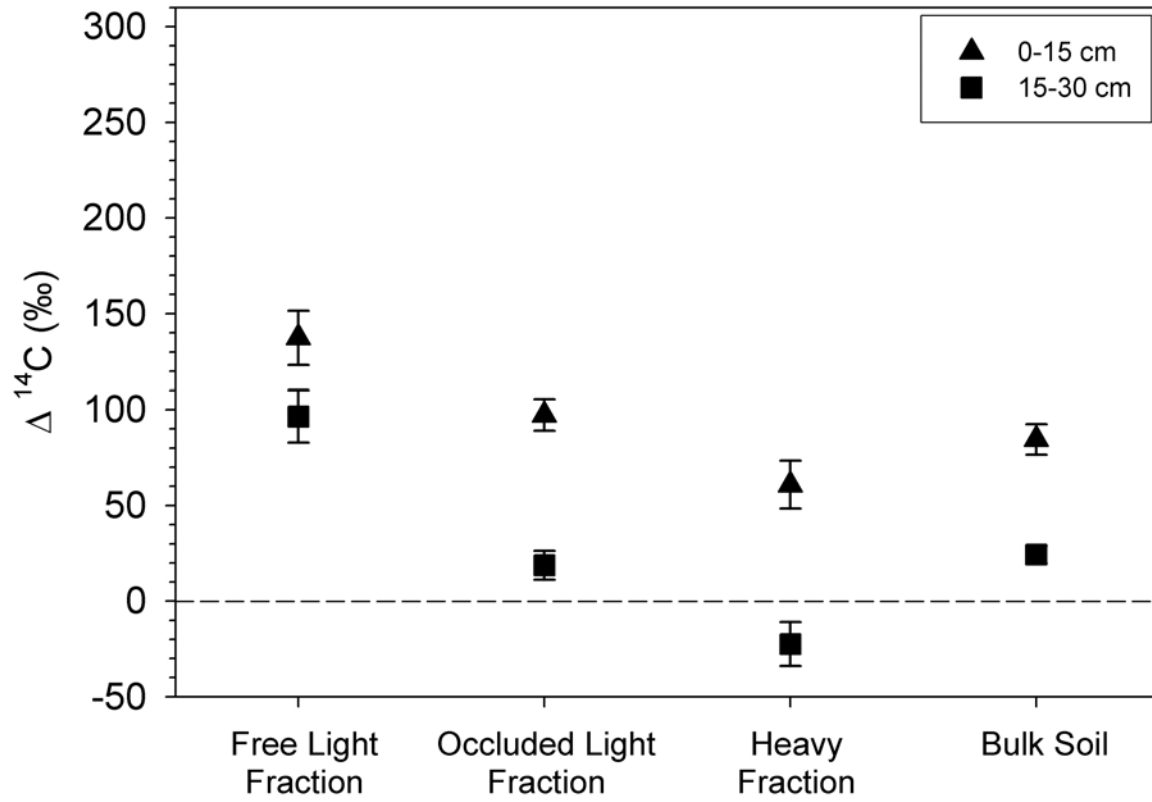
7

8 Figure 3.  $\Delta^{14}\text{C}$  (‰) of density fractions and bulk soil from 0-15 cm and 15-30 cm at TVA.  
9 Error bars are 1 SE.

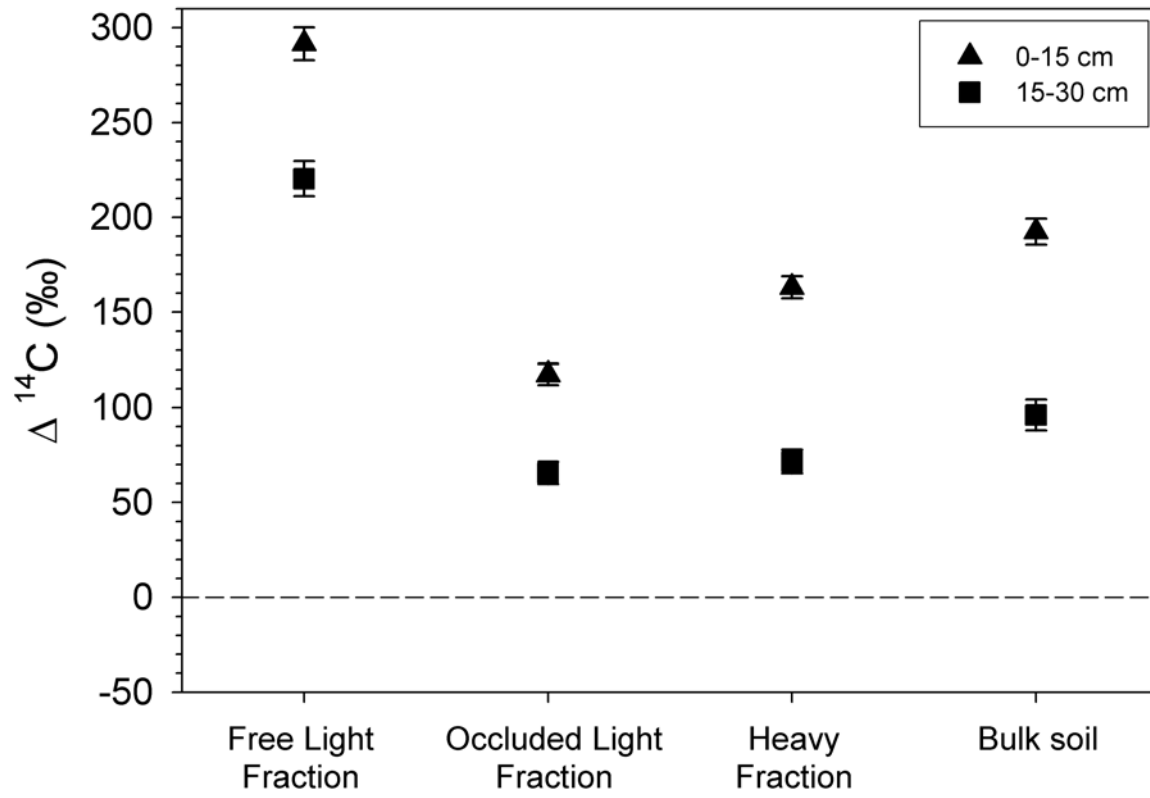
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Figure 3.