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**BIOLOGICAL SURVEYS ON THE
SAVANNAH RIVER IN THE VICINITY OF
THE SAVANNAH RIVER PLANT (1951-1976)**

ROBIN A. MATTHEWS

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**E. I. du Pont de Nemours & Co.
Savannah River Laboratory
Aiken, SC 29808**

PREPARED FOR THE U. S. DEPARTMENT OF ENERGY UNDER CONTRACT DE-AC09-76SR00001

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by

ROBIN A. MATTHEWS

Approved by:

T. V. Crawford, Research Manager
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ACKNOWLEDGEMENTS

The Savannah River Laboratory Data Base was created by J. E. Suich and J. H. Hightower, of the Savannah River Laboratory Computer Systems Division, and A. S. Dicks, of the Ecology Group. I would like to acknowledge their participation in that extensive project, and their many contributions to this report. Mike Odner, of the Academy of Natural Sciences of Philadelphia, has assisted in editing the computer data set. J. C. Corey, H. E. Mackey, T. V. Crawford and R. Patrick have reviewed this manuscript and provided editorial comments.

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COMMENTS

by

Dr. Ruth Patrick
Academy of Natural Sciences of Philadelphia
December 28, 1981

In 1951, the Academy of Natural Sciences of Philadelphia was contracted to initiate a long-term monitoring program to evaluate the effects of the Savannah River Plant on the Savannah River aquatic system. The purpose of this study was to establish conditions in the aquatic ecosystem:

- (1) in the Savannah River immediately above the Savannah River Plant,
- (2) just below various streams entering the Savannah River from the Savannah River Plant site, and
- (3) in the Savannah River as it leaves the area of the Savannah River Plant.

The program was designed to monitor at frequent intervals the easily assayed conditions of the Savannah River using diatometers and quarterly studies. The more subtle changes in the river were to be determined by thorough surveys of the associations of the major kinds of aquatic life and the chemical and physical conditions of the river. These surveys were made at approximately four year intervals. In designing the program of study, there was no attempt to place study areas so as to detect the effects from any particular effluent produced by the Plant, but rather to detect major effects that might appear after a reasonable degree of mixing in the river.

This monitoring program has been continued up to the present, and provides a comprehensive data base covering 25 years of sampling. One of the great advantages of this program is that it has been conducted in basically the same way so that gradual changes over time can be assayed. The Savannah River Laboratory has computerized the Savannah River data, and they are available as described in this report. The Academy of Natural Sciences played a major role in making sure the computerized taxonomy was as correct as possible and consistent throughout the data set. One of the greatest problems in placing a large data base such as this one in a computer where it may be used for a long period of time is to make sure that there is a reliable taxonomic classification and the sources or authorities for this classification are clearly set forth. In this work, the following publications and personal

communications are the basis on which the taxonomic classifications were made.

Algae: We have discussed the classification of the algae with Dr. Paul Silva, Dr. Francis Drouet, and Dr. Charles W. Reimer. The general taxonomy of all groups except diatoms followed Bourrelly (1970), Silva (1960), and Drouet (1973). For diatoms Patrick and Reimer (1966, 1975) was used.

Protozoa: For general organization we have used Kudo (1966), Jahn and Jahn (1949), Schaeffer (1926), Kahl (1930-34). Pascher (1927), Pascher and Lammermann (1912, 1914), and Page (1976). For general classification we have asked the advice of Dr. John Cairns, Jr.

Invertebrates: For general discussion, particularly concerning species of mollusca, Dr. Samuel L. H. Fuller was consulted. For developing the classification, the following authorities were used: Porifera and Platyhelminthes - Pennak (1978); Nemertea - Pennak (1964) and Hyman (1951); Aschelminthes - Hyman (1951) and Edmondson (1959); Tardigrada - Pennak (1978); Bryozoa - Rogick (1959); Annelida - Brinkhurst and Jamieson (1971), Sybil Parker (personal communication); Mollusca - Taylor and Sohl (1962), Newell (1965), Turner (personal communication); Mollusca - Taylor and Sohl (1962), Newell (1965), Turner (personal communication); Arthropoda - Pennak (1978).

Insects: The greatest responsibility has rested on Dr. Selwyn S. Roback, who has utilized many references in developing his opinions.

Fish: Our principal consultants have been Dr. James E. Bohlke and Dr. William Smith-Vaniz. This classification has largely depended on Dahlberger and Scott (1971). However, Bailey *et al.* (1979) has in some cases been used.

No attempt has been made by the Savannah River Laboratory to interpret the data. However, the Academy of Natural Sciences of Philadelphia has published several reports on these data and intends to publish several more papers utilizing these data now that they have been computerized. The publications to date are listed in the reference section of this report.

There have been 16 comprehensive river surveys from 1951-1976. In addition to the comprehensive river surveys, diatometers were installed in the river in 1953, and have been used continuously up to the present. The types of data collected from these diatometers have differed somewhat over time. In the beginning, four quarterly detailed readings, which involved the construction of truncated normal curves, were carried out four

times a year. Every two weeks, rough readings were made in which slides were scanned to see if there was any definite change in the species present; however, these taxonomic identifications were not very thorough, and are not included here. This report includes the data from the comprehensive river surveys and the quarterly detailed diatometer readings. The diatometer data represent a semi-continuous biomonitoring program that may be used to supplement the major survey data.

Representative specimens of all species collected on all the comprehensive surveys were placed in the permanent collections of the Academy of Natural Sciences of Philadelphia. This enables anyone in the future to bring together the species that were associated at any point in time during the studies. It also enables the Academy to revalidate identifications of the diatoms from the diatometer collection, which were also placed in the permanent collections in the Academy of Natural Sciences of Philadelphia diatom herbaria.

Some of the most important considerations in carrying out a baseline study are to make sure the locations of the sampling stations are correctly selected. In the Savannah River, several factors had to be carefully considered in the location of such stations. The first requirement is to place the stations so that they show the impacts of the operations of the Savannah River Plant on the Savannah River, and to separate these changes from upstream effects. For this reason, the upstream station had to be located in an area close to, but above, potential outfalls, but also in an area where mixing was fairly complete in the river.

It was essential to have similar current patterns in the areas of study, which in turn produce similar shoaling areas and cutting areas. There must also be areas in which sediments are eroding and sediments are being deposited. In the Savannah River, the bed is unconsolidated sand. The best habitats for aquatic organisms are in the shallow waters, on floating debris, or on solid substrates that are impinged in the river bed. In the shallow water, the river bed consisted mostly of sand or sands with mud and silt. For these reasons, one would like to have a point bar at each of the river stations. Unfortunately, this was not possible at the two upper stations (Stations 1 and 3). However, near these stations, revetments had been placed in the river by the U. S. Corps of Engineers prior to the time of the first surveys. These areas contained habitats similar to those of a point bar. Around each one of these revetments there were depositing and eroding areas, and areas of shallow water which would make seining of fish easy. We, therefore, located the two upper stations (Stations 1 and 3) in the vicinity of revetments and the two lower stations (Stations 5 and 6) to include point bars.

It is also important to have the original location of the stations in regards to East-West or North-South directions as similar as possible in order that the exposure of the banks to light will be similar. One also has to be sure that similar amounts of debris are present in each of the stations because the dead trees and branches provide an excellent habitat for aquatic life. The dredging of the river by the Corps of Engineers, which removed some debris, definitely affected the availability of habitats and the population of sizes and various species.

The river communities and populations were sampled qualitatively from a wide variety of habitats. Experience has shown that if one collects from one to two hours after one obtains the last new entity in insects, macro-invertebrates and fish, one has collected the species that were established in the area. For micro-organisms such as protozoa and algae, all available habitats must be sampled. As a result, it takes at least one day, and often two days, to sample any area. Separate species lists were developed for algae, protozoa, macroinvertebrates, insects, and fish. Appendices A and B provide a summary of the biological and chemical data available for each of the major surveys. Appendix B.1 summarizes the survey data, averaged for each station by survey number. Appendix B.3 includes the diatometer data derived from the studies in which the truncated normal curve model of the community was constructed.

When one examines the numbers of species present, one must remember that there are many environmental factors that affect the presence of species. Also important is one's ability to collect the species, i.e. high water as opposed to low flow. Furthermore, species react in aquatic systems according to the total stress from various causes, as long as one stress is not acute, and causes death. We have found that, in any given area, a 33 percent change in species numbers from the average for that area is natural, and only when the change is greater than 50 percent is severe perturbation indicated. Stated another way, generally, the natural functions of the transfer of nutrients in the food web can be maintained so long as 66 percent of the natural pattern of diversity of the various major groups is maintained. This applies to those groups in which there are many species. For these reasons, we have considered as separate groups, the algae, the protozoa, the invertebrates other than insects, the insects, and the fish.

Many things have happened over time in the Savannah River and some of these changes seem to be correlated with shifts in numbers of species in some of the groups. The algal flora in the river is dominated by diatoms. Station 6 has had the most radical shifts in numbers of diatom species. At all stations since 1968, there has been a small but consistent reduction in numbers of species. This decrease in species numbers seems to be correlated with the increase in chlorides in the river as evidenced by the chemical

analyses. These changes were correlated with the start-up of certain industries known to have chlorides in their effluents. Because the diatom species found in the Savannah River are typically soft water (low conductivity) species, one might expect such correlations. In the other groups of algae, which have very few species, more detailed studies are needed to interpret changes.

In the protozoa there seems to have been an increase in flagellates since 1972 in most stations that seems to be correlated with an increase in ammonia concentrations in the river. Likewise, the increase in ciliates seems to be correlated with increases in bacteria in the river.

Of the invertebrates other than insects, the mollusca were the only ones represented by a fairly diverse fauna. They show an irregular pattern, but seem to be more numerous since the building of Clarks Hill Dam, which has resulted in less fluctuation of water levels. Much more study is needed of these invertebrates to understand their patterns.

The more sensitive insects, such as stoneflies, have tended to be fewer in the surveys since 1955-56, and these changes may be correlated with the increased pollution load in the river. More study is needed to support or refute this statement. There was evidence that dredging activities, which varied in intensity at the different stations in different years, influenced the numbers of caddisflies, which are filter-feeders. These correlations were noted in the reports of the Academy of Natural Sciences of Philadelphia.

Of the fish, the suckers, minnows, sunfish and perch were more numerous as to species numbers in the river. These seemed to have increased since the building of Clarks Hill Dam. However, it must be pointed out that restricted use of rotenone was not used in the first few survey, and has been used since then. This method allows retrieval of some species not otherwise caught.

In conclusion, it should be stated that these observations can be refined by more careful study of the data. The Academy of Natural Sciences of Philadelphia plans to continue analyzing these data in order to reevaluate and continue developing their conclusions.

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ABSTRACT

In 1951, the Academy of Natural Sciences of Philadelphia was contracted by the Savannah River Plant to initiate a long-term monitoring program in the Savannah River. The purpose of this program was to determine the effect of the Savannah River Plant on the Savannah River aquatic ecosystem. The data from this monitoring program have been computerized by the Savannah River Laboratory, and are summarized in this report.

During the period from 1951 - 1976, 16 major surveys were conducted by the Academy in the Savannah River. Water chemistry analyses were made, and all major biological communities were sampled qualitatively during the spring and fall of each survey year. In addition, quantitative diatom data have been collected quarterly since 1953. Major changes in the Savannah River basin, in the Savannah River Plant's activities, and in the Academy sampling patterns are discussed to provide a historical overview of the biomonitoring program. Appendices to the data report include a complete taxonomic listing of species collected from the Savannah River, and summaries of the entire biological and physicochemical data base.

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**BIOLOGICAL SURVEYS ON THE SAVANNAH RIVER IN THE
VICINITY OF THE SAVANNAH RIVER PLANT (1951-1976)**

CHAPTER I. INTRODUCTION

The Savannah River Plant is a U. S. Department of Energy facility operated by E. I. du Pont de Nemours and Company. It is designed to produce nuclear materials for national defense. The Savannah River Plant is located about 25 miles southeast of Augusta, Georgia, along the South Carolina side of the Savannah River. Construction of the Savannah River Plant began in February, 1951, and involved a peak construction force of 39,000 workers. The first production reactor began operations on December 28, 1953.

In 1951, the Academy of Natural Sciences of Philadelphia was contracted to initiate a long-term monitoring program to evaluate the effects of the Savannah River Plant on the Savannah River aquatic system. This monitoring program has been continued up to the present, and provides a comprehensive data base covering more than 25 years of sampling. The Savannah River data collected from 1951 through 1976 have been computerized by the Savannah River Laboratory, and are available as described in this report. No attempt has been made by the Savannah River Laboratory to interpret the data; however, the Academy of Natural Sciences of Philadelphia has published a number of data interpretations.¹⁻¹¹

This report can be divided into two sections. The abstract and Chapters I through IX describe the Savannah River Plant and the Academy's surveys in detail. These chapters also provide a historical review of the Savannah River Basin from 1951 - 1976, and outline some of the changes that have occurred in the Savannah River Plant area that may have influenced the survey. The second section consists of Appendices A, B and C, which list the Savannah River Taxonomy and summarize much of the physical, chemical, and biological data from the Savannah River. These Appendices are included to show some examples of computer formats available for analyzing the data.

CHAPTER II. PERSONNEL

During the course of this study, Dr. Patrick has been responsible for the design of the studies and for the integration of results as set forth in the various Academy reports.

Members of the Academy of Natural Sciences, Philadelphia, who participated in the Savannah River surveys include the following individuals:

Survey 1 (June 25 - July 14, 1951)

John Cairns, Jr.	Protozoologist
Thomas Dolan, IV.	Entomologist
Harold W. Harry	Invertebrate Zoologist
John Lattin	Entomologist
Ruth Patrick	Director of Survey
Wilbur E. Wade	Algologist
John H. Wallace	Algologist
John M. Ward	Chemist and Bacteriologist
Charles B. Wurtz	Invertebrate Zoologist

Survey 2 (Oct. 15 - Nov. 31, 1951)

Fairie Lyn Carter	Chemist and Bacteriologist
Thomas Dolan	Entomologist
Sidney Kantor	Protozoologist
George W. McCammon	Ichthyologist
Ruth Patrick	Director of Survey and Algologist
Selwyn S. Roback	Entomologist
John H. Wallace	Algologist
Charles B. Wurtz	Invertebrate Zoologist

Survey 3 (Jan. 9 - 31, 1952)

Fairie Lyn Carter	Chemist and Bacteriologist
Thomas Dolan, IV.	Entomologist
George W. McCammon	Ichthyologist
Ruth Patrick	Director of Survey and Algologist
Selwyn S. Roback	Entomologist
John H. Wallace	Algologist
Ralph Wichterman	Protozoologist
Charles B. Wurtz	Invertebrate Zoologist

Survey 4 (May 5 - 22, 1952)

Fairie Lyn Carter	Chemist and Bacteriologist
Thomas Dolan, IV	Entomologist
George W. McCammon	Ichthyologist
Ruth Patrick	Director of Survey and Algologist
Selwyn S. Roback	Entomologist
John H. Wallace	Algologist
Ralph Wichterman	Protozoologist
Charles B. Wurtz	Invertebrate Zoologist

Survey 5 (Aug. 15 - 30, 1954)

Frederick A. Aldrich	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist
Harold de Ropp	Field Assistant
Ruth Patrick	Director of Survey
Selwyn S. Roback	Entomologist
John H. Wallace	Algologist

Survey 6 (Aug. 24 - Sept. 7, 1955)

Frederick A. Aldrich	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist, Field Director
Robert W. Haywood, III	Field Assistant
James Jacobs	Field Assistant
Ruth Patrick	Director of Survey
Charles W. Reimer	Algologist
Selwyn S. Roback	Entomologist
Yvonne H. Swabey	Chemist and Bacteriologist

Survey 7 (May 6 - 22, 1956)

Frederick A. Aldrich	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist, Field Director
Robert R. Grant, Jr.	Field Assistant
Ruth Patrick	Director of Survey
Charles W. Reimer	Algologist
Selwyn S. Roback	Entomologist
Yvonne H. Swabey	Chemist and Bacteriologist

Survey 8 (May 23 - June 4, 1960)

John M. Bates	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist, Project Supervisor
Phillip J. Halicki	Algologist
Nancy E. Hess	Chemist and Bacteriologist
Ruth Patrick	Director of Survey
Selwyn S. Roback	Entomologist

Survey 9 (Aug. 31 - Sept. 15, 1960)

John M. Bates	Invertebrate Zoologist
John Cairns, Jr.	Project Supervisor
Nancy E. Hess	Chemist and Bacteriologist
Thomas Lloyd	Field Assistant
Samson McDowell	Protozoologist
Ruth Patrick	Director of Survey
Charles Reimer	Algologist
Selwyn S. Roback	Entomologist

Survey 10 (May 30 - June 8, 1965)

John M. Bates	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist
Neal Foster	Ichthyologist
Robert R. Grant, Jr.	Algologist and Field Supervisor
Jules J. Loos	Field Assistant
Ruth Patrick	Director of Survey
Selwyn S. Roback	Entomologist

Survey 11 (Sept. 21 - 30, 1965)

John M. Bates	Invertebrate Zoologist
John Cairns, Jr.	Protozoologist
Neal Foster	Ichthyologist
C. W. Hart, Jr.	Invertebrate Zoologist and Field Supervisor
Jules J. Loos	Field Assistant
Samson McDowell	Protozoologist
Nicholas Nitti	Chemist and Bacteriologist
Ruth Patrick	Director of Survey
Charles Reimer	Algologist
Selwyn S. Roback	Entomologist

Survey 12 (May 27 - June 5, 1968)

Robert R. Grant, Jr.	Algologist and Field Supervisor
Robert W. Haug	Chemist
Irwin R. Isquith	Protozoologist
Edward Jankowski	Field Assistant
Gerald J. Lauer	Ichthyologist
Ruth Patrick	Director of Survey
Jay W. Richardson	Invertebrate Zoologist
Selwyn S. Roback	Entomologist

Survey 13 (Aug. 24 - Sept. 2, 1968)

Robert R. Grant, Jr.	Algologist and Field Supervisor
Robert W. Haug	Chemist
Irwin R. Isquith	Protozoologist
Edward Jankowski	Field Assistant
Gerald J. Lauer	Ichthyologist
Ruth Patrick	Director of Survey
Jay W. Richardson	Invertebrate Zoologist and Entomologist

Survey 14 (May 22 - June 2, 1972)

Martin DeGraw	Field Assistant
Neal Foster	Ichthyologist
Samuel Fuller	Invertebrate Zoologist
Robert R. Grant, Jr.	Phycologist and Field Supervisor
Edward Jankowski	Field Assistant
Ruth Patrick	Director of Survey
Jay W. Richardson	Entomologist
Jesse Steelman	Chemist
William Yongue	Protozoologist

Survey 15 (Sept. 13 - Oct. 1, 1972)

Neal Foster	Ichthyologist
Samuel Fuller	Invertebrate Zoologist
Robert R. Grant, Jr.	Field Supervisor
Edward Jankowski	Field Assistant
Joy Morrill	Phycologist
Ruth Patrick	Director of Survey
Jay W. Richardson	Entomologist
Selwyn Roback	Entomologist
Jesse Steelman	Chemist
William Yongue	Protozoologist

Survey 16 (Aug. 10-16, 1976)

Jules Loos	Ichthyologist
Samuel Fuller	Invertebrate Zoologist
Clyde Goulden	Director of Survey
Robert R. Grant, Jr.	Phycologist and Field Supervisor
Edward Jankowski	Field Assistant
Jay W. Richardson	Entomologist
Larry Lyons	Chemist
Irwin Isquith	Protozoologist
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The following authorities assisted in identification of a portion of the specimens collected:

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Leonora K. Gloyd Illinois Natural History Survey Urbana, Illinois (Surveys 1-4)	John C. Lutz American Entomological Societies Philadelphia, Pennsylvania (Surveys 1-9)
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Michigan State University
East Lansing, Michigan
(Surveys 1-7)

Employees of E. I. du Pont de Nemours and Company, Inc. who have been directly involved in the Savannah River surveys include Raymond S. Harvey, John H. Horton, C. M. Patterson, Everett W. Rabon, William C. Reinig, and Daniel I. Ross.

CHAPTER III. SAVANNAH RIVER BASIN HISTORY*

The Savannah River drainage basin has a total area of 10,579 square miles ($27,388 \text{ km}^2$), and encompasses all or part of 41 counties in Georgia, South Carolina and North Carolina (Figure 1). The Savannah River Basin is located in three physiographic regions: the Mountain Province, the Piedmont, and the Coastal Plain (Figure 2). The Mountain Province contains most of the major tributaries of the Savannah River, including the Seneca,

* Much of the information provided in this chapter was obtained from references 12-19.

NORTH CAROLINA

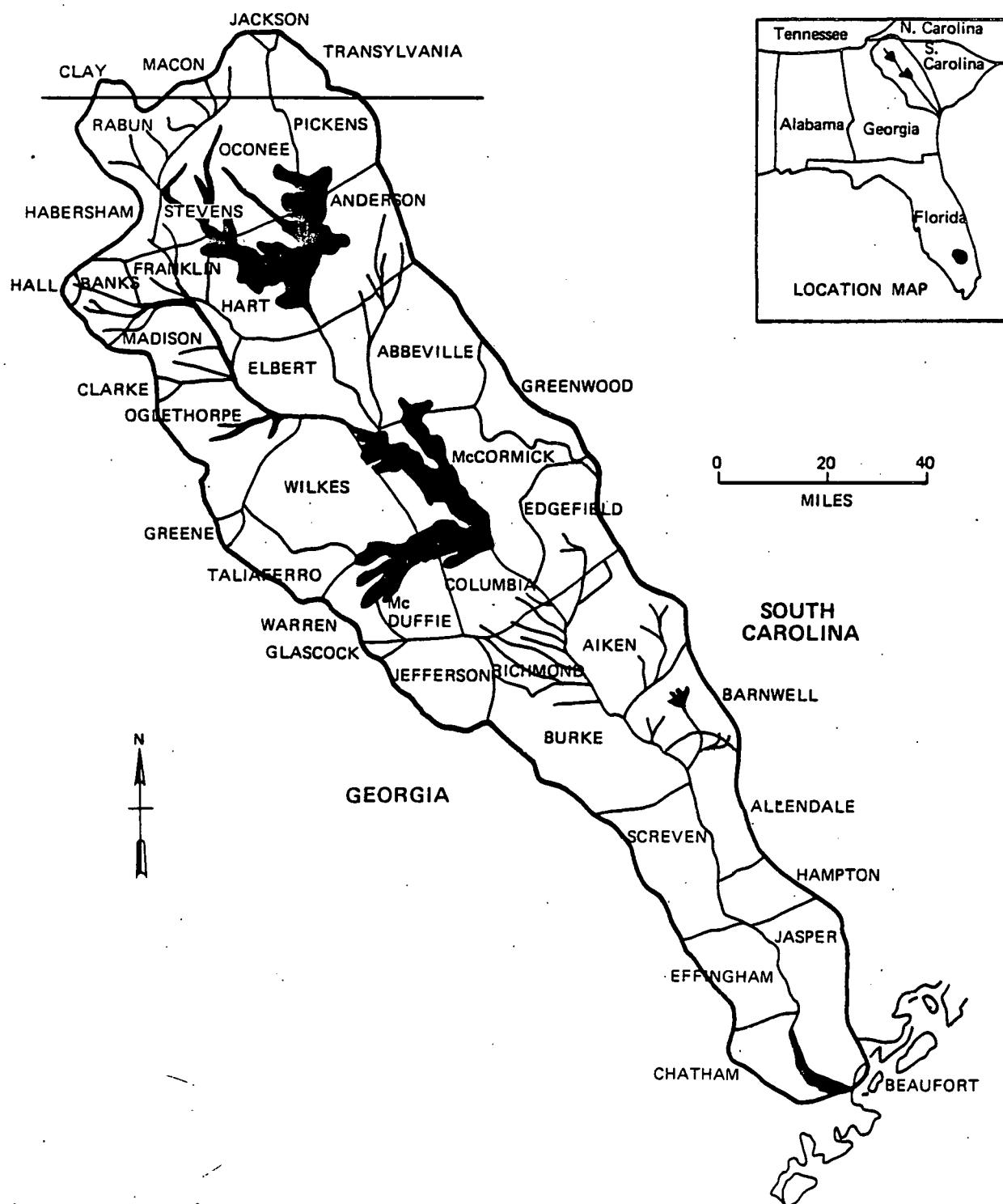


Figure 1. Counties Included Within the Savannah River Drainage Basin.

Reference 12.

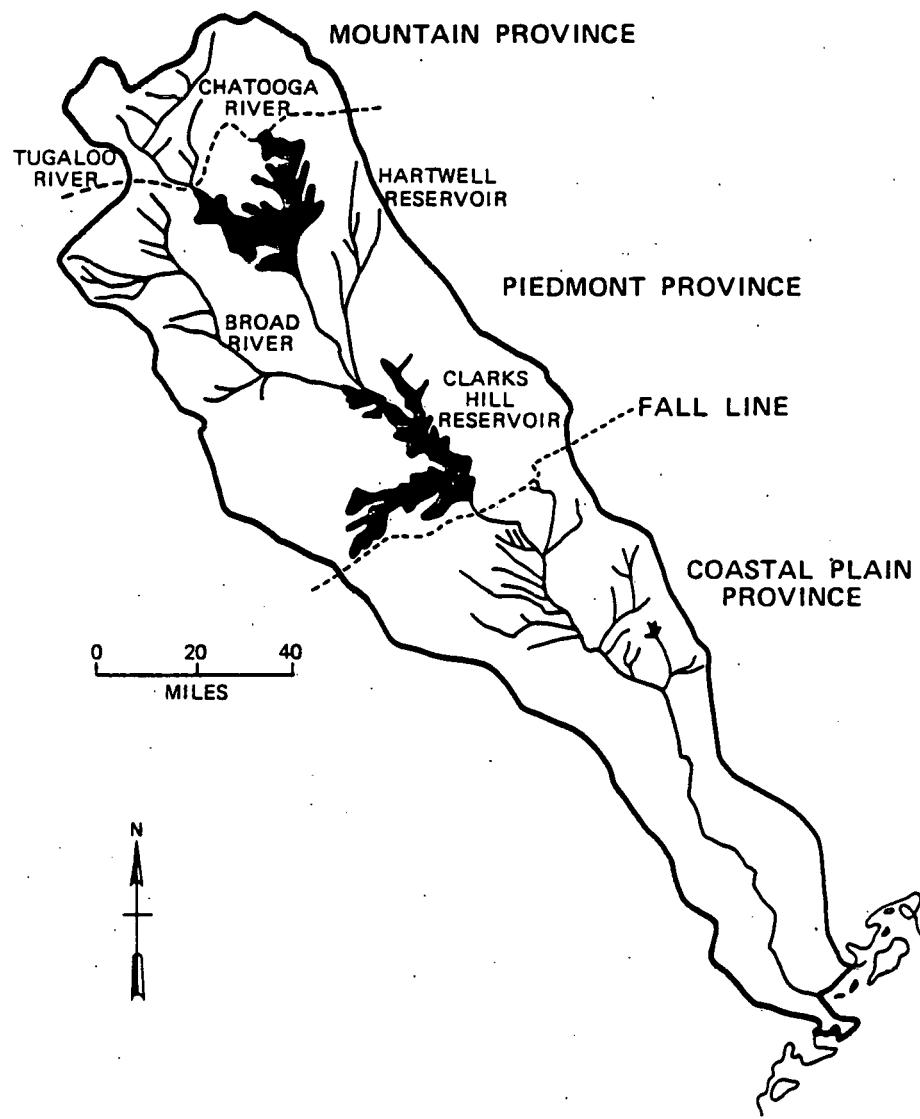


Figure 2. Major Tributaries and Physiographic Regions Included in the Savannah River Drainage Basin.

Reference 12.

Tugaloo and Chattooga Rivers. The region is characterized by a relatively steep gradient, ranging from about 5,500 to 1,000 ft (1676 to 305 m), and includes 2,022 sq mi (5235 km²) of the total drainage basin. The Mountain Province lies in the Blue Ridge, and has a bedrock composed of gneisses, granites, schists and quartzites; the subsoil is composed of brown and red sandy clays. In this region the Savannah River and its tributaries have the character of mountain streams, with shallow riffles, clear creeks, and a fairly steep gradient. The stream bed is mainly sand and rubble, and the banks are sloping and grass-covered.

The Piedmont Region has an intermediate gradient, with elevations ranging from 1,000 to 200 ft (305 to 61 m). This region includes 5,233 sq mi (13,548 km²) of the total drainage basin. Soils in the Piedmont are primarily red, sandy or silty clays, with weathered bedrock consisting of ancient sediments containing granitic intrusions. The Piedmont is bordered by the Fall-Line, an area where the sandy soils of the Coastal Plain meet the rocky terrain of the Piedmont foothills. The city of Augusta, Georgia, is located near this line.

The Savannah River becomes more turbid in the Piedmont Region, picking up the majority of its silt load. The river often meanders, and, despite the stabilizing effect of Clarks Hill Dam, the outer banks on curves are relatively unstable. Sand bars are deposited downstream from many inside banks along the curves.

The Coastal Plain has a negligible gradient ranging from an elevation of 200 ft (61 m) to sea level. The soils of this region are primarily stratified silts, clays, and sands. The Coastal Plain contains 3,334 sq mi (8631 km²) of the total Savannah River drainage area, and includes the city of Savannah, Georgia (Figure 2). In the Coastal Plain, the Savannah River is quite slow moving. Tidal effects may be observed near the mouth of the river, and a salt water tongue extends upstream along the bottom of the river bed. The river bed is often muddy, and the river proper merges with the surrounding swamps.

Flow data for the Savannah River at Augusta (measured at Butler Creek) are given in Table 1. Typically, low flow occurs during the fall while high water occurs in late winter or early spring. The average discharge at Augusta calculated from 72 years of data, is 10,300 cfs (292 m³/sec). More detailed flow and temperature data are given in Appendix C.

The water quality of the Savannah River and its tributaries varies considerably, ranging from clean to heavily contaminated with industrial and domestic effluents. The 1974 stream use

TABLE 1

**Minimum, Maximum, and Average Flows of the Savannah River
(Measured at Augusta)***

<u>Water Year (Oct-Sept)</u>	<u>Minimum (cfs)</u>	<u>Maximum (cfs)</u>	<u>Average (cfs)</u>
1950-51	1,710	41,400	6,766
1951-52	1,770	38,600	8,596
1952-53	3,260	34,800	6,561
1953-54	5,460	22,300	7,293
1954-55	4,180	22,400	5,487
1955-56	3,580	14,600	5,398
1956-57	5,170	15,200	6,572
1957-58	5,000	57,100	11,360
1958-59	5,260	28,200	7,125
1959-60	5,350	33,200	12,450
1960-61	4,930	30,300	8,873
1961-62	4,760	28,200	9,276
1962-63	5,130	30,600	9,554
1963-64	6,120	84,500	16,580
1964-65	6,300	33,000	12,940
1965-66	6,160	34,500	9,509
1966-67	5,740	20,900	8,372
1967-68	5,890	32,200	9,043
1968-69	5,800	44,100	9,812
1969-70	5,870	23,200	7,032
1970-71	4,460	59,700	8,668
1971-72	6,220	32,700	10,240
1972-73	5,460	38,100	13,200
1973-74	5,450	29,300	9,822
1974-75	5,520	43,900	11,690
1975-76	6,750	31,900	12,110
1976-77	6,000	32,200	11,030

* Reference 19.

classifications for the Savannah River Basin are given in Table 2. These classifications indicate the water quality at the time of the last Academy survey.

Historically, the Augusta, North Augusta, and Aiken County areas have provided the major sources of pollution to the Savannah River in the area around the Savannah River Plant. The city of Augusta did not have a secondary sewage treatment facility until 1975. Prior to that time most domestic and industrial wastes were discharged untreated or inadequately treated into the Savannah River, or into Hawks Gully, Butler Creek, and Spirit Creek, which flow into the Savannah River (Table 3). In the North Augusta and Aiken County area, domestic and industrial effluents entered the Savannah River directly and via Horse Creek and Lower Three Runs Creek (Table 4). Treatment facilities for the North Augusta and Aiken County area was not in operation until 1979. The Savannah River Plant also discharged waste water into the Savannah River. These discharges include thermal effluents as well as domestic and industrial wastes. The Savannah River Plant effluents are discussed in more detail in the next chapter.

Other pollution sources may be identified, particularly in the Savannah River Estuary near Savannah, Georgia. However, these sources did not have as great a potential for influencing the Academy of Natural Sciences of Philadelphia surveys as did those pollution sources from the Augusta area.

During the survey period there were a number of dams and impoundments built or in operation on the Savannah River. The largest of these were the Clarks Hill and the Hartwell Dams, located upstream from Augusta (Figure 2). The Clarks Hill Dam was completed in March, 1953 and is located 27 miles (43 km) above Augusta. Since the construction of Clarks Hill Dam, the Academy of Natural Sciences of Philadelphia survey teams have noted several changes in the Savannah River at their sampling station. River flow regulation increased the stability of the river banks by allowing vegetation, which previously had been torn loose from the banks by periodic floods, to build up along the river. In addition, there seemed to be an increase in water clarity as sediments settled out behind the dam. This increased clarity may have been partially responsible for the growth of dense algal mats in shallow river areas. The Academy hypothesized that because light was now able to penetrate farther into the water, the productivity of the river increased. The Academy also attributed part of the increased production to increasing industrial and domestic nutrient input into the river as cities and communities grew in the Savannah River Basin. Finally, the Academy noted a reduction in the number of snags and trash piles along the river.

TABLE 2

Stream Use Classification of the Savannah River (1974)*

<u>Stream</u>	<u>Reach</u>	<u>Classification**</u>
Chattooga	Georgia-North Carolina State Line to Tugalo Reservoir	Wild & Scenic
West Fork - Chattooga River	Confluence of Overflow Creek and Clear Creek to Chattooga River	Wild & Scenic
Savannah River	Headwaters of Tugalo Reservoir to Clarks Hill Dam	Recreation
Savannah River	Clarks Hill Dam to Augusta, 13th Street Bridge	Drinking Water
Butler Creek (and its tributaries)	Headwaters in Augusta to confluence with Savannah River	Urban
Cason's Dead River (and its tributaries)	Headwaters in Augusta to confluence with Savannah River	Urban
Savannah River	Augusta, 13th Street Bridge to U. S. Highway 301 Bridge	Fishing
Savannah River	U. S. Highway 301 Bridge to U. S. Highway 17 Bridge	Drinking Water
Savannah River	U. S. Highway 17 Bridge to Field's Cut	Industrial Navigation
Savannah River	Field's Cut to Fort Pulaski	Fishing
Savannah River	Fort Pulaski to open sea and all littoral waters of Tybee Island	Recreation

* Reference 14.

** Water quality classifications are defined in the Georgia Department of Natural Resources, Rules of the Environmental Protection Division.¹³

TABLE 3

**Partial List of the Savannah River Pollution Sources from the
Augusta Area***

<u>Name of Industry</u>	<u>Const. Date</u>	<u>Type of Wastewater</u>	<u>Est. Vol. of Wastewater (mgd) Prior to 1970</u>	<u>Treatment Prior to 1970**</u>	<u>Receiving Stream</u>
Augusta Chemical Co.	?	Organic, acidic, with sodium salts	0.13	Some Neutralization	Camille St. Ditch to
Augusta Plating Co.	1966	Cd, Cr, Cu, Zn, cyanide plating wastes	0.10	None	Oates Creek
Augusta Waste Water Treatment Facility	1969	Domestic sewage, textile finishing (23.0 max. wastes, chemical cap.) manufacturers wastes, slaughterhouse wastes	8.0	Primary and chlorination	Butler Creek
Babcock & Wilcox	1928	Oil, kaolin, sawdust	0.15	Settling tank	Unnamed tributary to Rocky Creek
Buckeye Cotton Oil Div. of Buckeye Cellulose Corp.	1902	Suspended & floating organics, oils	0.30	None	Camille St. Ditch to Beaverdam Ditch
Burris Chemical, Inc.	1969	Cooling water & chloride solutions	0.12	Retention area	Savannah River
Castleberry's Food Co.	1926	Organic	?	None	City sewer to Oates Creek
Columbia Nitrogen Corp. (Nitrogen Plant)	1963	Contaminated cooling water (Ammonia), domestic wastes	?	Containment and reuse, pH neutralization	Butler Creek

* Reference 15.

** Most Augusta industries currently direct their wastewaters to the Augusta Waste Water Treatment Facility, which was improved in 1975 to provide secondary treatment. Columbia Nitrogen Corp. and Continental Forest Industries currently have their own secondary treatment facility.

TABLE 3 (Contd)

<u>Name of Industry</u>	<u>Const. Date</u>	<u>Type of Wastewater</u>	<u>Estimated Vol. of Wastewater (mgd)</u>	<u>Treatment Prior to 1970</u>	<u>Receiving Stream</u>
Columbia Nitrogen Corp. (Caprolactam Plant)	1963	Cooling water domestic wastes oil stripper bottoms	0.40	None (Activated sludge in approx. 1975)	Savannah River
Continental Forest Industries	1960	Kraft Pulp Mill Effluent	0.50	Settling ponds, Spirit Creek aeration	
E. I. du Pont Nemours & Co.	1962	Inorganic caustics	?	Settling ponds	Butler Creek
Graniteville Mills, Sibley Division	1870	Dye, slasher, and domestic	.28	None	Water wheel tailrace to Savannah River
IMC (International Minerals & Chemicals	1908	Cooling water and scrubber wastewater	.002	None	Oates Creek
J. P. King Mfg. Co.	1881	Dye, slasher, and domestic	.03	None	Water wheel tailrace to Savannah River
Monsanto Company	1962	Cooling water, boiler blowdown,	.20	Cooling ditch	Ditch to Butler Creek
Olin Corp.	1964	Hg, chloride	2.5	Hg recovery and pH neutralization	Savannah River
Philadelphia Quartz Co.	?	Inorganic caustics	.008	Two holding ponds	Savannah River
Riverside Mills	?	Organic acidic	.30	None	City sewer to Third Level Canal
Scott Meat Packers, Inc.	1939	Blood, washwater	.12	None	Rocky Creek

TABLE 3 (Contd)

<u>Name of Industry</u>	<u>Const. Date</u>	<u>Type of Wastewater</u>	<u>Estimated Vol. of Wastewater (mgd)</u>	<u>Treatment Prior to 1970</u>	<u>Receiving Stream</u>
Shapiro Packing Co., Inc.	1940	Organic	?	Grease trap	Most of waste to sanitary sewers, small amount to Camille Street Ditch
Swift Fresh Meats Co.	1897	Organic	?	Floor grates	Second Level Canal
Taylor-Piedmont Co.	?	Creosote, oils	.90	None	Rocky Creek

TABLE 4

Partial List of the Savannah River Pollution Sources from the North Augusta and Aiken County Areas*

Name of Industry	Const. Date	Type of Wastewater	Estimated Vol. of Wastewater (mgd)	Treatment Prior to 1970**	Receiving Stream
Aiken STP (improved 1963)	1951	Domestic & industrial wastes	0.93	Secondary (Trickling filter)	Kelly Creek
N. Augusta Sewer Outfall	?	Domestic & industrial wastes	0.95	None	Savannah River
Bath Mill	1929	Domestic & industrial wastes	0.303	none	Horse Creek
Clearwater Finishing	1929	Finishing plant and domestic wastes	4.52	none	Little Horse
Graniteville Mill	1890	Finishing plant wastes, chromium	4.65	none	Horse Creek
Kimberly Clark	1968	Domestic & industrial wastes	6.89	Retention Lagoon	Savannah River
Seminole Mills	1924	Boiler blowdown, air conditioning, wastewater, domestic wastes	0.04	none	Little Horse Creek
J. P. Stevens Co.	1966	Domestic & industrial wastes, wool scouring wastes	0.38	none	Lower Three Runs Creek
Valchem Chemical Co.	1947	Domestic & industrial wastes, chromium, ammonia-N	1.61	none (?)	Horse Creek
Warrenville STP	1946	Domestic & industrial wastes	0.33	Secondary (Activated sludge)	Kelly Creek

* Reference 18.

** Most North Augusta and Aiken County industries currently direct their wastes to the Horse Creek Pollution Control Facility (constructed in 1979) for secondary treatment.

The second major dam is Hartwell Dam, which was completed in June, 1962. Hartwell Dam is located in the headwater region of the Savannah River about 65 miles (105 km) upstream from the Clarks Hill Dam. A third large dam, the Richard B. Russell Dam, is being built between the Clarks Hill and Hartwell Dams, but it will not be completed until 1984 or 1985. Several other small dams are located along the river, including three near Augusta: New Savannah Bluff Lock and Dam (completed in 1937), Stevens Creek Dam (completed in 1914), and Augusta City Dam (rebuilt in 1863).

A program that may have had considerable short-term implications on the surveys, was the dredging operations conducted by the U. S. Corps of Engineers. This program, initiated in October, 1958, was designed to dredge and maintain a 9 ft navigation channel in the Savannah River from the city of Savannah to Augusta.* A total of 61 sets of pile dikes were placed to constrict the river flow, thereby increasing flow rates, and a total of 37,645 linear ft of wood and stone revetment was laid to reduce erosion on banks opposite from the dikes. In addition, the channel was dredged, and 31 cutoffs were made, reducing the total river distance from Augusta to Savannah by about 15 miles (24.1 km). The project was completed in July, 1965; however, periodic dredging was continued to maintain the channel. A table of dredging dates and locations is included in Table 5.

The Savannah River Plant, which began construction in 1951, was another source of potential impact on the Savannah River. The major purpose of the Academy's surveys in the Savannah River was to monitor the effects of the Savannah River Plant. The next chapter will discuss the Savannah River Plant activities from a historical aspect to evaluate some of the possible sources of impact by the Savannah River Plant on the Savannah River.

CHAPTER IV. SAVANNAH RIVER PLANT ACTIVITIES

The Savannah River Plant (Figures 3-4) consists of a total of five nuclear production reactors and supporting facilities.^{20,21} Three of the reactors are currently operational (P, K, and C), and two are on stand-by (R and L). In addition, a small test reactor (U) is on stand-by. Nuclear fuel and targets are manufactured on the site in the fuel and target fabrication facility (M). The irradiated reactor products are processed and packaged for shipment in the chemical separations areas (F and H). Deuterium,

* Some dikes and revetments had been laid earlier, and were in place at the start of the Savannah River surveys in 1951.

TABLE 5

Savannah River Dredging Activities, 1964 through 1974
 (Partial Listing)*

<u>Dates</u>	<u>River Miles Dredged</u>
1964 Dec. 7-10	176.0 - 175.4
Dec. 10-12	175.2 - 174.8
Dec. 13	165.5
Dec. 14-17	158.4 - 159.0
Dec. 18-22	158.1 - 157.2
Dec. 23-30	141.3 - 140.8
Dec. 31	137.3 - 136.8
1965 Jan. 2-5	135.6 - 135.3
Jan. 5	129.2
Jan. 6	118.7 (Rt 301 Bridge)
1966 April 22-25	173.8**
April 26 - May 2	162.0**
May 3-4	151.0**
May 4-9	149.0**
May 9-10	144.0**
May 11-12	126.0**
May 12-13	124.5**
1967 June 3-5	197.9 - 179.8
June 6-16	179.8 - 175.0
June 17-22	175.0 - 174.3
June 22 - July 9	174.3 - 169.0
July 10 - Aug. 4	169.0 - 135.0
Aug. 4 - Aug. 10	135.0 - 134.4
Aug. 10	128.4
Aug. 11	59.4
1968 Sept. 8	above 167.1
Sept. 9	167.1
Sept. 24-25	151.3 - 151.1
Sept. 26	144.2 - 144.1
Sept. 27	136.5 - 136.3
Sept. 26 - Oct. 4	129.3 - 128.9
Oct. 5-6	124.6 - 124.0
Oct. 6-7	114.5 - 114.2

* Reference 16.

** Only average river mile values were available.

TABLE 5 (Contd)

<u>Dates</u>	<u>River Miles Dredged</u>
1970 - Aug. 21-27	179.7
	Aug. 28 - Sept. 9
	151.3
	Sept. 10-23
	150.8
	Sept. 24-29
	149.6 - 149.4
	Sept. 30 - Oct. 8
	149.3 - 149.0
	Oct. 9-14
1970 - Oct. 15-22	148.5 - 148.3
	146.6 - 146.2
	Oct. 23 - 29
	145.7 - 145.4
	Oct. 30 - Nov. 12
1970 - Nov. 13-19	128.4 - 128.2
	124.4 - 124.2
	Nov. 20
179.7 - 78.4	
1973 Oct 22 - 30	187**
	Oct. 31 - Nov. 2
	183.1**
	Nov. 3-6
	151.3**
	Nov. 7-24
	149.4**
	Nov. 25
	148**
	Nov. 26 - Dec. 1
146**	
1973 Dec. 2-4	144**
	Dec. 5-7
	136**
Dec. 8	
117**	

** Only average river mile values were available.

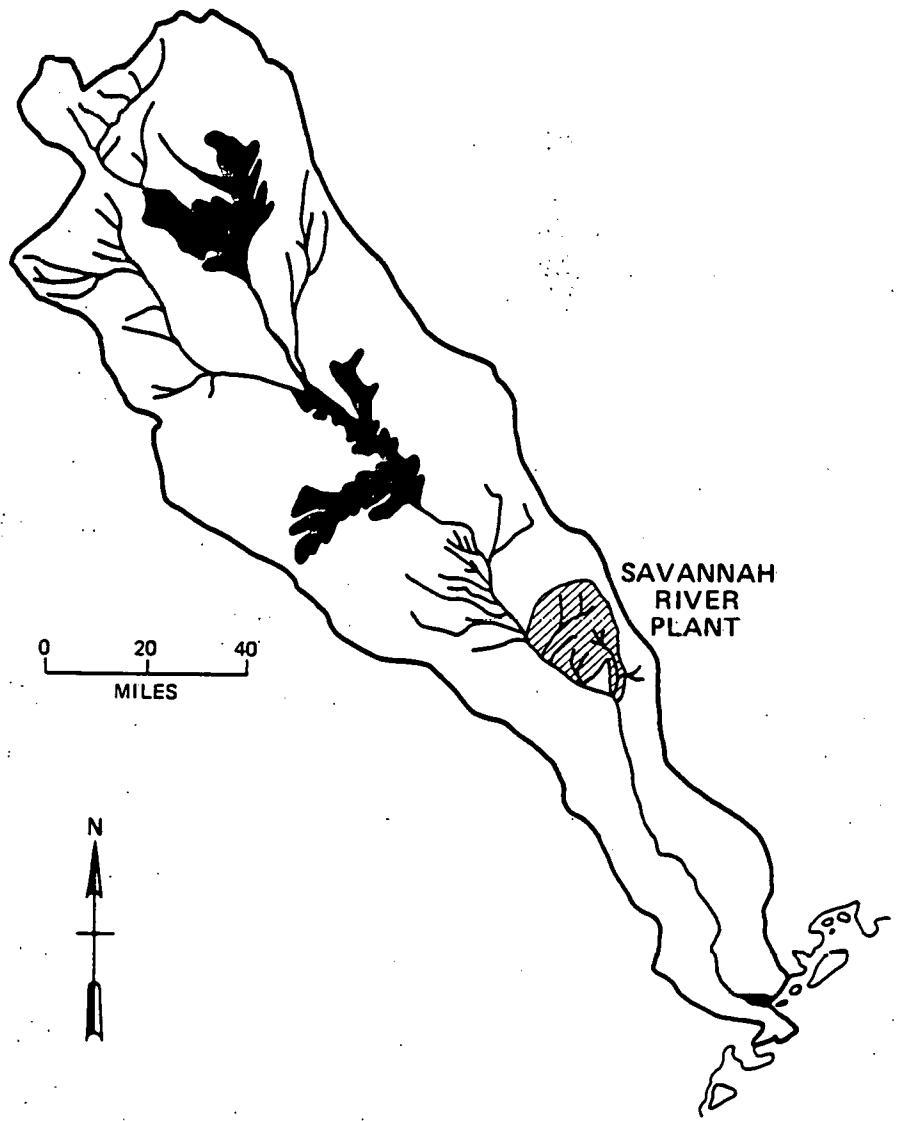


Figure 3. Location of the Savannah River Plant in the Savannah River Drainage Basin.

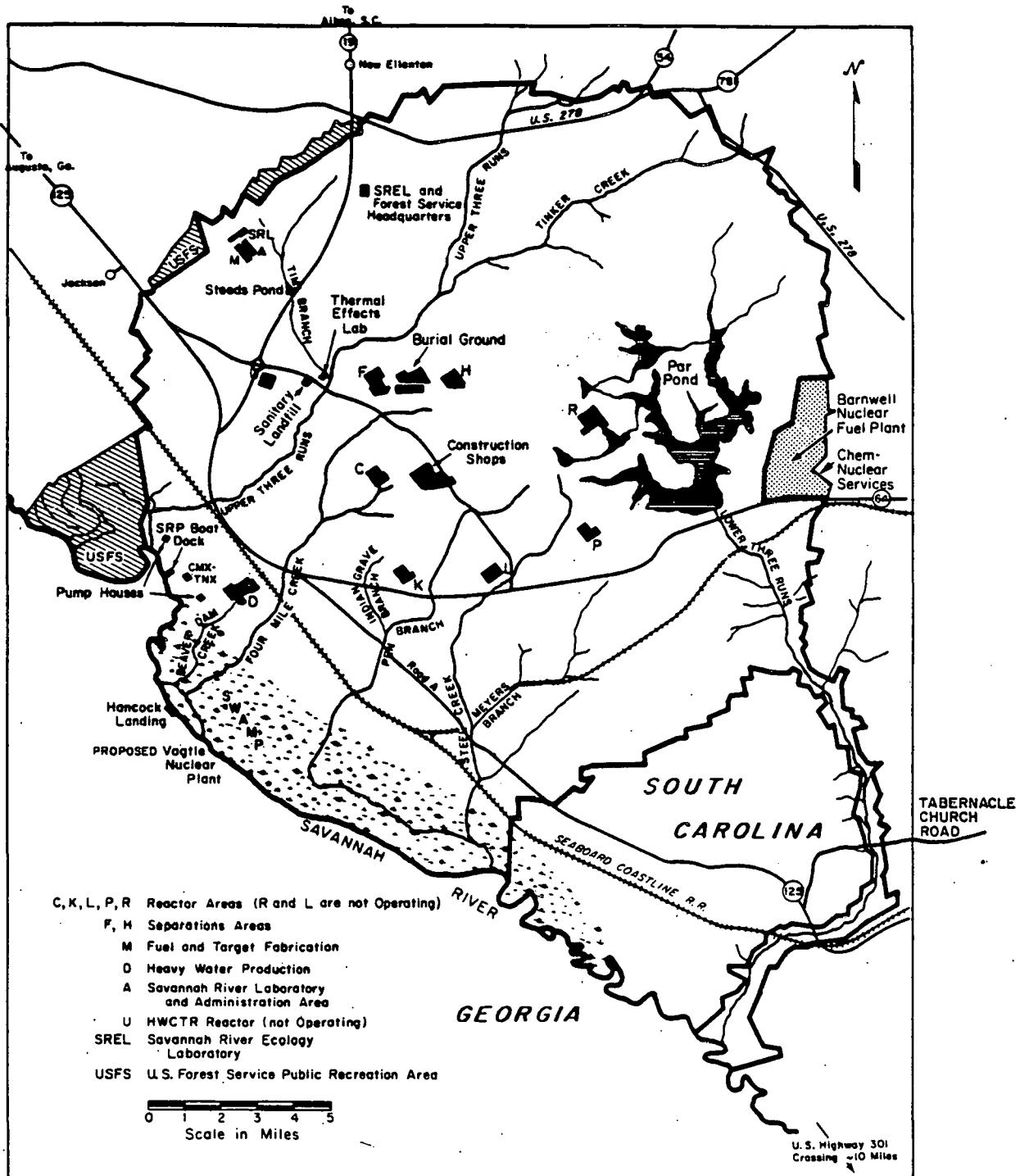


Figure 4. The Savannah River Plant Site.

which is used to moderate the neutron activity inside the reactor, is separated from Savannah River water in the heavy water extraction and recovery area (D). Other facilities include the Savannah River Laboratory, a process development laboratory, and administrative areas (A).

The major products produced at the Savannah River Plant are ^{239}Pu and ^3H . Smaller quantities of other nuclear materials have been produced. Some of these products include:

- 1) ^{238}Pu , a self-contained energy source used as a power supply in pacemakers and for the space program
- 2) ^{252}Cf , an active neutron source useful in radiography, activation analysis, and possible cancer therapy
- 3) ^{60}Co , a gamma ray source used in cancer therapy and self-contained power sources
- 4) ^{244}Cm , a potential power source
- 5) ^{233}U , a research material loaned to Universities and other research facilities.

The Savannah River Plant operations produce nuclear wastes, as well as a variety of industrial and domestic wastes. Most nuclear wastes are processed and stored on the site. Industrial and domestic wastes, and some radionuclides are processed if necessary, and released to drainage ditches and seepage basins. As part of a comprehensive program to monitor the effects of these wastes on the environment, the Academy of Natural Sciences of Philadelphia was contracted to evaluate the impact of the Savannah River Plant on the health of the Savannah River.

Effluents released to the Savannah River, or to seepage basins that drain into tributaries of the Savannah River, contain exceedingly low levels of radionuclides and other materials. Tables 6 through 9 list the major compounds found in Savannah River Plant effluents,²⁰ and show the effect of these compounds on chemical concentrations and bacterial numbers in the Savannah River.

One of the most important potential impacts that the Savannah River Plant has is from thermal effluents. Large quantities of river water are pumped through heat exchangers in the reactor buildings, then are returned to the river by way of Four Mile Creek, Beaver Dam Creek, and Pen Branch Creek (Figure 4). Steel Creek and Lower Three Runs Creek have also been used during the 1951-1976 survey period to transport thermal effluents. A summary of the history of reactor operations is shown in Table 10.

TABLE 6

Radioactivity in Savannah River Water (1975)*

<u>Sampling Point</u>	Alpha Emitters** (10^{-9} $\mu\text{Ci}/\text{cc}$)			Nonvolatile Beta Emitters** (10^{-9} $\mu\text{Ci}/\text{cc}$)		
	<u>Max</u>	<u>Min</u>	<u>Ave</u>	<u>Max</u>	<u>Min</u>	<u>Avg</u>
Above SRP, 1 mile upstream from Upper Three Runs	1.0	ND†	ND	6	ND	ND
Below SRP, 8 miles downstream from Lower Three Runs at Highway 301	1.5	ND	ND	7	ND	ND

* Reference 20.

** Tritium and small amounts of ^{137}Cs and ^{90}Sr are the only radionuclides of SRP origin detectable by routine monitoring in river water at the downstream location.

† ND = not detectable (less than sensitivity of analysis).
 Alpha emitter minimum sensitivity = $0.2 \times 10^{-9} \mu\text{Ci}/\text{cc}$
 Nonvolatile beta emitter minimum sensitivity = $4.0 \times 10^{-9} \mu\text{Ci}/\text{cc}$.

TABLE 7

Comparison of Changes in Savannah River Water Quality
with Drinking Water Standards (1975)^a(Assumes all materials discharged to streams reach
the Savannah River)

Material	Changes in Savannah River Concentrations Resulting From SRP Release (mg/L) ^b	Drinking Water Standard, (mg/L)
Sulfate	0.2	250
Chloride	0.05	250
Nitrate	0.01	10
Phosphate	3×10^{-3}	d
Barium	1.7×10^{-3}	1.0
Iron	4.9×10^{-4}	0.3
Boron	1.8×10^{-4}	1.0
Zinc	1.5×10^{-4}	5
Chromium	4×10^{-5}	0.05
Manganese	9×10^{-5}	0.05
Arsenic	1×10^{-5}	0.05
Mercury	9×10^{-8}	0.002
Copper	1.3×10^{-6}	1
Phenols	1.7×10^{-7}	0.001
Cyanide	c	0.2
Cadmium	c	0.01
Lead	c	0.05
Selenium	c	0.01
Silver	c	0.05

a. References 20, 22.

b. Assumes minimum flow of 6000 cfs. Concentrations downstream of SRP are all below the standards.

c. Not normally discharged to streams at SRP.

d. No drinking water standard in 1975.

TABLE 8

Pesticides Analyzed by USGS^a in Savannah River Sediment and Water Samples (1975).

	<u>River Sediment, µg/kg</u>	
	<u>Above Plant</u>	<u>Below Plant</u>
Aldrin ^b	d	d
Chlordane ^b	d	d
DDD ^b	1.0	3.2
DDE ^b	1.2	3.4
DDT ^b	1.3	4.1
Diazinon ^b	d	d
Dieldrin ^b	0.5	1.4
Endrin ^b	d	d
Ethion ^b	d	d
Ethyl-Parathion ^b	d	d
Ethyl-Trithion ^b	d	d
Heptachlor ^b	d	d
Heptachlor-Epox ^b	d	d
Lindane ^b	d	d
Malathion ^b	d	d
Methoxychlor ^c	d	d
Methyl-Parathion ^b	d	d
Methyl-Trithion ^b	d	d
PCB ^b	d	9.0
PCN ^b	d	d
Silvex ^b	d	d
Toxaphene ^b	d	d
2, 4-D ^b	d	d
2, 4-DP ^b	d	d
2, 4, 5-T ^b	d	d

a. Limit of sensitivity of each analysis is 0.1 µg/L of water and 1.0 µg/kg of sediment. Reference 20.

b. Water and sediment analyses.

c. Water analyses only.

d. Below limit of sensitivity.

TABLE 9

Fecal Coliform Bacteria Measurements (1975)*

	Weekly Samples, Colonies/100 mL		
	<u>Maximum</u>	<u>Mean (Geom.)</u>	<u>Minimum</u>
Savannah River			
Upstream from SRP Highway 301 Bridges	26,000 1,200	2,300 440	110 <10
Upper Three Runs			
SRP Road F	530	90	0
SRP Road A	1,400	120	20
400-D Area Effluent	8,600	830	0
Four Mile Creek - Road A	4,600	450	0
Pen Branch - Road A	6,600	80	0
Steel Creek - Road A	1,800	130	0
Lower Three Runs			
Tabernacle Church Road	4,400	100	10
Road A	3,400	350	10

* Reference 20. Data from the Academy of Natural Sciences of Philadelphia are included in Table B.4.23 (Appendix B) for comparison.

TABLE 10

Startup and Stand-by Dates of the Savannah River Plant Reactors

<u>Reactor</u>	<u>Startup</u>	<u>Stand-by</u>
R*	12/28/53	6/15/64
P**	2/20/54	
L†	7/02/54	2/18/68
K††	10/14/54	
C¶	3/28/55	

* Discharge to Lower Three Runs until 1958,
then discharge to Par Pond.

** Discharge to Steel Creek until 1963, then discharge
to Par Pond.

† Discharge to Steel Creek.

†† Discharge to Pen Branch Creek.

¶ Discharge to Four Mile Creek.

Typically, the thermal effluents from the Savannah River Plant area create plumes in the river which mix slowly with the rest of the river. Thermal profiles for the Savannah River below Four Mile Creek and Steel Creek (Pen Branch effluent) are shown in Tables 11 and 12. The maximum river temperature recorded below the Savannah River Plant after mixing was 29.4°C on August 25, 1959 (Table 13).

The Academy of Natural Sciences of Philadelphia has conducted 16 major surveys on the Savannah River to monitor the effluence of the Savannah River Plant. These surveys, and the Academy's monitoring program, are described in more detail in the next chapters.

CHAPTER V. CHRONOLOGY OF SURVEYS AND STUDIES

There have been a total of 16 major surveys conducted by the Academy of Natural Sciences of Philadelphia on the Savannah River from 1951 to 1976 (Table 14). These surveys have produced a comprehensive, taxonomic data base covering the entire span of the Savannah River Plant's operations. The biological and chemical data from these surveys have been used to evaluate the impact of thermal effluents from the Savannah River Plant reactors, particularly L reactor, on the Savannah River.¹⁰ Other comparisons between biological data and reactor activity may be made using one or more of the data formats discussed in Chapter VIII (Scope of the Savannah River Plant Data Base).

In addition to the comprehensive river surveys, diatometer data have been collected quarterly from 1953 to 1976 at Stations 1, 6, and AB. These data represent a semi-continuous biomonitoring program that may be used to supplement the major survey data. Chapter IX and Appendices A and B provide a summary of the biological and chemical data available for each survey.

The Savannah River has been influenced by many changes that have occurred during the 25 year sampling period. The most notable impacts resulted from dredging of the river, industrialization along the river banks, and construction of upstream dams. The impacts resulting from these operations are detailed in Chapters III, IV, and VI.

CHAPTER VI. SAMPLING STATIONS

A number of criteria were used to select the location of sampling stations on the Savannah River.¹⁻⁹ The Academy survey team wanted stations with comparable ecological habitats. The total area to be sampled was of less importance than the inclusion of a similar variety of habitats. In addition, the stations should have similar physical and chemical characteristics

TABLE 11

River Temperatures Near Four Mile Creek (Measured 4/10/72)*

Location**	Depth, ft	Temperatures Above Ambient at Various Distances† from Right Bank, °C					Air Temperature, °C
		75%	50%	37%	25%	12%	
I	1	0.1	0.0		0.2		18.4
	5	0.0	0.0		0.2		
	10	0.0	0.0		0.2		
			(at 9.5')		(at 10.5')		
II	1	0.1	0.2	0.7	7.8	13.8	
	2	0.1	0.2	0.6	5.8	13.9	
	3	0.1	0.2	0.6	3.4	14.0	
	5	0.0	0.2	0.6	1.0	14.0	
	8	0.0	0.2	0.6	0.9	(at 3.5')	
	10	0.0		0.5	0.8		
			(at 9.5')	(at 9.5')			
III	1	0.0	0.2	0.3	2.9	3.9	
	2	0.1	0.1	0.5	2.4	2.8	
	3	0.1	0.1	0.5	2.0	2.3	
	5	0.0	0.1	0.5	0.9	1.8	
		0.0	0.1	1.0	0.9	1.6	
		(at 11')	(at 9')	(at 8')	(at 7.5')	(at 8.5')	
IV	1	0.6		0.9	1.4	1.4	
	3	0.5		0.8	0.9	1.4	
	5	0.4		0.9	(at 2.5')		

* River flow measured at SRP boat docks: 7750 cfs
 River stage measured at SRP boat docks: 85.9 ft
 River ambient temperature measured at Location I: 14.6°C.
 Reference 20.

** Locations:

- I - Control, on Savannah River, 200 yards above Four Mile Creek mouth.
- II - On Savannah River, at downstream side of Four Mile Creek mouth.
- III - On Savannah River, 100 yards below Location II.
- IV - On Savannah River, 0.5 mile below Location III.

† Percent of distance across stream from right bank, facing upstream.

TABLE 12

River Temperatures Near Steel Creek (Measured 4/13/72)*

Location**	Depth, ft	Temperatures Above Ambient at Various Distances† from Right Bank, °C					Temperature, °C
		75%	50%	37%	25%	12%	
I	1	0.0	0.0		0.0		27.8
	5	0.0	0.0		0.0		
	10	0.0	0.0		0.0		
		0.0			0.0		
		(at 11.5')			(at 11')		
II	1	0.2	0.0	0.0	0.0	2.4	28.6
	2	0.2	0.0	0.0	1.2	3.0	
	3	0.1	0.0	0.0	1.1	3.0	
	5	0.1	0.0	0.0	1.1	1.4	
	10	0.1	0.0	0.0	0.3	1.1	
		(at 7')	(at 10.5')	0.0	0.1	0.4	
III	1	0.1	0.0	0.0	0.1	0.9	30.0
	2	0.1	0.0	0.0	0.0	1.2	
	3	0.1	0.0	0.0	1.1	1.2	
	5	0.0	0.0	0.0	0.1	1.1	
	10	0.0	0.0	0.0	0.1	0.8	
		(at 9.5')	0.0	0.0	0.1	0.8	
IV	1	0.4	0.1	0.2	0.5	0.9	28.9
	5	0.2	0.1	0.2	0.5	0.8	
		0.2	0.1	0.2	0.5	0.9	
		(at 7')	(at 8')	(at 8.5')	(at 10.5')	(at 12.6')	
V	1	0.5	0.6		0.7		28.9
	5	0.4	0.6		0.7		
	10	0.3	0.5		0.7		
		0.3	0.3		0.7		
		(at 23')	(at 20')		(at 13')		
VI	1	0.6	0.5		0.5		29.0
	5	0.5	0.4		0.5		
	10	0.5	0.4		0.5		
		0.5	0.4		0.5		
		(at 11')	(at 14')		(at 15.5')		

* River flow measured at SRP boat docks: 7450 cfs

River stage measured at SRP boat docks: 85.5 ft

River ambient temperature measured at Location I: 16.9°C. Reference 20.

** Locations:

I - Control, on Savannah River, above Steel Creek mouth.

II - On Savannah River, 20 ft below Steel Creek.

III - On Savannah River, 100 yards below Steel Creek.

IV - On Savannah River, 0.6 mile below Steel Creek.

V - On Savannah River, one mile below Location IV.

VI - On Savannah River, one mile below Location V.

† Percent of distance across stream from right bank, facing upstream.

TABLE 13

Maximum Temperature Increases Caused by the Savannah River Plant
in the Savannah River*

<u>Criterion</u>	<u>South Carolina Standard</u>	<u>Maximum SRP Value</u>
Maximum temperature below SRP after mixing	32.2°C (90°F)	29.4°C
Maximum temperature increase	2.8°C (5°F)	1.4°C
Maximum mixing zone (% of cross-sectional area)	25%	<20%
% of surface area	33-1/3%	<25%

* Reference 20. Calculated using classified information for two reactors discharging to the river, at minimum river flow.

TABLE 14

Chronology of Surveys Conducted by the Academy of Natural Sciences of Philadelphia on the Savannah River (1951-1976)

<u>Survey No.</u>	<u>Survey Dates</u>	<u>Reactors in Operation*</u>			
1	1951: June 25 - July 14	None			
2	Oct. 15 - Nov. 31	None			
3	1952: Jan. 9-31	None			
4	May 5-22	None			
5	1954: Aug. 15-30	R	P	L	
6	1955: Aug. 24 - Sept. 7	R**	P	L	K C**
7	1956: May 6-22	R	P	L	K C
8	1960: May 23 - June 4	R	P	L	K C
9	Aug. 31 - Sept. 15	R**	P	L	K C
10	1965: May 30 - June 8		P	K	C
11	Sept. 21-30		P	L	K C
12	1968: May 27 - June 5			K	C
13	Aug. 24 - Sept. 2		P	K	C
14	1972: May 22 - June		P	K	C
15	Sept. 13-22		P	K	C
16	1976: Aug. 10-16		P	K	C

* R reactor discharged into Lower Three Runs Creek until 1958 then discharged into Par Pond until it was shutdown in 1964;

P reactor discharged into Steel Creek until 1963, then discharged into Par Pond;

L reactor discharged into Steel Creek until it was placed on stand-by in 1968;

K reactor discharges into Pen Branch Creek;

C reactor discharges into Four Mile Creek

** Reactor up for part of survey.

including similar substrates, current velocities and river contours. There should be shallow water and beaches for collecting (particularly for seining fish) and the area should be workable during high and low water conditions.

The Savannah River stations were all located in slow-moving, meandering sections of the river. The river bed consisted mostly of sand, with mud and silt in the pools and backwater areas. There were no large rocks in the river bed, although there were some limestone outcroppings in the area. The largest of these was Blue Bluff, located about 1/4 mile (0.4 km) downstream from Four Mile Creek (Figure 5). A considerable amount of floating and partially submerged debris was present at all stations, which provided increased surface area in the shallow, photosynthetic region of the river. After the completion of Clarks Hill Dam in 1953 there were noticeable changes in the productivity of the Savannah River stations. These changes included increased algal, invertebrate and fish diversities, as discussed in Chapter III.

The major Savannah River stations were numbered 1, 3, 5 and 6, as shown in Figure 5. These stations were used for all comprehensive surveys since 1951. A descriptive summary of the Savannah River stations is given in Table 15.

Station 1 (Figure 6) was located on a straight portion of the river at Mile 160.7 (formerly Mile 175.1), and was above any possible discharge from the Savannah River Plant. Station 1 included all of the dikes on the left bank* and the opposite area on the right bank. The right bank was a gentle slope with over-hanging willows. A stone revetment had been built along part of the right bank, and a small stream entering at the downstream boundary. There were sandy beaches between the left bank dikes. At this point the river flowed almost due south.

Station 3 was ecologically very similar to Station 1. It was located on a straight section of the river at Mile 143.9 (formerly 157.5), about 6 miles (9.6 km) downstream from Four Mile Creek (Figure 7). There were dikes with sandy beaches along the left bank, and the right bank was moderately steep and overhung with willows. There was a stone revetment near the right downstream boundary, and a small stream entered along the right bank.

Station 5 was originally located on a bend in the river at Mile 149.5 (Figure 8). Despite superficial differences from Stations 1 and 3, the ecological habitats were quite similar at Station 5. Sandy beaches and a considerable amount of fallen debris at Station 5 provided habitats similar to the sandy, shallow areas between dikes at Stations 1 and 3.

* Bank sides were determined by facing downstream.

TABLE 15

**Descriptive Summary of Stations Used by the Academy of Natural Sciences
of Philadelphia for Surveys on the Savannah River**

<u>Station</u>	<u>Location</u>	<u>Survey Numbers</u>	<u>Description</u>
1	Mile 160.7 (old 175.1)	1-16	Dikes and sandy beaches on L. bank. Gentle slope with stone revetment and small stream on R. bank. Overhanging willows, straight section of river.
3	Mile 143.9 (old 157.5)	1-16	Dikes and sandy beaches or L. bank. Fairly steep R. bank with stone revetment and small stream. Overhanging willows, straight section of river.
5 (old)	Old Mile 149.5	1-7	Sandy beaches on both banks with small stream on L. bank. Much debris. Curved section of river, with steep outer banks.
5 (new)	Mile 135.8 (old 149.5)	8-16	Similar to Old Station 5 Sandy beaches with small stream on L. bank. Much debris. Curved section of river with steep outer banks.
6	Mile 123.1 (old 134.5)	1-16	Sandy beaches on curve of river. Slough on R. bank designated Station 6B as opposed to river proper at 6A. Much debris.

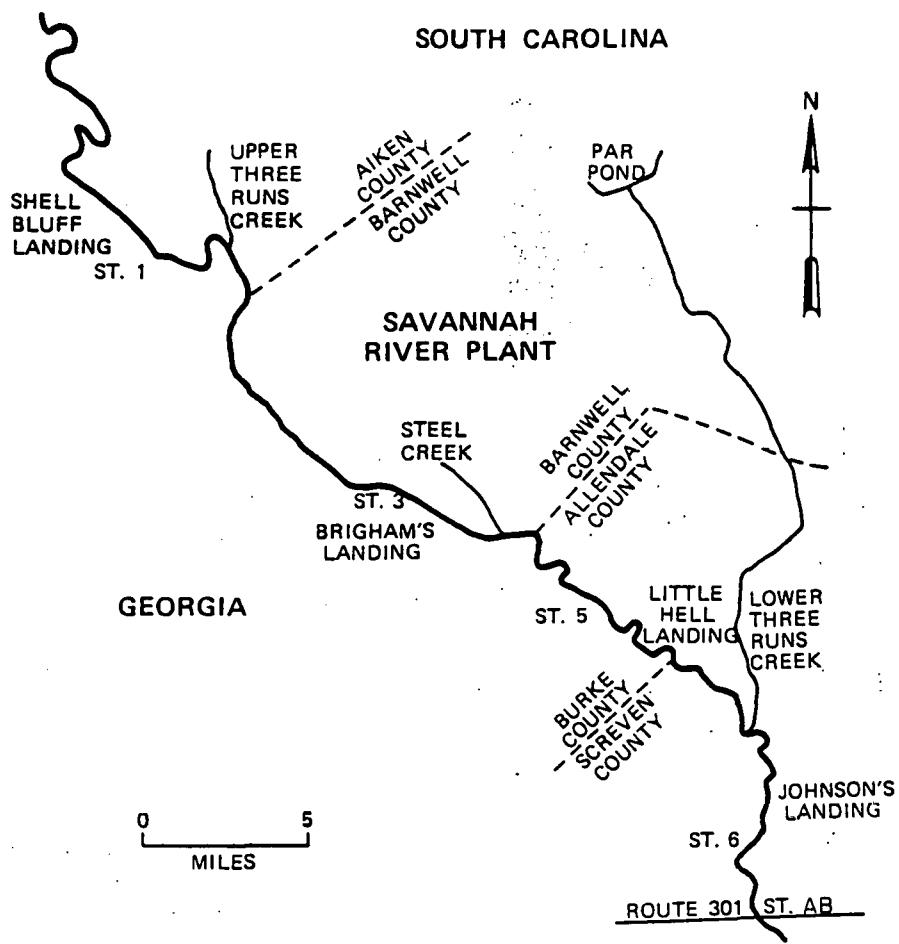


Figure 5. Survey Stations Used By the Academy of Natural Sciences of Philadelphia on the Savannah River (1951-1976).

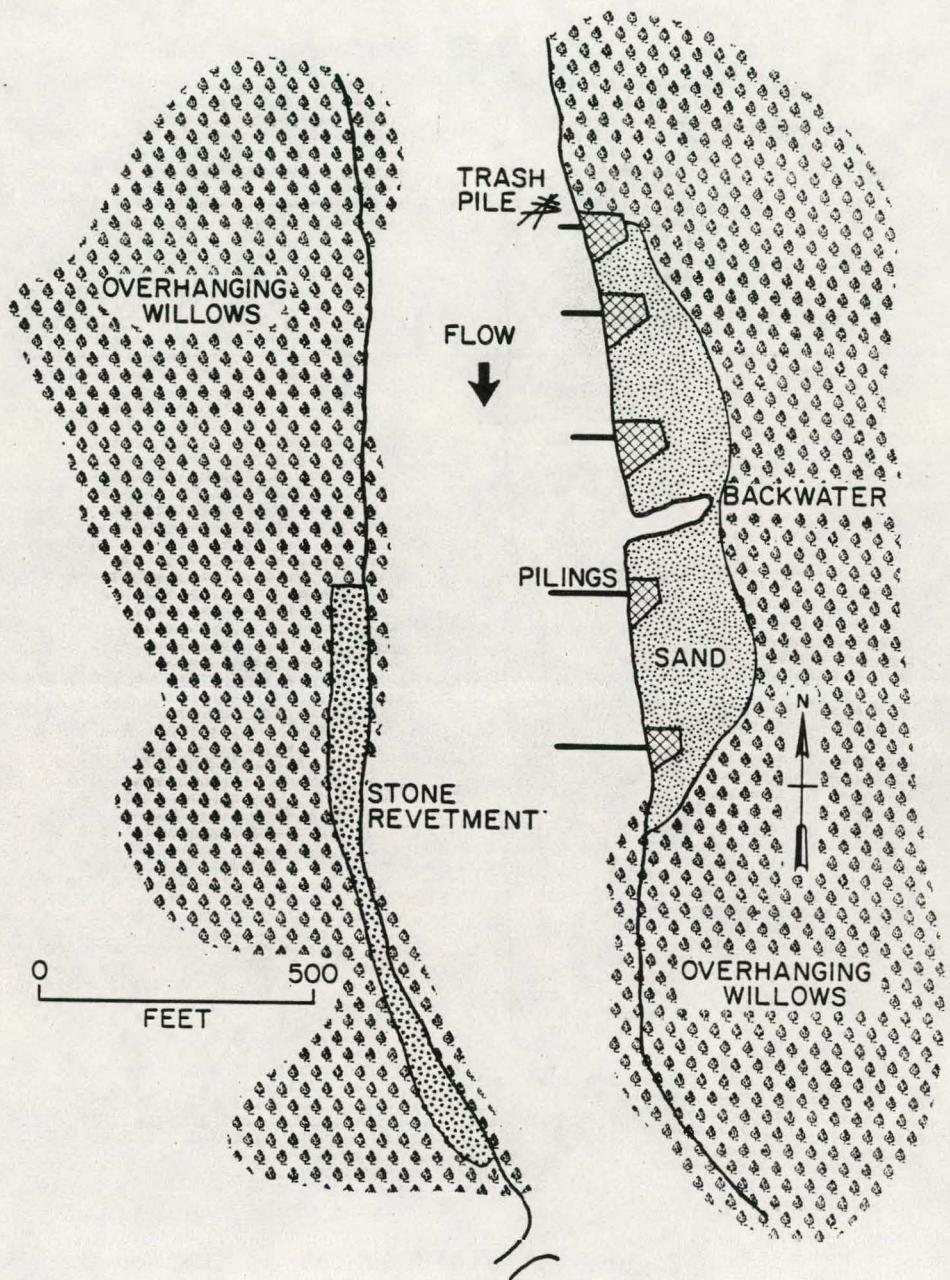


Figure 6. Field Sketch of Station 1.

References 1-9.

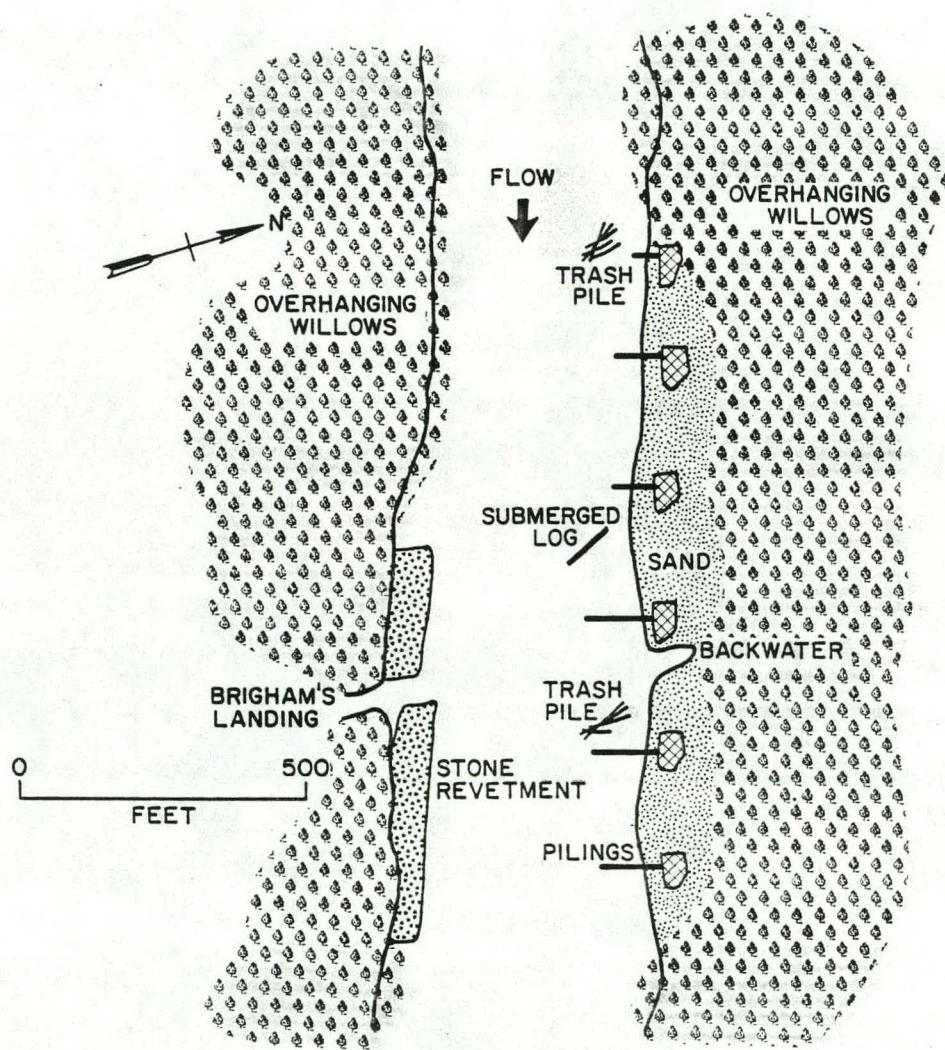


Figure 7. Field Sketch of Station 3.

References 1-9.

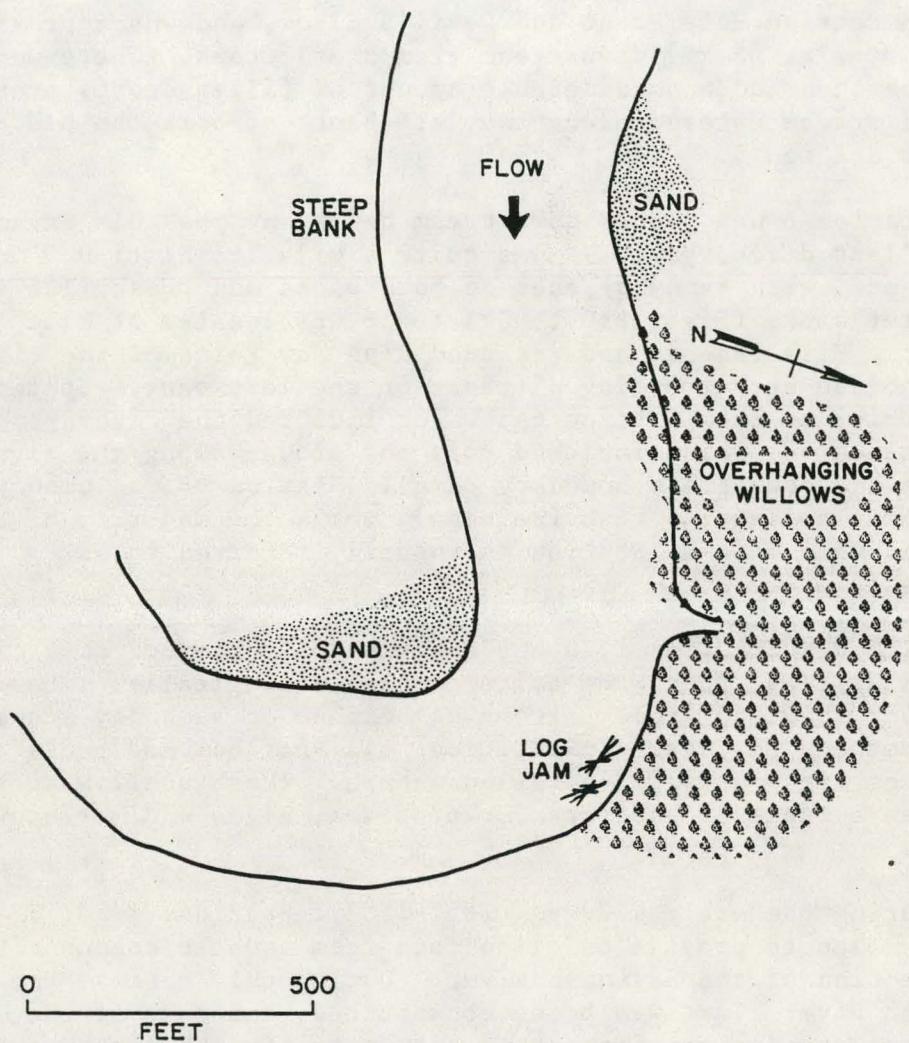


Figure 8. Field Sketch of Original Station 5.

References 1, 2.

In 1958 a dredging project was initiated by the U. S. Corps of Engineers. This project altered the river channel and caused Station 5 to become an oxbow. Station 5 was relocated to Mile 135.8 (formerly 149.5) for the 1960 surveys, and for the subsequent surveys. The new Station 5 was so similar in appearance to old Station 5 that the two were difficult to distinguish without a map. New Station 5 (Figure 9) was located on a bend in the river between Catfish Hole Point and Devil's Elbow, and was approximately 5 miles (8 km) downstream from Steel Creek. There were sandy beaches and a considerable amount of fallen debris present. A small stream entered along the left banks of both the old and the new Station 5.

Station 6 was placed downstream below any possible Savannah River Plant discharges. It was quite similar to Station 5 in appearance, with sandy beaches on both banks and trashpiles along the outer banks (Figure 10). Station 6 was located at Mile 123.1 (formerly Mile 134.5), and included Ring Jaw Point on the right bank and the corresponding distance on the left bank. Station 6 was subdivided into Station 6A, which included the river proper, and Station 6B, which included only the slough along the right bank at the downstream boundary of 6A. Station 6B was used primarily for collecting fish (seining), while the majority of collections were made at Station 6A (usually referred to as Station 6).

Although Stations 1, 3, 5, and 6 had a number of physical dissimilarities, they were quite similar ecologically. They all had sandy, shallow areas, either as beaches or as sandy accumulations between pilings. In addition, all stations had pools and backwater areas as well as fallen debris. These shallow areas provided a range of habitats for protozoa, algae and macroinvertebrates.

During the early surveys (1951-1952), Stations 1, 3, 5, and 6 were sampled to provide baseline data from aquatic communities in this section of the Savannah River. During this period, the Savannah River Plant was being constructed, and none of the reactors were discharging heated effluent into the survey area. After 1954, Station 5 received heated effluent from P and L reactors by way of Steel Creek, and indirectly from K reactor which discharged into Pen Branch, a tributary of Steel Creek. Station 3 received effluent from C reactor by way of Four Mile Creek. The effluent from P reactor was redirected into Par Pond in 1963, and L reactor was placed on stand-by in 1968. Reactors C and K are still in operation.

During the period that K, L, and C reactors were all operational, the temperature increase observed in the Savannah River caused by Savannah River Plant operations was usually no

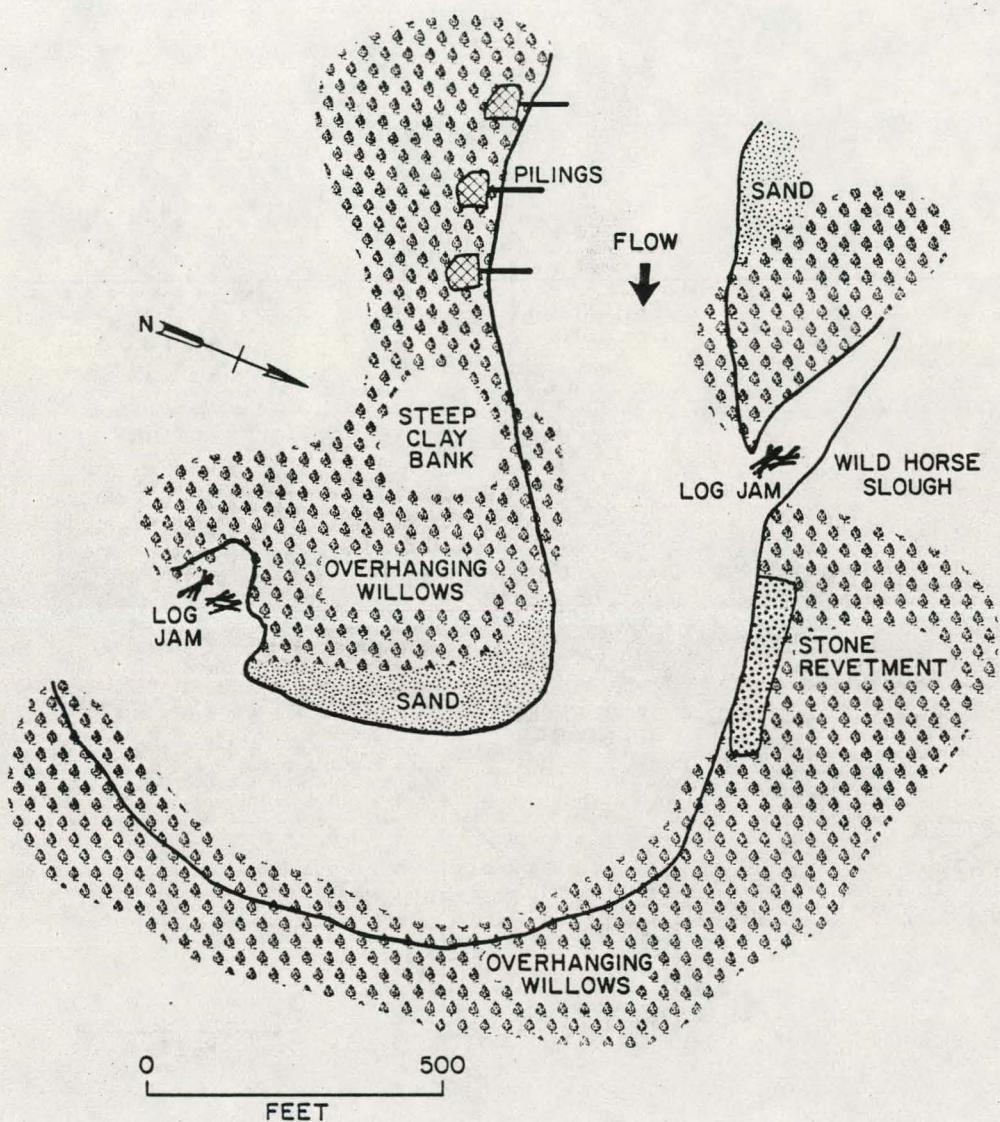


Figure 9. Field Sketch of Relocated Station 5.

References 3 through 9.

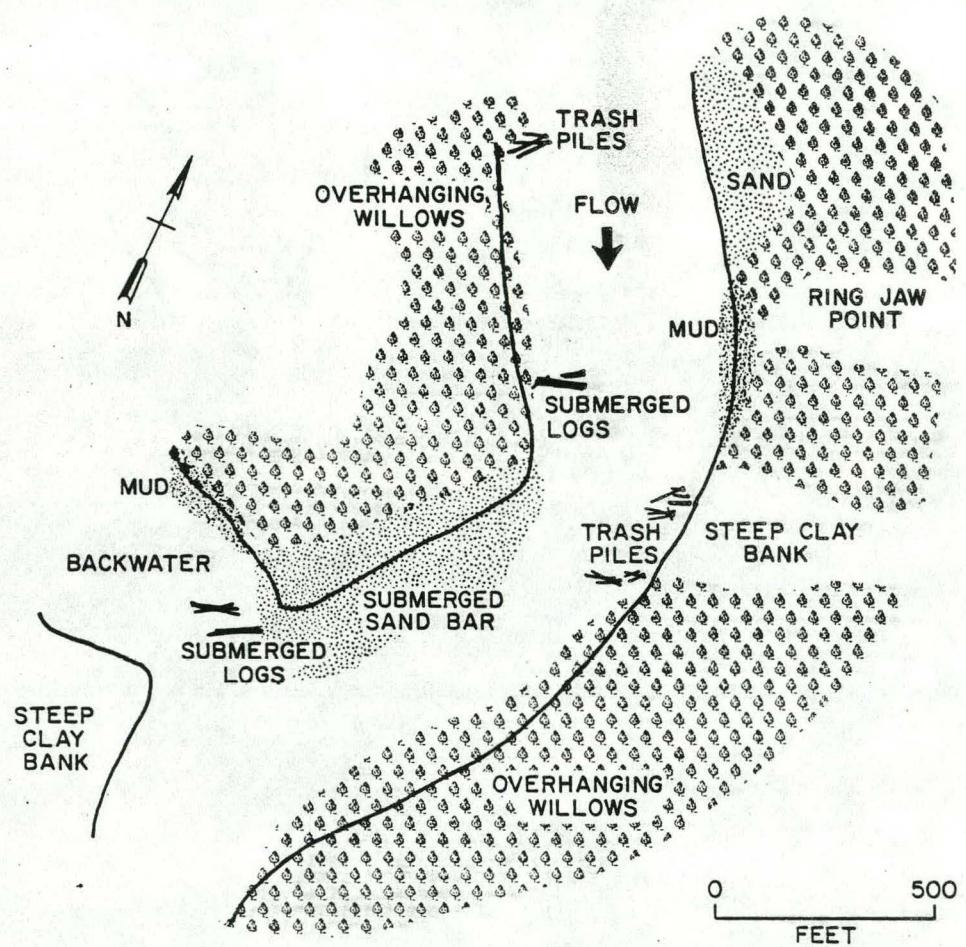


Figure 10. Field Sketch of Station 6.

References 1 through 9.

greater than 5°C (2.8°C; Table 13, Chapter IV). The thermal plume from Steel Creek was obliterated when the river passed a meander. The water temperatures at both Stations 3 and 5 were essentially homogeneous.

One additional station, Station AB, was located on the Savannah River, and was used for quarterly diatom surveys. This station was located at Mile 118.7, upstream from the Allendale Bridge on Rt. 301. Station AB was used as a reference station, and was situated well downstream from any Savannah River Plant thermal sources. The station was ecologically similar to Stations 1, 3, 5 and 6, with debris along the left bank, and overhanging vegetation along the right bank. Station AB was not used for any of the comprehensive river surveys, and field sketches of the station were not provided in any of the Academy survey publications.

CHAPTER VI. METHODS

The Academy of Natural Sciences of Philadelphia attempted to sample all river habitats qualitatively during the major river surveys to insure that most of the biota in the river were collected on each survey. Survey field teams usually consisted of a phycologist, an entomologist, an ichthyologist, a water chemist, a protozoologist, an invertebrate zoologist, and field assistants. Except where noted, the collecting, preservation and identification techniques were consistent for all major river surveys.

Much of the sampling was done by boats and, because of the size of the sampling area, took several weeks to complete. The first samples collected at each station were water samples for physical, chemical, and bacterial analyses. The methods for these analyses are summarized in Table 16. The station averages for each water chemistry measurement are given by survey number in Appendix B.4.

After the water samples were collected, several days were spent at each station sampling the major biological communities and populations in the river. Collecting was initiated at the downstream boundary of each station and proceeded upstream. All organisms, except protozoa, were sorted and preserved in the field, then transported to the Academy's laboratories for identification and incorporation into permanent collections. Protozoa were identified in the field laboratory because they cannot be preserved satisfactorily.

The river communities and populations were sampled qualitatively from a wide variety of habitats. Separate species lists were developed for algae and diatoms, insects, microinvertebrates,

TABLE 16

Summary of Methods used for Physical, Chemical, and Bacteriological Analyses of Savannah River Water Samples*

<u>Measurement</u>	<u>Method</u>
Alkalinity	
a) Methyl Orange	a) Methyl orange titration
b) Phenolphthalein	b) Phenolphthalein titration
c) Bicarbonate	c) Calculated from M.O. & P Alkalinitities
d) Carbonate	d) Calculated from M.O. & P Alkalinitities
Bacteriological	
a) Total bacteria	a) Membrane filtration + M-TGE broth incubation
c) Coliform	b) Membrane filtration + M-endo broth incubation
Bicarbonate	Calculated from Alkalinity
Carbon Dioxide	Calculated from pH and Alkalinity
Chlorides	Mohr titration, Silver Nitrate/Potassium Chromate titration
Conductivity	Specific conductivity meter (Cond. bridge)
Hardness	
a) Total	a) Compleximetric or EDTA titration, procedure and reagents from Betz & Co., Philadelphia, Pennsylvania, or calculated from Atomic Absorpton data
b) Calcium	b) C.4 Calcium hardness method, Betz & Co. reagents and procedure or calculated from A. A. data
c) Magnesium	c) Betz & Co. reagents and procedure or calculated from A. A. data
Metals	
a) Calcium	a) Measured on Perkin Elmer Model 303 Atomic Absorption Spectrophotometer or calculated from Hardness data.
b) Iron	b) Phenanthroline method or measured on A. A.
c) Magnesium	c) Calculated from Total and Calcium Hardness, or measured on A. A.
d) Manganese	d) Measured on A. A.
e) Potassium	e) Measured on A. A.
f) Sodium	f) Measured on A. A.

* All methods are from Standard Methods²³⁻²⁶ unless indicated.

TABLE 16 (Contd)

<u>Measurement</u>	<u>Method</u>
Nitrogen	
a) Ammonia	a) Direct Nesslerization method, automated Phenate method ²⁷ or measured with an Orion Ammonia electrode
b) Kjeldahl	b) Kjeldahl method or measured with a Technicon Auto Analyzer II
c) Nitrate	c) Phenoldesulphonic acid method, Cadmium Reduction Auto Analyzer ²⁷ and Spectrophotometric Brucine method
d) Nitrite	d) Diazotization method, and Diazotization using Auto Analyzer ²⁷
Oxygen	
a) Biological Oxygen Demand	a) 5-day B.O.D. Test
b) Chemical Oxygen Demand	b) Microdichromate Reflux Method ²⁸
c) Dissolved Oxygen	c) Azide modification of the Winkler test
pH	La Motte Co., Baltimore, Maryland pH Kit or Orion 401 pH Meter
Phosphate	
a) Total	a) Persulfate digestion/Ascorbic acid method
b) Ortho	b) Colorimetric Stannous Chloride method, Ascorbic Acid method, and Auto Analyzer (Ascorbic Acid) Method ²⁷
Silica	Colorimeters silicomolybdate method (intermediate range) and Auto Analyzer (Molybdate - Reactive Silica)
Solids (Residue)	
a) Total	a) Filtration/gravimetric method
b) Dissolved	b) Filtration/gravimetric method
c) Fixed	c) Filtration/gravimetric method
d) Suspended	d) Filtration/gravimetric method
e) Volatile	e) Filtration/gravimetric method
Sulfate	Turbidimetric method
Temperature	Field measurement using mercury-filled glass thermometer
Transparency	Black and white Secchi disc
Turbidity	Jackson Candle method, photometric measurement at 420 μm using standard suspension from Harleco Co., Philadelphia Pennsylvania, or Hach 2100 Turbidimeter, Standardized with formazin suspension

protozoa, and fish. A summary of these data averaged for each station by survey number, are presented in Appendix B.1.

Algae were sampled from shallow and deep water, logs and twigs, stones, moss and rootlets, soft mud, and any other substrate present inside the station boundaries. All samples were placed in 17 ml vials or small jars and examined in the laboratory to separate nonliving material from the sample. The samples were preserved with formalin, ethanol or Transeau's Fixative (6 parts water: 3 parts 95% ethanol: 1 part formalin). Diatoms were cleared and washed prior to examination. All organisms were identified to species or taxa. Diatom taxa represented by fewer than six specimens were not considered to be established in the samples, and were excluded from the species lists.

Diatoms were also collected from glass slide substrates that had been suspended for two weeks at Stations 1, 6, and AB using Catherwood Diatometers.²⁹ The diatometers were placed near the right and left banks at each station, and collections were made biweekly by Savannah River Plant personnel. The colonized slides were mailed to the Academy for processing and quantitative species identifications. Complete quantitative counts were made quarterly on representative slides from the biweekly samples. The diatom data in the Savannah River Laboratory data base are segregated into Diatom data (from quarterly quantitatives counts) and Diatom Taxonomy Data (from qualitative major river surveys).

Protozoa were collected using an aspirator bottle or a plastic tube connected to a rubber bulb. Approximately 15 to 20 half-pint jars were collected at each station. Samples were kept in an insulated container or wet bag, and were transported to the laboratory within one hour to avoid loss of sensitive species. At the laboratory the samples were placed near a light source and allowed to settle. Subsamples were taken from the miniscus and surface of the sediments near and away from the light source. Species were identified with the aid of conventional stains or by using a Phase Contrast microscope, which eliminated the necessity of stains. Only species represented by 6 or more specimens per slide, or species identified in 3 out of 4 subsamples were considered to be established and were included in the species lists.

Macroinvertebrates were collected from a variety of habitats, including sand bars, clay banks, mud flats, stone revetments, tributary mouths, pilings, floating debris and snags. Slow moving and sedentary forms were collected by hand, while other forms were collected using dip nets, dredges, mesh screens and bottom scrapers (Needham Scraper). Some specimens were placed directly in vials with alcohol or formalin. Substrate and plant materials were taken to the laboratory for sorting under a dissecting microscope.

Insects were placed in vials containing 50 percent alcohol, then transferred to 70 percent alcohol for storage. Tendipedid larvae were mounted directly on microscope slides. Arthropods, annelids and poriferans were preserved in 70 percent alcohol. All other macroinvertebrates were collected alive, then narcotized to prevent excessive contraction in death, before being fixed and preserved in alcohol.

Fish were collected by a number of methods. Initially, seines were used in backwater areas, along with hoop nets, gill nets, wire traps and hook and line fishing techniques. After 1952, rotenone was also used for collecting fish. Potassium permanganate was applied after the collections were made to detoxify the rotenone and to restrict its effects to the desired area. Fish were preserved with 10 percent formalin and transferred to 40 percent isopropyl alcohol for storage.

CHAPTER VIII. SCOPE OF THE SAVANNAH RIVER LABORATORY DATA BASE

The Savannah River Laboratory version of the 25-year data base is stored on IBM 370/VS direct access devices, using local FORTRAN/VSAM interface routines. It is expected, therefore, that parties interested in analyzing these data will require the assistance of Savannah River Laboratory Computer Science personnel in retrieving and reformatting selected data items. This chapter will describe the content of the data base. Appendix A alphabetically lists the taxa for which data are available. Appendix B gives an overview of the Savannah River Survey results in conveniently available formats.

There are four major data sets in the data base:

- 1) A taxonomic nomenclature coding structure;
- 2) Presence/absence data, organized by
 - Each Station and Date for all Taxa
 - Each Taxon for all Stations and Dates;
- 3) Diatometer Taxa frequency counts, organized by
 - Each Station and Date for all Taxa
 - Each Taxon for all Stations and Dates;
- 4) Water Quality physical and chemical species concentrations, organized by
 - Each Station and Date for all species
 - Each species for all Stations and Dates.

Each of these data sets are described in the following subsections.

A. Savannah River Taxonomy

The Savannah River taxonomic coding structure uses three distinct labels for each item of nomenclature:

- 1) An Accession Number identifies each specific class of organism. This unique number is never changed, and is treated in all computer programs as a surrogate for the organism's taxonomic identity. The Accession Number is maintained as a non-reuseable JSS Key Access Number (KAN).
- 2) Each Accession Number (KAN) is correlated to appropriate biological taxonomic nomenclature from the Savannah River data. This is the data portion of the KAN record in the Taxonomic Data Set.
- 3) Taxonomic data are placed into a classification hierarchy from Phylum to Forma, comprised of 18 possible levels (Table 17).

The Key portion of the JSS taxonomic data set is comprised of the 18 hierarchical levels concatenated into a 72 byte Key. Unused levels, and levels below the one being defined, are zero filled. The Key field values may be changed to alter the natural presentation order of taxonomy, but such changes do not alter the assigned KAN. For example, the organism "Alasmidonta arcula (Lea)" with the following taxonomy:

Phylum	Mollusca
Class	Bivalvia
Subclass	Paleoheterodonta
Order	Unionoida
Superfamily	Unionacea
Family	Unionidae
Subfamily	Anodontinae

might be represented by a Key of:

phy. subp cls. subc ord. subo grp. ser. supf fam. subf
0140 0000 0200 0100 0100 0000 0000 0100 0100 0100
trb. grp. gen. subg spc. subs frm. (not used)
0000 0000 0100 0000 0010 0000 0000 0000 0000

where numeric codes have been recorded in corresponding hierarchical positions.

In short, the Accession Number creates a computer identity for each organism in the Savannah River. As we alter the taxonomic

TABLE 17

**Taxonomic Hierarchy used for the Classification of the Savannah
River Biological Data**

<u>Taxonomic Level</u>	<u>Key Positions</u>
Phylum/Division	1-4
Subphylum/Subdivision	5-8
Class	9-12
Subclass	13-16
Order	17-20
Ssborder	21-24
Group	25-28
Series	29-32
Superfamily	33-36
Family	37-40
Subfamily	41-44
Tribe	45-48
Group	49-52
Genus	53-56
Subgenus	57-60
Species	61-64
Subspecies/Variety	65-68
Forma	69-72

cross references for the computer's Accession Numbers, the printed names for organisms may change; however, their computer identity will not. This system allows the taxonomic nomenclature to be updated when future changes or revisions are necessary to maintain a current and accurate taxonomy. A listing of the taxonomic accession numbers is included in Appendix A. Whenever possible, the accession number should be used when requesting data for a particular organism or group of organisms.

The taxonomic data may be formatted in a variety of ways. A complete listing of the organisms identified from the Savannah River during the entire survey period is extensive. This lengthy list may be condensed into similar orders, classes or phyla, for easier interpretation. One example of such a condensation is presented in Appendix B.1. Here the data are grouped into major biological subsets, and graphed by station to show the total number of species identified for each survey.

B. Presence/Absence Data

The Presence/Absence data are binary values associated with each accession number, at each of up to four stations, at each of the 16 survey dates. An example of one possible Presence/Absence data format is given in Appendix B.2.

The surveys and stations for which data are available are as follows:

1951: spring	1,3,5,6	1968: spring	1,3,5,6
fall	1,3,5,6	fall	1,3,5,6
1952: spring	1,3,5,6	1972: spring	1,3,5,6
fall	1,3,5,6	fall	1,3,5,6
1954: fall	1, 6	1976: fall	1,3,5,6
1955: fall	1,3,5,6		
1956: spring	1,3,5,6		
1960: spring	1,3,5,6		
fall	1,3,5,6		
1965: spring	1,3,5,6		
fall	1,3,5,6		

C. Diatometer Data

The Diatometer data are recorded as counted frequency values associated with each accession number, at each of up to four stations, for each of four seasons, in each of the 24 study years. These data are different from the diatom taxonomy data, which include only species listings. The diatometer data may be formatted in a number of different ways. One example is presented in Appendix B.3.

The studies and diatometer numbers* for which data are available are as follows:

1953: fall	2,4	1962: winter	2,4,8	1970: winter	2,4,8
1954: winter	2,4	spring	2,4,8	spring	2,4,8
spring	2,3	summer	2,4,8	summer	2,4,8
summer	1,3	fall	2,4,8	fall	2,4,8
fall	2,3				
1955: winter	2,3	1963: winter	2,4,8	1971: winter	2,4,8
spring	1,3	spring	2,4,8	spring	2,4,8
summer	2,3	summer	2,4,8	summer	2,4,8
fall	2,4	fall	2,4,8	fall	2,4,8
1955: winter	2,3	1963: winter	2,4,8	1971: winter	2,4,8
spring	1,3	spring	2,4,8	spring	2,4,8
summer	2,3	summer	2,4,8	summer	2,4,8
fall	2,4	fall	2,4,8	fall	2,4,8
1956: winter	2,4	1964: winter	1,4,8	1972: winter	2,4,8
spring	2,4	spring	2,4,8	spring	2,4,8
summer	2,4	summer	2,4,8	summer	2,4,8
fall	2,4	fall	2,4,8	fall	2,3,8
1957: winter	2,4	1965: winter	2,4,9	1973: winter	2,4,8
spring	1,4	spring	2,4,9	spring	2,4,9
summer	2,4	summer	2,4,8	summer	2,4,8
fall	1,4	fall	2,4,8	fall	2,4,8
1958: winter	2,4	1966: winter	2,4,8	1974: winter	2,4,8
spring	1,4	spring	1,4,8	spring	2,4,9
summer	2,4	summer	2,4,8	summer	2,4,8
fall	2,4	fall	2,4,8	fall	2,4,8
1959: winter	2,4,8	1967: winter	2,4,8	1975: winter	2,4,8
spring	2,4,8	spring	2,4,9	spring	2,4,8
summer	2,3,8	summer	2,4,8	summer	2,4,8
fall	2,4,8	fall	2,4,8	fall	2,4,8
1960: winter	2,3,8	1968: winter	2,4,9	1976: winter	2,4,8
spring	2,4,8	spring	2,4,9	spring	2,4,8
summer	2,4,8	summer	2,4,8	summer	2,4,8
fall	1,4,8	fall	2,4,8	fall	2,4,8
1961 winter	2,4,8	1969: winter	2,4,8		
spring	1,4,8	spring	2,4,8		
summer	2,4,8	summer	2,4,8		
fall	2,4,8	fall	2,4,8		

* Two diatometers were placed at each station:

Station 1: Diatometer 1 near the left bank
 Diatometer 2 near the right bank

Station 6: Diatometer 3 near the left bank
 Diatometer 4 near the right bank

Station AB: Diatometer 8 near the left bank
 Diatometer 9 near the right bank

D. Chemical Data

The chemical data are real values of the concentration of the following suite of water-quality parameters:

	(abbreviation)
Alkalinity, Methyl Orange (mg/L as CaCO ₃)	ALKAMO
Alkalinity, Phenylthphlein (mg/L as CaCO ₃)	ALKALP
Alkalinity, Carbonate (mg/L)	CO3
Alkalinity, Bicarbonate (mg/L)	HCO3
Bacteria, Coliform Count (col/100 mL)	COLCTN
Bacteria, MPN (cell/100 mL)	MPNCTN
Bacteria, Plate Count (col/mL)	PLACTN
Bacteria, Total Count (col/mL)	TOTCTN
Bacteria, Viable Count (col/mL)	VIACTN
Carbon Dioxide (mg/L)	CO2
Dissolved Oxygen (mg/L)	DO
Chloride (mg/L)	CL
Specific Conductivity (μmho at 25°C)	SPCOND
Hardness, Total (mg/L as CaCO ₃)	HARD
Hardness, Ca (mg/L as CaCO ₃)	CACO3
Hardness, Mg (mg/L as CaCO ₃)	MGC03
Calcium (mg/L)	CA
Iron (mg/L)	FE
Magnesium (mg/L)	MG
Manganese (mg/L)	MN
Potassium (mg/L)	K
Sodium (mg/L)	NA
Ammonia (mg/L)	NH3-N
Total Kjeldahl Nitrogen (mg/L)	TKN
Nitrate (mg/L)	NO3-N
Nitrite (mg/L)	NO2-N
Biological Oxygen Demand (mg/L)	BOD
Chemical Oxygen Demand (mg/L)	COD
Total Phosphorus (mg/L)	TOTP
Ortho-Phosphorus (mg/L)	PO4-P
Silica (mg/L)	SIO2
Total Solids (mg/L)	TS
Total Dissolved Solids (mg/L)	TDS
Fixed Solids (mg/L)	FIXR
Suspended Solids (mg/L)	SS
Volatile Solids (mg/L)	VS
pH	PH
Temperature (°C)	TEMP
Sulfate (mg/L)	SO4
Transparency (in.)	CLEAR
Turbidity (mg/L)	TURB

The data are available at up to each of four stations in each of the 16 survey years. Examples of Chemical data are presented in Appendix B.4. These examples indicate two ways the data may be formatted.

The studies and stations for which data are available are as follows:

1951:	spring	1,3,5,6
	fall	1,3,5,6
1952:	spring	1,3,5,6
	fall	1,3,5,6
1955:	fall	1,3,5,6
1956:	spring	1,3,5,6
1960:	spring	1,3,5,6
	fall	1,3,5,6
1965:	spring	1,3,5,6
	fall	1,3,5,6
1968:	spring	1,3,5,6
	fall	1,3,5,6
1972:	spring	1,3,5,6
	fall	1,3,5,6
1976:	fall	1,3,5,6

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Appendix A. Savannah River Taxonomic Accession List

- A.1 Algae*
- A.2 Fishes
- A.3 Macro-invertebrates*
- A.4 Protozoa
- A.5 Unidentified Taxa

* Synonyms are included at the ends of these subgroups.

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

<u>NOMENCLATURE</u>	<u>ACCESSION NO.</u>
DIVISION:	
Bacillariophyta	3087
Chlorophyta	3000
Chromophyta	3079
Cyanophyta	3158
Rhodophyta	3204
CLASS:	
Bacillariophyceae	3088
Charophyceae	3074
Cyanophyceae	3159
Euchlorophyceae	3001
Rhodophyceae	3205
Ulothricophyceae	3025
Xanthophyceae	3080
Zygophyceae	3055
ORDER:	
Achnanthales	3118
Acrochaetiales	3211
Bacillariales	3149
Biddulphiales	3096
Chaetophorales	3026
Chamaesiphonales	3161
Charales	3075
Chlorococcales	3002
Chroococcales	3166
Compsopogonales	3207
Epithemiales	3144
Eunotiales	3113
Eupodiscales	3089
Fragilariales	3103
Naviculales	3123
Nemalionales	3216
Nostocales	3176
Oedogoniales	3033
Siphonocladales	3036
Stigonematales	3199

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

<u>NOMENCLATURE</u>	<u>ACCESSION NO.</u>
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ORDER:

Surirellales	3154
Tetrasporales	3021
Tribonematales	3084
Ulotrichales	3043
Ulvales	3052
Vaucheriales	3081
Zygnematales	3056

FAMILY:

Achnanthaceae	3119
Acrochaetiaceae	3212
Anaulaceae	3097
Audouinellaceae	3214
Bactrachospermaceae	3217
Biddulphiaceae	3099
Chaetophoraceae	3027
Chamaesiphonaceae	3162
Characeae	3076
Chroococcaceae	3167
Cladophoraceae	3037
Clastidiaceae	3164
Coccomyxaceae	3003
Compsopogonaceae	3208
Coscinodiscaceae	3090
Cylindrocapsaceae	3044
Cymbellaceae	3136
Desmidiaeae	3057
Entomoneidaceae	3142
Entophysalidaceae	3173
Epithemiaceae	3145
Eunotiaceae	3114
Fragilariaaceae	3104
Gomphonemaceae	3139
Hydrodictyaceae	3006
Lemaneaceae	3219
Mesotaeniaceae	3065
Microsporaceae	3046
Naviculaceae	3124

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

ACCESSION NO.

FAMILY:

Nitzchiaceae	3150
Nostocaceae	3177
Oedogoniaceae	3034
Oocystaceae	3008
Oscillatoriaceae	3181
Palmellaceae	3014
Rivulariaceae	3190
Scenedesmaceae	3018
Schizomeridaceae	3053
Scytonemataceae	3194
Stigonemataceae	3200
Surirellaceae	3155
Tetrasporaceae	3022
Thalassiosiraceae	3094
Tribonemataceae	3085
Ulotrichaceae	3048
Vaucheriaceae	3082
Zygnemataceae	3070

GENUS & SPECIES:

Achnanthes affinis Grun.	269
Achnanthes austriaca v. helvetica Hust.	237
Achnanthes biporoma Hohn & Hell.	238
Achnanthes clevei v. rostrata Hust.	240
Achnanthes clevei Grun.	239
Achnanthes decipiens Reim.	241
Achnanthes deflexa Reim.	242
Achnanthes detha Hohn & Hell.	243
Achnanthes exigua v. heterovalva Krasske	245
Achnanthes exigua Grun.	244
Achnanthes flexella (Kutz.) Brun	246
Achnanthes harveyi Reim.	247
Achnanthes hauckiana Grun.	248
Achnanthes hungarica (Grun.) Grun.	249
Achnanthes inflata (Kutz.) Grun.	250
Achnanthes lanceolata (Breb.) Grun.	251
Achnanthes lanceolata v. abbreviata Reim.	252
Achnanthes lanceolata v. apiculata Patr.	253

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Achnanthes lanceolata</i> v. <i>dubia</i> Grun.	254
<i>Achnanthes lanceolata</i> v. <i>omissa</i> Reim.	255
<i>Achnanthes lapidosa</i> Krasske	270
<i>Achnanthes laponica</i> v. <i>ninckei</i> (Guerm. & Mang.) Reim.	272
<i>Achnanthes laterostrata</i> Hust.	256
<i>Achnanthes linearis</i> (W. Sm.) Grun.	257
<i>Achnanthes marginulata</i> Grun.	273
<i>Achnanthes microcephala</i> (Kutz.) Grun.	275
<i>Achnanthes minutissima</i> Kutz.	258
<i>Achnanthes monela</i> Hohn & Heil.	259
<i>Achnanthes nodosa</i> Cl.	260
<i>Achnanthes peragalli</i> v. <i>fossilis</i> Temp. & Perag.	263
<i>Achnanthes peragalli</i> v. <i>parvula</i> (Patr.) Reim.	262
<i>Achnanthes peragalli</i> Brun & Herib.	261
<i>Achnanthes pinnata</i> Hust.	264
<i>Achnanthes</i> sp. 1	274
<i>Achnanthes stewartii</i> Patr.	265
<i>Achnanthes subhudsonis</i> v. <i>kraeuselli</i> Choln.	267
<i>Achnanthes trinodes</i> (W. Sm.) Grun.	268
<i>Actinella punctata</i> Lewis	200
<i>Amphipleura pellucida</i> Kutz.	285
<i>Amphithrix janthina</i> (Mont.) B. & F.	801
<i>Amphora birugula</i> Hohn	560
<i>Amphora normanii</i> Rabh.	561
<i>Amphora ovalis</i> (Kutz.) Kutz.	562
<i>Amphora ovalis</i> v. <i>pediculus</i> (Kutz.) V. H. ex Det.	563
<i>Amphora perpusilla</i> (Grun.) Grun.	564
<i>Amphora submontana</i> Hust.	565
<i>Amphora veneta</i> Kutz.	566
<i>Anabaina</i> (= <i>Anabaena</i>) <i>circinalis</i> Rabh.	739
<i>Anabaina</i> (= <i>Anabaena</i>) <i>licheniformis</i> Bory	740
<i>Anabaina</i> (= <i>Anabaena</i>) <i>oscillarioides</i> Bory	741
<i>Anabaina</i> (= <i>Anabaena</i>) sp.	743
<i>Anabaina</i> (= <i>Anabaena</i>) <i>variabilis</i> Kutz.	742
<i>Anacystis cyanea</i> (Kutz.) Dr. & Daily	728
<i>Anacystis montana</i> (Lightf.) Dr. & Daily	729
<i>Anacystis montana</i> f. <i>minor</i> (Walle) Dr. & Daily	730
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs	8
<i>Anomoeoneis serians</i> (Breb. ex Kutz.) Cl.	286

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Anomoeoneis serians v. brachysira (Breb. ex Kutz.) Hust.	287
Anomoeoneis vitrea (Grun.) Ross	288
Aphanocapsa sp.	731
Aphanochaete repens A. Braun	33
Asterionella formosa Hass.	145
Asterionella ralfsii v. americana Koerner	147
Audouinella violacea (Kutz.) Hamel	819
Bacillaria paradoxa Gmel.	633
Bactrachospermum ectocarpum ? Sird.	821
Bactrachospermum moniliforme Roth	820
Bactrachospermum spp.	822
Basicladia chelonum (Coll.) Hoff. & Tild.	47
Biddulphia laevis Ehr.	143
Caloneis alpestris (Grun.) Cl.	289
Caloneis bacillum (Grun.) Cl.	290
Caloneis hyalina Hust.	291
Caloneis lewisii v. inflata (Schultze) Patr.	293
Caloneis lewisii Patr.	292
Caloneis limosa (Kutz.) Patr.	294
Caloneis schroderi Hust.	295
Caloneis ventricosa (Ehr.) Meist.	296
Caloneis ventricosa v. alpina (Cl.) Patr.	297
Caloneis ventricosa v. truncatula (Grun.) Meist.	298
Calothrix juliana (Menegh.) B. & F.	802
Calothrix parietana (Nag.) Thur.	803
Calothrix sp.	804
Capartogramma crucicula (Grun. ex Cl.) Ross	299
Chaetomorpha sp.	48
Chaetophora attenuata Hazen	34
Chaetophora elegans (Roth) Ag.	35
Chaetophora incrassata (Huds.) Hazen	36
Chaetophora pisiformis (Roth) Ag.	37
Chaetophora sp.	38
Chamaesiphon incrustans Grun.	726
Chara sp.	117
Chroococcus sp.	732
Cladophora glomerata (L.) Kutz.	49
Closterium acerosum (Schrank) Ehr.	68
Closterium leibleinii Kutz.	69

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Closterium libellula</i> v. <i>interruptum</i> (West & West) Donat	70
<i>Closterium lunula</i> (Mull.) Kutz.	71
<i>Closterium moniliferum</i> (Bory) Ehr.	72
<i>Closterium parvulum</i> Naeg.	73
<i>Closterium</i> sp.	76
<i>Closterium tumidum</i> Johns.	74
<i>Closterium venus</i> Kutz.	75
<i>Coccconeis diminuta</i> Pant.	276
<i>Coccconeis disculus</i> (Schum.) Cl.	283
<i>Coccconeis fluviatilis</i> Wall.	277
<i>Coccconeis pediculus</i> Ehr.	282
<i>Coccconeis placentula</i> v. <i>euglypta</i> (Ehr.) Cl.	279
<i>Coccconeis placentula</i> v. <i>lineata</i> (Ehr.) V. H.	280
<i>Coccconeis placentula</i> Ehr.	278
<i>Coccconeis thumensis</i> A. May.	281
<i>Coelastrum cambricum</i> Arch.	16
<i>Coelastrum microsporum</i> Naeg.	17
<i>Compsopogon coeruleus</i> (Balb.) Mont.	817
<i>Cosmarium granatum</i> Breb.	77
<i>Cosmarium laeve</i> Rabh.	78
<i>Cosmarium polygonum</i> (Naeg.) Arch.	79
<i>Cosmarium pseudoconnatum</i> Nordst.	80
<i>Cosmarium punctulatum</i> v. <i>subpunctulatum</i> (Nordst.) Borg.	81
<i>Cosmarium pygmaeum</i> Arch.	82
<i>Cosmarium pyramidatum</i> Breb.	83
<i>Cosmarium quadratum</i> Ralfs	84
<i>Cosmarium regnelli</i> Wille	85
<i>Cosmarium sexangulare</i> f. <i>minima</i> Nordst.	86
<i>Cosmarium</i> sp.	91
<i>Cosmarium subcrenatum</i> Hantz.	87
<i>Cosmarium subprotumidum</i> Nordst.	88
<i>Cosmarium subspeciosum</i> Nordst.	89
<i>Cosmarium turpinii</i> Breb.	90
<i>Cyclotella atomus</i> Hust.	121
<i>Cyclotella comta</i> (Ehr.) Kutz.	122
<i>Cyclotella meneghiniana</i> Kutz.	123
<i>Cyclotella pseudostelligera</i> Hust.	124
<i>Cyclotella stelligera</i> Cl. & Grun.	125
<i>Cyclotella striata</i> (Kutz.) Grun.	126

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

GENUS & SPECIES:

	<u>ACCESSION NO.</u>
<i>Cylindrocapsa conferta</i> West	52
<i>Cylindrocystis brebissonii</i> Menegh.	104
<i>Cylindrocystis</i> sp.	105
<i>Cylindrospermum catenatum</i> Ralfs	744
<i>Cylindrospermum licheniforme</i> (Bory) Kutz.	745
<i>Cylindrospermum majus</i> Kutz.	746
<i>Cylindrospermum musicola</i> Kutz.	747
<i>Cylindrospermum</i> sp.	748
<i>Cymbella affinis</i> Kutz.	567
<i>Cymbella aspera</i> (Ehr.) H. Per.	568
<i>Cymbella suspidata</i> Kutz.	581
<i>Cymbella delicatula</i> Kutz.	569
<i>Cymbella erdmannensis</i> Foged	580
<i>Cymbella hybrida</i> Grun. ex. Cl.	570
<i>Cymbella lunata</i> W. Sm.	571
<i>Cymbella mexicana</i> (Ehr.) Cl.	582
<i>Cymbella microcephala</i> Grun.	572
<i>Cymbella minuta</i> v. <i>pseudogracilis</i> (Choln.) Reim.	574
<i>Cymbella minuta</i> v. <i>silesiaca</i> (Bleisch ex Rabh.) Reim.	575
<i>Cymbella minuta</i> Hilse ex Rabh.	573
<i>Cymbella naviculiformis</i> Auersw. ex Heib.	576
<i>Cymbella prostrata</i> v. <i>auerswaldii</i> (Rabh.) Reim.	584
<i>Cymbella sinuata</i> Greg.	577
<i>Cymbella triangulum</i> (Ehr.) Cl.	578
<i>Cymbella tumida</i> (Breb. ex Kutz.) V. H.	579
<i>Denticula elegans</i> Kutz.	620
<i>Denticula tenuis</i> Kutz.	621
<i>Desmidium grevillii</i> (Kutz.) DeBary	92
<i>Desmidium swartzii</i> Ag.	93
<i>Desmococcus vixidis</i> (Ag.) Brand	39
<i>Desmogonium rabenhorstianum</i> v. <i>elongatum</i> Patr.	235
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I. ALGAE

NOMENCLATURE

ACCESSION NO.

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1. ALGAE

NOMENCLATUREACCESSION NO.

GENUS & SPECIES:

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1. ALGAE

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<i>Plectonema</i> spp.	810
<i>Plectonema terebrans</i> B. & F.	807
<i>Plectonema tomasinianum</i> (Kutz.) Born.	808
<i>Porphyrosiphon notarisii</i> (Menegh.) Kutz.	790
<i>Porphyrosiphon splendidus</i> (Grev.) Dr.	791
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APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

1. ALGAE

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Rhopalodia gibberula (Ehr.) O. Mull.	632
Scenedesmus abundans (Kirch.) Chod.	18
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Scenedesmus armatus (Chod.) G.M.Smith	25
Scenedesmus bijuga (Turp.) Lagerheim	26
Scenedesmus brasiliensis Bohlin	20
Scenedesmus dimorphus (Turp.) Kutz.	21
Scenedesmus incrassatulus v. mononae G.M.Sm.	22
Scenedesmus opoliensis P. Richter	27
Scenedesmus quadricauda (Turp.) de Breb.	23
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Schizochlamys gelatinosa A. Braun	28
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Schizothrix arenaria (Berk.) Gom.	792
Schizothrix calcicola (Ag.) Gom.	793
Schizothrix friesii (Ag.) Gom.	794
Schizothrix purpurascens (Kutz.) Gom.	795
Schizothrix rivularis (Wolle) Dr.	796
Schizothrix rubella Gom.	797
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Scytonema hofmannii Ag.	812
Scytonema mirabile (Dillw.) Born.	811
Selenastrum minutum (Naeg.) Collins	12
Sphaerocystis schroeteri Chod.	15
Spirogyra spp.	115
Staurastrum alternans Breb.	98
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<i>Stauroneis thermicola</i> v. <i>lanceolata</i> Hust.	559
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<i>Surirella alicula</i> Hohn & Hell.	708
<i>Surirella angusta</i> Kutz.	709
<i>Surirella biseriata</i> Breb.	710
<i>Surirella delicatissima</i> Lewis	711
<i>Surirella guatemalensis</i> Ehr.	712
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1. ALGAE

<u>NOMENCLATURE</u>	<u>ACCESSION NO.</u>
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GENUS & SPECIES:

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<i>Tetraspora gelatinosa</i> (Vauch.) Desv.	30
<i>Tetraspora lubrica</i> (Roth) Ag.	31
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<i>Thalassiosira fluviatilis</i> Hust.	141
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<i>Tolypothrix tenuis</i> Kutz.	813
<i>Tribonema bombycinum</i> (C. A. Ag.) Derb. & Sol.	120
<i>Tuomeya fluviatilis</i> Harv.	823
<i>Ulothrix aequalis</i> Kutz.	62
<i>Ulothrix oscillarina</i> Kutz.	63
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<i>Uronema elongatum</i> Hodg.	66
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<i>Zygnema</i> spp.	116

Synonyms of the Division Cyanophyta according to Frances Drouet

<u>Synonym</u>	<u>ID. No.</u>	<u>Computer Nomenclature</u>
<i>Agmenellum quadruplicatum</i> (Menegh.) Breb	734 735	<i>Merismopedia tenuissima</i> Lemm. <i>M. tranquilla</i> (Ehr.) Trevis.
<i>Entophysalis lemaniae</i> (Ag.) Dr. & Daily	726	<i>Chamaesiphon incrassans</i> Grun.
<i>Stigonema muscicola</i> (Thur.) Borzi	814	<i>Hapalosiphon Fontinale</i> (Ag.) Born.
<i>S. ocellatum</i> (dillw.) Thur.	816	<i>Stigonema mesentericum</i> Geitl.
<i>Scytonema hofmannii</i> Ag.	806 811	<i>Thorea wrangelii</i> Ag. <i>Scytonema mirabile</i> (Dillw.) Born.
	813	<i>Tolypothrix tenuis</i> Kutz.
<i>Schizothrix calcicola</i> (Ag.) Gom.	807	<i>Plectonema terebrans</i> B. & F.
	809	<i>P. nostocorm</i> ? Born.
	801	<i>Amphithrix janthina</i> (Mont.) B. & F.
	758	<i>Lyngbya epiphytica</i> Hieronymus
	757	<i>L. contorta</i> Lemm.
	760	<i>L. versicolor</i> (Wartm.) Gom.
	770	<i>Oscillatoria geminata</i> Schw.
	785	<i>Phormidium purpurascens</i> (Kutz.) Gom.
	787	<i>P. tenue</i> (Menegh.) Gom.
<i>Porphyrosiphon notarisii</i> (Menegh.) Kutz	808	<i>Plectonema tomasinianum</i> (Kutz.) Born.
	768	<i>Oscillatoria chalybea</i> Mert.
	792	<i>Schizothrix aernaria</i> (Berk.) Gom.
<i>Calothrix parietana</i> (Nag.) Thur.	802	<i>Calothrix juliana</i> (Menegh.) B. & F.
	751	<i>Nostoc linckia</i> (Roth) Born & Thuret
<i>Nostoc commune</i> Vauch.	739	<i>Anabaina (=Anabaena) circinalis</i> Rabh.
	749	<i>Nostoc carneum</i> Ag.
	752	<i>N. muscorum</i> Ag.
<i>Anabaina oscillarioides</i> Bory	742	<i>Anabaina variabilis</i> Kutz.

<u>Synonym</u>	<u>ID. No.</u>	<u>Computer Nomenclature</u>
<i>Anabaina licheniformis</i> Bory	744	<i>Cylindrospermum catenatum</i> Ralfs
	745	<i>C. licheniforme</i> (Bory) Kutz.
	746	<i>C. majus</i> Kutz.
	747	<i>C. musicola</i> Kutz.
<i>Microcoleus vaginatus</i> (Vauch.) Gom	754	<i>Lyngbya aerugineo-caerulea</i> (Kutz.) Gom.
	781	<i>Phormidium ambiguum</i> Gom.
	782	<i>P. autumnale</i> (Ag.) Gom.
	783	<i>P. favosum</i> (Bory) Gom.
	788	<i>P. uncinatum</i> (Ag.) Gom.
<i>M. lyngbyaceus</i> Ketz.	755	<i>Lyngbya aestuarii</i> (Mert.) Liebmann
	767	<i>Oscillatoria articulatata</i> Gardn.
	771	<i>O. limosa</i> (Roth) Ag.
	775	<i>O. rubescens</i> DeCand.
	778	<i>O. tenuis</i> Ag.
<i>Oscillatoria retzii</i> Ag.	756	<i>Lyngbya bicolor</i> Wood
	786	<i>Phormidium retzii</i> (Ag.) Gom.
	800	<i>Symploca muscorum</i> (Ag.) Gom.
<i>Schizothrix mexicana</i> Gom.	759	<i>Lyngbya putealis</i> Mont. ex Gom.
<i>S. arenaria</i> (Berk.) Gom.	762	<i>Microcoleus chthonoplastes</i> (Mert.) Zanard
<i>S. rubella</i> Gom.	765	<i>M. paludosus</i> (Kutz.) Gom.
	784	<i>Phormidium inundatum</i> Kutz.
	799	<i>Symploca muralis</i> Kutz.
<i>Porphyrosiphon animals</i> (Ag.)	769	<i>Oscillatoria formosa</i> Bory
Dr.		
<i>Oscillatoria princeps</i> Vauch.	773	<i>O. proboscidea</i> Gom.
<i>Porphyrosiphon splendidus</i> (Grev.) Dr.	776	<i>O. splendida</i> Grev.
<i>Schizothrix friesii</i> (Ag.)	796	<i>Schizothrix rivularis</i> (Wolle) Dr.
Gom.		

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

2. FISHES

NOMENCLATURE

ACCESSION NO.

PHYLUM:

Chordata	4313
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CLASS:

Osteichthyes	4314
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ORDER:

Amiiformes	4315
Anguilliformes	4318
Atheriniformes	4323
Clupeiformes	4332
Cypriniformes	4339
Perciformes	4355
Percopsciformes	4374
Pleuronectiformes	4379
Salmoniformes	4384
Semionotiformes	4389
Siluriformes	4392

FAMILY:

Amblyopsidae	4375
Amiidae	4316
Anguillidae	4319
Aphredoderidae	4377
Atherinidae	4324
Belonidae	4326
Catostomidae	4340
Centrarchidae	4356
Clupeidae	4333
Cyprinidae	4346
Cyprinodontidae	4328
Echelidae	4321
Engraulidae	4337
Esocidae	4385
Ictaluridae	4393
Lepisosteidae	4390
Mugilidae	4364

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NOMENCLATURE

ACCESSION NO.

FAMILY:

Paralichthidae	4380
Percichthyidae	4366
Percidae	4368
Poeciliidae	4330
Sciaenidae	4372
Soleidae	4382
Umbridae	4387

GENUS & SPECIES:

Acantharchus pomotis (Baird)	2404
Alosa aestivalis (Mitchill)	2361
Alosa chrysocloris (Rafinesque)	2362
Alosa mediocris (Mitchill)	2363
Alosa sapidissima (Wilson)	2364
Alosa sp.	2365
Amia calva Linnaeus	2353
Anchoa mitchilli (Valenciennes)	2369
Anguilla rostrata (Lesueur)	2354
Aphredoderus sayanus (Gilliams)	2439
Aploodonotus grunniens Rafinesque	2437
Brevoortia tyrannus (Latrobe)	2366
Carpiodes carpio (Rafinesque)	2370
Carpiodes cyprinus (Lesueur)	2371
Carpiodes spp.	2372
Centrarchus macropterus (Lacepede)	2405
Chologaster cornuta Agassiz	2438
Cyprinus carpio Linnaeus	2380
Dorosoma cepedianum (Lesueur)	2367
Dorosoma petenense (Gunther)	2368
Elassoma zonatum Jordan	2406
Enneacanthus gloriosus (Holbrook)	2407
Enneacanthus sp.	2408
Erimyzon oblongus (Mitchill)	2373
Erimyzon spp.	2375
Erimyzon suetta (Lacepede)	2374
Esox americanus vermiculatus Lesueur	2443
Esox americanus Gmelin	2442
Esox niger Lesueur	2444

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

2. FISHES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Esox sp.	2445
Etheostoma fricksium Hildebrand	2425
Etheostoma fusiforme (Girard)	2426
Etheostoma fusiforme barratti (Holbrook)	2427
Etheostoma hopkinsi (Fowler)	2428
Etheostoma inscriptum (Jordan & Brayton)	2429
Etheostoma nigrum Rafinesque	2430
Etheostoma olmstedi Storer	2431
Etheostoma serriferum (Hubbs & Cannon)	2432
Etheostoma spp.	2433
Fundulus lineolatus (Agassiz)	2358
Fundulus notti (Agassiz)	2359
Gambusia affinis (Baird & Girard)	2360
Hybognathus nuchalis regius Girard	2382
Hybognathus nuchalis Agassiz	2381
Hybopsis amblops (Rafinesque)	2383
Hybopsis rubrifrons (Jordan)	2384
Hypentelium nigricans (Lesueur)	2376
Ictalurus brunneus (Jordan)	2450
Ictalurus catus (Linnaeus)	2451
Ictalurus melas (Rafinesque)	2452
Ictalurus natalis (Lesueur)	2453
Ictalurus nebulosus (Lesueur)	2454
Ictalurus platycephalus (Girard)	2455
Ictalurus punctatus (Rafinesque)	2456
Labidesthes sicculus (Cope)	2356
Lepisosteus oculatus (Winchell)	2447
Lepisosteus osseus (Linnaeus)	2448
Lepisosteus platyrhincus DeKay	2449
Lepomis auritus (Linnaeus)	2409
Lepomis cyanellus Rafinesque	2410
Lepomis gibbosus (Linnaeus)	2411
Lepomis gulosus (Cuvier)	2412
Lepomis macrochirus Rafinesque	2413
Lepomis marginatus (Holbrook)	2414
Lepomis megalotis (Rafinesque)	2415
Lepomis microlophus (Gunther)	2416
Lepomis punctatus (Valenciennes)	2417
Micropterus punctulatus (Rafinesque)	2418

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

2. FISHES

<u>NOMENCLATURE</u>	<u>ACCESSION NO.</u>
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GENUS & SPECIES:

<i>Micropterus salmoides</i> (Lacepede)	2419
<i>Minytrema melanops</i> (Rafinesque)	2377
<i>Morone chrysops</i> (Rafinesque)	2423
<i>Morone saxatilis</i> (Walbaum)	2424
<i>Moxostoma anisurum</i> Rafinesque	2378
<i>Moxostoma</i> spp.	2379
<i>Mugil cephalus</i> Linnaeus	2422
<i>Myrophis punctatus</i> Lutken	2355
<i>Nocomis leptocephalus bellus</i> Girard	2386
<i>Nocomis leptocephalus</i> Girard	2385
<i>Nocomis micropogon</i> (Cope)	2387
<i>Notemigonus crysoleucas</i> (Mitchill)	2388
<i>Notropis ardens</i> (Cope)	2389
<i>Notropis chalybaeus</i> (Cope)	2390
<i>Notropis cummingsae</i> Myers	2391
<i>Notropis emiliae</i> (Hay)	2392
<i>Notropis hudsonius</i> (Clinton)	2393
<i>Notropis leedsi</i> Fowler	2394
<i>Notropis lutipinnis</i> (Jordan & Brayton)	2395
<i>Notropis maculatus</i> (Hay)	2396
<i>Notropis niveus</i> (Cope)	2397
<i>Notropis petersoni</i> Fowler	2398
<i>Notropis spilopterus</i> (Cope)	2399
<i>Notropis</i> spp.	2401
<i>Notropis xanoccephalus</i> (Jordan)	2400
<i>Noturus gyrinus</i> (Mitchill)	2457
<i>Noturus insignis</i> (Richardson)	2458
<i>Noturus leptacanthus</i> Jordan	2459
<i>Noturus</i> sp.	2460
<i>Paralichthys lethostigma</i> Jordan & Gilbert	2440
<i>Perca flavescens</i> (Mitchill)	2434
<i>Percina caprodes</i> (Rafinesque)	2435
<i>Percina nigrofasciata</i> (Agassiz)	2436
<i>Pimephales vigilax</i> (Baird & Girard)	2402
<i>Pomoxis annularis</i> Rafinesque	2420
<i>Pomoxis nigromaculatus</i> (Lesueur)	2421
<i>Pylodictis olivaris</i> (Rafinesque)	2461
<i>Semotilus atromaculatus</i> (Mitchill)	2403
<i>Strongylura marina</i> (Walbaum)	2357

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

2. FISHES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Trinectes maculatus (Block & Schneider)
Umbra pygmaea (DeKay)

2441
2446

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

PHYLUM:

Annelida	3735
Arthropoda	3877
Aschelminthes	3649
Bryozoa	3719
Mollusca	3808
Nemertea	3644
Platyhelminthes	3623
Porifera	3615
Tardigrada	3714

CLASS:

Arachnoidea	3928
Bivalvia	3849
Crustacea	3878
Demospongae	3616
Ectoprocta	3720
Enopla	3645
Eutardigrada	3715
Gastropoda	3809
Gastrotricha	3699
Hirudinoidia	3736
Insecta	3959
Nematoda	3703
Nematomorpha	3710
Oligochaeta	3760
Rotifera	3650
Turbellaria	3624

ORDER:

Alloeocoela	3638
Amphipoda	3911
Arhynchobdellidae	3754
Basommatophora	3831
Bdelloidea	3654
Branchiura	3900
Catenulida	3629
Cladocera	3881

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

ORDER:

Coleoptera	4100
Decapoda	3919
Diptera	4208
Ephemeroptera	3994
Eucopepoda	3896
Gordioidea	3711
Gymnolaemata	3721
Haplosclerina	3617
Haplotaixida	3767
Hemiptera	4059
Hoploneurtea	3646
Hydrocarina	3929
Isopoda	3906
Lepidoptera	4205
Lumbriculida	3761
Macrobiotoidea	3716
Megaloptera	4094
Mermithoidea	3707
Mesogastropoda	3811
Monogononta	3657
Neorhabdoocela	3634
Neuroptera	4091
Odonata	3960
Phylactolaemata	3726
Plecoptera	4037
Rhynchobellae	3741
Trichoptera	4169
Tricladida	3625
Unionoida	3851
Veneroida	3867

FAMILY:

Aeolosomatidae	3806
Aeshnidae	3977
Ancylidae	3838
Argulidae	3902
Arrenuridae	3930
Asellidae	3908

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

FAMILY:

Astacidae	3923
Baetidae	4000
Belostomatidae	4083
Brachionidae	3681
Brachycentridae	4194
Caenidae	4030
Calopterygidae	3962
Capniidae	4044
Catenulidae	3630
Ceratopogonidae	4224
Chaoboridae	4222
Chironomidae	4229
Chydoridae	3886
Coenagrionidae	3965
Corbiculidae	3870
Corduliidae	3984
Corixidae	4087
Corydalidae	4097
Culicidae	4219
Cyclopidae	3898
Dalyelliidae	3635
Dicranophoridae	3667
Dolichopodidae	4304
Dryopidae	4159
Dytiscidae	4108
Elmidae	4162
Empididae	4302
Enchytraeidae	3770
Ephemerellidae	4019
Ephemeridae	4032
Ephydriidae	4311
Erpobdellidae	3755
Eylaidae	3933
Flosculariidae	3697
Fredericellidae	3727
Gammaridae	3914
Gastropodidae	3674
Gelastocoridae	4077
Gerridae	4062

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

FAMILY:

Glossiphoniidae	3744
Glossoscolecidae	3792
Gomphidae	3970
Gordiidae	3712
Gyrinidae	4130
Haliplidae	4104
Heptageniidae	4009
Hydrobiidae	3821
Hydrometridae	3935
Hydrometridae	4074
Hydrophilidae	4135
Hydropsychidae	4178
Hydroptilidae	4184
Hygrobatidae	3937
Krendowskiiidae	3941
Lebertiidae	3943
Lecanidae	3691
Lepidostomatidae	4197
Leptoceridae	4199
Leptophlebiidae	4016
Libellulidae	3988
Limnephilidae	4189
Limnesiidae	3945
Lindiidae	3665
Lophopodidae	3729
Lumbricidae	3796
Lumbriculidae	3762
Lymnaeidae	3833
Macrobiotidae	3717
Macromiidae	3982
Megascolecidae	3801
Mermithidae	3708
Mesovelidae	4072
Naididae	3778
Naucoridae	4081
Nemouridae	4040
Neoephemeridae	4028
Nepidae	4079
Notommatidae	3660

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

FAMILY:

Notonectidae	4085
Oligoneuriidae	4007
Omophronidae	4102
Oxidae	3948
Palaemonidae	3920
Paludicellidae	3722
Perlidae	4052
Perlodidae	4049
Philopotamidae	4171
Physidae	3847
Pionidae	3950
Piscicolidae	3750
Plagiostomidae	3639
Planariidae	3626
Planorbidae	3842
Pleuroceridae	3828
Plumatellidae	3732
Polycentropodidae	4173
Pteronarcidae	4047
Pyralidae	4206
Rhagionidae	4299
Sciomyzidae	4308
Sialidae	4095
Sididae	3884
Simuliidae	4289
Siphlonuridae	3995
Sisyridae	4092
Sphaeriidae	3872
Spongillidae	3618
Stenostomidae	3632
Stratiomyidae	4294
Synchaetidae	3677
Tabanidae	4296
Taeniopterygidae	4042
Talitridae	3912
Testudinellidae	3695
Tetrastemmatidae	3647
Tipulidae	4211
Trichocercidae	3671

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

FAMILY:

Tricorythidae	4025
Tubificidae	3773
Unionicolidae	3954
Unionidae	3853
Valvatidae	3818
Veliidae	4068
Victorellidae	3724
Viviparidae	3815

GENUS & SPECIES:

Ablabesmyia mallochi (Walley)	2265
Ablabesmyia nr. monilis (L.)	2266
Ablabesmyia nr. philosophagnos Beck & Beck	2268
Ablabesmyia poss. aspera Roback	2267
Ablabesmyia sp. e Roback	2269
Ablabesmyia spp.	2270
Acroneuria abnormis (Newman)	1994
Acroneuria arida (Hagen)	1995
Acroneuria georgiana Banks	1996
Acroneuria sp.	1998
Acroneuria sp. a. Frison	1997
Aeolosoma sp(p?).	1761
Aeshna sp.	1939
Alasmidonta arcula Lea	1797
Alasmidonta triangulata (Lea)	1798
Alona guttata Sars	1864
Amnicola (Lyogyrus) sp.	1770
Amnicola limosa (Say)	1767
Amnicola n. sp. a	1768
Amnicola n. sp. b	1769
Anacaena infuscata Motschulsky	2131
Ancyronyx sp.	2177
Ancyronyx variegatus (Germar)	2176
Anodonta cataracta Say	1799
Anodonta couperiana Lea	1801
Anodonta hallenbeckii Lea	1802
Anodonta imbecillis Say	1803
Anodonta nr. cataracta Say	1800

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATUREACCESSION NO.

GENUS & SPECIES:

Anopheles sp.	2250
Arcteonais lomondi (Martin)	1727
Argia apicalis (Say)	1914
Argia moesta (Hagen)	1915
Argia sedula (Hagen)	1916
Argia sp.	1920
Argia tibialis (Rambur)	1917
Argia translata Hagen	1918
Argia violacea (Hagen)	1919
Argulus sp.	1869
Arrenurus sp(p?).	1889
Asellus attenuatus Richardson	1870
Asellus communis (Say)	1871
Asellus militaris Hay	1872
Aspelta aper (Herring)	1612
Atanytarsus sp.	2336
Atherix sp.	2348
Atractides sp.	1893
Atrichopogon sp.	2253
Aulophorus furcatus (Muller)	1728
Baetis spp.	1959
Basiaeschna janata (Say)	1940
Batracobdella phalera (Graf)	1682
Belostoma flumineum Say	2038
Belostoma lutarium (Stal)	2039
Belostoma sp.	2040
Berosus aculeatus Leconte	2132
Berosus exiguus Say	2133
Berosus fraternus Leconte	2134
Berosus infuscatus Leconte	2135
Berosus pantherinus Leconte	2136
Berosus peregrinus (Herbst)	2137
Berosus pugnax ? Leconte	2139
Berosus sp.	2140
Berosus striatus (Say)	2138
Bezzia or Probezzia sp.	2254
Bidessus obesus ? Shp.	2075
Boyeria vinoso (Say)	1941
Brachionus calyciflorus Pallas	1630

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Brachionus havanaensis Rousselet	1631
Brachionus quadridentatus Hermann	1632
Brachycentrus numerosus (Say)	2226
Brillia par (Coquillett) var.	2273
Caborius sp.	2222
Caenis spp.	1983
Callibaetis sp.	1960
Calopteryx maculata Beauvais	1910
Calopteryx sp.	1911
Campeloma decisum (Say)	1763
Campeloma lima Anthony	1764
Campeloma sp.	1765
Carunculina parva (Barnes)	1805
Carunculina patrickae Bates	1806
Carunculina pulla (Conrad)	1807
Castrella sp.	1588
Catenula sp.	1586
Centroptilum sp.	1961
Cephalodella auriculata (Muller)	1597
Cephalodella eva (Gosse)	1598
Cephalodella forficula (Ehrenberg)	1599
Cephalodella gibba (Ehrenberg)	1600
Cephalodella gracilis (Ehrenberg)	1601
Cephalodella hiulca Myers	1602
Cephalodella hoodi (Gosse)	1603
Cephalodella intuta Myers	1604
Cephalodella megalcephala (Glasscott)	1605
Cephalodella tenuior (Gosse)	1606
Ceraclea maculata (Banks)	2229
Ceraclea nr. mentiae (Walker)	2231
Ceraclea nr. spongillivorax (Resh)	2232
Ceraclea transversa (Hagen)	2230
Cercyon sp.	2167
Chaetogaster diaphanus (Gruithuisen)	1729
Chaetogaster limnaei von Baer	1730
Chaetogaster sp(p?).	1731
Chaoborus punctipennis (Say)	2252
Chauliodes sp.	2052
Chernovskia sp.	2298

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Cheumatopsyche spp.	2209
Chimarra nr. feria Ross	2202
Chimarra prob. aterrima Hagen	2201
Chimarra prob. socia Hagen	2203
Chimarra sp.	2204
Chironomus decorus Johannsen	2299
Chromogaster ovalis (Bergendal)	1623
Chrysops sp.	2346
Cladopelma nais ? (Townes)	2300
Climacea areolaris Hagen	2050
Clinotanypus pinguis (Loew)	2259
Cloeon sp.	1962
Coelambus (Hygrotus) spp.	2080
Coelambus dissimilis Gemminger & Harold	2076
Coelambus inaequalis (Fabricius)	2077
Coelambus laccophilinus Leconte	2078
Coelambus spp.	2079
Coelotanypus scapularis (Loew)	2257
Coelotanypus tricolor (Loew)	2258
Colurella anodonta Carlin	1633
Colurella colura (Ehrenberg)	1635
Colurella obtusa (Gosse)	1636
Colurella sinistra Carlin	1637
Colurella uncinata (Muller)	1638
Copelatus glyphicus (Say)	2096
Copelatus sp.	2097
Coptotomus interrogatus (Fabricius)	2098
Coptotomus spp.	2099
Corbicula fluminea (Muller)	1841
Corbicula manilensis (Philippi)	1842
Cordites sp.	2274
Corydalus cornutus (Linnaeus)	2053
Corynoneura (Corynoneura) nr. celeripes Winnertz	2275
Corynoneura (Corynoneura) nr. taris Roback	2276
Corynoneura (Thienemanniella) nr. xena Roback	2277
Corynoneura (Thienemanniella) sp.	2278
Crangonyx gracilis Smith	1875
Crenitis subcupreus (Say)	2141
Cricotopus bicinctus (Meigen)	2279

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Cricotopus nr. slossonae Malloch	2281
Cricotopus poss. fugax (Johannsen)	2280
Cricotopus sp. ("Sylvestris group")	2282
Cryptochironomus digitatus (Malloch)	2301
Cryptochironomus nr. argus Roback	2302
Cryptochironomus sp.	2303
Cryptocladopelma amachaerus (Townes)	2304
Cryptocladopelma poss. viridulus (Linnaeus)	2305
Cryptopleurum minutum (Fabricius)	2168
Cryptotendipes pseudotener (Goetghebuer)	2306
Culex sp.	2251
Culicoides ? sp.	2255
Cybbiodyta lacustris Leconte	2142
Dalyellia sp.	1589
Dapidia calpidia Myers	1639
Demicryptochironomus sp.	2307
Dero digitata (Muller)	1732
Dero sp.	1733
Deronectes sp.	2081
Desmopachria convexa Aube	2082
Desmopachria granum (Leconte)	2083
Dicranomyia sp.	2244
Dicranophorus artamus Harring & Myers	1613
Dicranophorus myriophylli (Harring)	1614
Dicrotendipes modestus (Johannsen)	2308
Dicrotendipes nervosus (Staeger)	2309
Dicrotendipes nr. neomodestus (Malloch)	2310
Dicrotendipes sp.	2311
Dictya sp.	2351
Dina parva Moore	1704
Dina sp. 1	1705
Dina sp. 2	1706
Dineutes analis Regimbart	2105
Dineutes angustus Leconte	2106
Dineutes assimilis (Kirby)	2107
Dineutes carolinus Leconte	2108
Dineutes discolor Aube	2109
Dineutes emarginatus Say	2110
Dineutes horni Roberts	2111

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Dineutes spp.	2113
Dineutes vittatus (Germar)	2112
Diplodontus despiciens (Muller)	1892
Diurella bidens Luck	1616
Dubiraphia bivittata Leconte	2178
Dubiraphia quadrinotata (Say)	2179
Dubiraphia sp.	2181
Dubiraphia vittata (Melsheimer)	2180
Dugesia tigrina (Girard)	1584
Eclipidrilus sp.	1711
Eisenella tetraedra (Savigny)	1751
Elliptio arctatus (Conrad)	1819
Elliptio complanata (Lightfoot)	1820
Elliptio complanatus (Solander)	1821
Elliptio complanatus northamptonensis (Lea)	1822
Elliptio congareea (Lea)	1823
Elliptio crassidens (Lamarck)	1824
Elliptio crassidens congareus Lea	1825
Elliptio fisherianus Lea	1826
Elliptio forbesiana (Lea)	1827
Elliptio fraterna (Lea)	1828
Elliptio hopetonensis Lea	1829
Elliptio icterina (Conrad)	1830
Elliptio incrassatus Lea	1831
Elliptio lanceolata (Lea)	1832
Elliptio lecontianus (Lea)	1833
Elliptio obnubilus (Lea)	1834
Elliptio productus (Conrad)	1835
Elliptio subinflatus (Conrad)	1836
Elliptio tuomeyi (Lea)	1837
Enallagma civile (Hagen)	1921
Enallagma divagans Selys	1922
Enallagma exsulans (Hagen)	1923
Enallagma signatum (Hagen)	1924
Enallagma spp.	1926
Enallagma weewa Byers	1925
Encentrum grande (Western)	1615
Endochironomus nigricans (Johannsen)	2312
Enochrus cinctus (Say)	2143

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Enochrus nebulosus</i> (Say)	2144
<i>Enochrus ochraceus</i> Melsheimer	2145
<i>Enochrus perplexus</i> Leconte	2146
<i>Enochrus</i> spp.	2147
<i>Ephemerella</i> (<i>Dannella</i>) <i>simplex</i> McD.	1973
<i>Ephemerella</i> (<i>Ephemerella</i>) <i>rotunda</i> Morgan	1974
<i>Ephemerella</i> (<i>Ephemerella</i>) spp.	1975
<i>Ephemerella</i> (<i>Eurylophella</i>) nr. <i>temporalis</i> McD.	1977
<i>Ephemerella</i> (<i>Eurylophella</i>) spp.	1978
<i>Ephemerella</i> (<i>Eurylophella</i>) <i>temporalis</i> McD.	1976
<i>Ephemerella</i> (<i>Serratella</i>) <i>deficiens</i> Morgan	1979
<i>Ephemerella</i> spp.	1972
<i>Ephydra</i> sp.	2352
<i>Epicordulia princeps</i> (Hagen)	1946
<i>Epicordulia</i> sp.	1947
<i>Erpetogomphus designatus</i> Hagen	1931
<i>Erpobdella punctata</i> (Leidy)	1707
<i>Erythemis simplicicollis</i> (Say)	1951
<i>Euchlanis dilatata</i> Ehrenberg	1640
<i>Euchlanis meneta</i> Myers	1641
<i>Euchlanis parva</i> Rousselet	1642
<i>Euchlanis triquetra</i> Ehrenberg	1643
<i>Eucrangonyx gracilis</i> Smith	1876
<i>Eukiefferiella</i> poss. <i>sordens</i> Johannsen	2283
<i>Eukiefferiella</i> sp.	2284
<i>Eupera cubensis</i> (Prime)	1843
<i>Eupera singleyi</i> Pilsbry	1844
<i>Eupera</i> spp.	1845
<i>Eurycercus lamellatus</i> (Muller)	1865
<i>Eylais</i> sp.	1891
<i>Ferrissia fusca</i> (C.B. Adams)	1783
<i>Ferrissia rivularis</i> (Say)	1782
<i>Ferrissia</i> spp.	1784
<i>Fredericella sultana</i> (Blumenbach)	1673
<i>Galba humilis modicella</i> (Say)	1777
<i>Gammarus fasciatus</i> Say	1877
<i>Gammarus</i> sp.	1878
<i>Gastropus hytopus</i> (Ehrenberg)	1624
<i>Gelastocoris oculatus</i> (Fabricius)	2031

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Geosargus sp.	2345
Geranomyia sp.	2246
Gerris alacris Hussey	2007
Gerris canaliculatus Say	2008
Gerris conformis Uhler	2009
Gerris marginatus Say	2010
Gerris nebularis Drake & Hottes	2011
Gerris nr. buenoi Hungerford	2012
Gerris sp.	2013
Glossiphonia complanata (Linnaeus)	1683
Glossiphonia heteroclita (Linnaeus)	1684
Glossiphonia nepheloidea (Graf)	1685
Glyptotendipes lobiferus ? (Say)	2313
Glyptotendipes spp.	2315
Glyptotendipes spp. nr. senilis (Johannsen)	2314
Gomphus (Gomphurus) lividus Selys	1932
Gomphus (Gomphurus) spp.	1934
Gomphus (Gomphurus) vastus Walsh	1933
Gomphus (Stylurus) plagiatus Selys	1935
Gomphus (Stylurus) sp.	1936
Goniobasis caternia (Say)	1774
Goniobasis sp.	1775
Gordius spp.	1669
Gyraulus parvus (Say)	1787
Gyraulus sp(p).	1788
Gyrinus affinis Aube	2114
Gyrinus analis Say	2115
Gyrinus borealis lugens Leconte	2116
Gyrinus dichrous Leconte	2117
Gyrinus fraternus Couper	2118
Gyrinus gibber Leconte	2119
Gyrinus limbatus Say	2120
Gyrinus lugens Leconte	2121
Gyrinus minutus Fabricius	2122
Gyrinus nr. rockinghamensis Leconte	2125
Gyrinus opacus Sahlberg	2123
Gyrinus pectoralis Leconte	2124
Gyrinus spp.	2126
Hagenius brevistylus Selys	1937

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3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Haliphus borealis</i> Leconte	2055
<i>Haliphus punctatus</i> Aube	2056
<i>Haliphus</i> sp.	2058
<i>Haliphus triopsis</i> Say	2057
<i>Helichus fastigiatus</i> (Say)	2171
<i>Helichus lithophilus</i> Germar	2172
<i>Helichus</i> sp.	2174
<i>Helichus striatus</i> Leconte	2173
<i>Helisoma anceps</i> (Menke)	1789
<i>Helisoma trivolvis</i> (Say)	1790
<i>Helobdella elongata</i> (Castle)	1686
<i>Helobdella fusca</i> (Castle)	1687
<i>Helobdella lineata</i> (Verrill)	1688
<i>Helobdella nepheoloidea</i> (Graf)	1689
<i>Helobdella papillata</i> Moore	1690
<i>Helobdella punctata</i> Moore	1691
<i>Helobdella</i> spp.	1693
<i>Helobdella staghialis</i> (Linnaeus)	1692
<i>Helochares</i> (<i>Creniphilus</i>) sp.	2150
<i>Helochares maculicollis</i> Mulsant	2148
<i>Helochares</i> sp.	2149
<i>Helophorus lineatus</i> Say	2127
<i>Helophorus</i> sp.	2129
<i>Helophorus tuberculatus</i> Gyll.	2128
<i>Hemerodromia</i> sp.	2349
<i>Heptagenia</i> spp.	1966
<i>Herpobdella punctata</i> (Leidy)	1708
<i>Hesperophylax</i> sp.	2223
<i>Hetaerina americana</i> (Fabricius)	1912
<i>Hetaerina titia</i> (Drury)	1913
<i>Heterocloeon</i> sp.	1963
<i>Heteromeyenia ryderii</i> Potts	1578
<i>Hexagenia</i> sp.	1984
<i>Homeoneuria dolani</i> Edmunds, Berner, Travis	1965
<i>Hyalella azteca</i> (Saussure)	1874
<i>Hyalinella punctata</i> (Hancock)	1676
<i>Hydaticus bimarginatus</i> Say	2103
<i>Hydaticus laevipennis</i> Thompson	2104
<i>Hydrobaenus pilipes</i> ? (Malloch)	2285

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3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Hydrobius tesselatus</i> Zimm.	2151
<i>Hydrobius tumidus</i> Leconte	2152
<i>Hydrocanthus iricolor</i> Say	2068
<i>Hydrochoreutes</i> sp.	1904
<i>Hydrochus subcupreus</i> Rand.	2130
<i>Hydrolimax grisea</i> Haldeman	1590
<i>Hydrometra hungerfordi</i> Bueno	2029
<i>Hydrometra martini</i> Kirkaldy	2030
<i>Hydrophilus obtusatus</i> Say	2153
<i>Hydrophilus</i> sp.	2154
<i>Hydroporus?</i> sp.	2350
<i>Hydroporus clypealis</i> Sharp	2084
<i>Hydroporus consimilis</i> Leconte	2085
<i>Hydroporus lynceus</i> Sharp	2086
<i>Hydroporus mellitus</i> Leconte	2087
<i>Hydroporus</i> nr. <i>mixtus</i> Leconte	2093
<i>Hydroporus pilatei</i> Fall.	2088
<i>Hydroporus proximus</i> Aube	2089
<i>Hydroporus pulcher</i> Leconte	2090
<i>Hydroporus</i> spp.	2094
<i>Hydroporus undulatus</i> Say	2091
<i>Hydroporus wickhami</i> Zaitzev	2092
<i>Hydropsyche</i> nr. <i>cuanis</i> Ross	2212
<i>Hydropsyche orris</i> Ross	2210
<i>Hydropsyche</i> prob. <i>frisoni</i> Ross	2213
<i>Hydropsyche</i> prob. <i>venularis</i> Banks	2211
<i>Hydropsyche</i> sp.	2214
<i>Hydroptila</i> sp.	2219
<i>Hydrovatus pustulatus</i> (Melsheimer)	2095
<i>Hygrobates</i> sp.	1894
<i>Hypsibius</i> sp.	1670
<i>Ilinobdella moorei</i> Meyer	1701
<i>Ilybius ignarus</i> Leconte	2100
<i>Ischnura posita</i> (Hagen)	1927
<i>Ischnura ramburi</i> ? (Selys)	1929
<i>Ischnura</i> sp.	1930
<i>Ischnura verticalis</i> (Say)	1928
<i>Isogenus</i> sp.	1992
<i>Isonychia</i> spp.	1958

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NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Isoperla</i> spp.	1993
<i>Keratella cochlearis</i> (Gosse)	1644
<i>Keratella gracilenta</i> Ahlstrom	1645
<i>Krendowskia similis</i> Viets	1896
<i>Labrundinia pilosella</i> (Loew)	2271
<i>Laccobius agilis</i> Rand.	2155
<i>Laccobius</i> sp.	2156
<i>Laccophilus fasciatus</i> Aube	2071
<i>Laccophilus maculosus</i> Say	2072
<i>Laccophilus proximus</i> Say	2073
<i>Laccophilus</i> sp.	2074
<i>Laevapex fuscus</i> (C. B. Adams)	1785
<i>Lampsilis cariosa</i> (Say)	1808
<i>Lampsilis dolabraeformis</i> (Lea)	1809
<i>Lampsilis radiata</i> (Lea)	1810
<i>Lampsilis radiata splendida</i> (Lea)	1811
<i>Lampsilis recta</i> Lamarck	1812
<i>Lampsilis</i> sp.	1814
<i>Lampsilis splendida</i> (Lea)	1813
<i>Lebertia</i> sp. 1	1897
<i>Lebertia</i> sp. 2	1898
<i>Lebertia</i> sp. 3	1899
<i>Lebertia</i> sp(p?).	1900
<i>Lecane aegaena</i> Harring	1654
<i>Lecane bulla</i> (Gosse)	1655
<i>Lecane closterocerca</i> (Schmarda)	1656
<i>Lecane elachis</i> (Harring & Myers)	1657
<i>Lecane flexilis</i> (Gosse)	1658
<i>Lecane hamata</i> Stokes	1659
<i>Lecane luna</i> (Muller)	1660
<i>Lecane quadridentata</i> (Ehrenberg)	1661
<i>Lepadella acuminata</i> (Ehrenberg)	1646
<i>Lepadella ovalis</i> (Muller)	1647
<i>Lepadella patella</i> (Muller)	1648
<i>Lepadella</i> sp.	1650
<i>Lepadella venifica</i> Myers	1649
<i>Lepidostoma</i> sp.	2228
<i>Leptohyphes dolani</i> Allen	1980
<i>Leptophlebia</i> sp.	1970

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ACCESSION NO.

GENUS & SPECIES:

<i>Libellula</i> sp.	1953
<i>Libellula vibrans</i> Fabricius	1952
<i>Limnephilus</i> sp.	2224
<i>Limnesia</i> sp(p?).	1901
<i>Limnodrilus hoffmeisteri</i> Claparedé	1718
<i>Limnodrilus</i> sp.	1720
<i>Limnodrilus udekemianus</i> Claparedé	1719
<i>Limogonus hesione</i> Kirkaldy	2014
<i>Limogonus</i> sp.	2015
<i>Limonia</i> sp.	2247
<i>Lindia pallida</i> Harring & Myers	1611
<i>Lirceus brachyurus</i> (Harger)	1873
<i>Lophopodella carteri</i> (Hyatt)	1674
<i>Lumbriculus inconstans</i> Smith	1712
<i>Lumbricus variegatus</i> (Müller)	1752
<i>Lymnaea columella</i> Say	1778
<i>Lymnaea humilis</i> Say	1779
<i>Lymnaea</i> sp.	1780
<i>Machronychus glabratus</i> (Say)	2182
<i>Machronychus</i> sp.	2183
<i>Macrobrachium acanthurus</i> (Wiegmann)	1879
<i>Macrobrachium ohione</i> (Smith)	1880
<i>Macromia georgina</i> (Selys)	1943
<i>Macromia</i> sp.	1945
<i>Macromia taeniolata</i> Rambur	1944
<i>Macronema carolina</i> Banks	2215
<i>Macronema</i> sp.	2217
<i>Macronema zebratum</i> (Hagen)	2216
<i>Matus bicarinatus</i> Say	2101
<i>Megapus</i> ? sp.	1895
<i>Menetus dilatatus</i> (Gould)	1791
<i>Mesovelia bisignata</i> Uhler	2027
<i>Mesovelia mulsanti</i> White	2028
<i>Metrobates hesperius</i> Uhler	2016
<i>Micrasema</i> sp.	2227
<i>Microcylloepus pusillus</i> (Leconte)	2184
<i>Micromya ogeecheensis</i> (Conrad)	1815
<i>Micromya vibex</i> Conrad	1816
<i>Micropsectra</i> prob. <i>flavella</i> Zetterstedt	2337

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NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Microvelia sp.	2024
Monommata astia Myers	1607
Monommata sp.	1608
Monopylephorus sp.	1721
Monostyla lunaris (Ehrenberg)	1662
Mooreobdella microstoma (Moore)	1709
Mooreobdella sp(p?).	1710
Musculium spp.	1847
Musculium transversum (Say)	1846
Mystacides nr. sepulchralis (Walker)	2233
Myzobdella lugubris Leidy	1702
Naidium osborni Walton	1734
Naidium sp(p?).	1735
Nais communis Piguet	1736
Nais obtusa (Gervais)	1737
Nais sp(p?).	1738
Nasiaeschna pentacantha (Rambur)	1942
Nectopsyche nr. candida (Hagen)	2234
Nectopsyche nr. exquisita (Walker)	2235
Nectopsyche spp.	2236
Nemocapnia carolina Banks	1989
Nemoura sp.	1987
Neoephemera sp.	1982
Neoperla clymene (Newman)	1999
Neumania sp.	1907
Neureclipsis sp.	2205
Neurocordulia molesta Walsh	1948
Neurocordulia sp.	1949
Notholca longispina (Kellicott)	1651
Notommata pachyura (Gosse)	1609
Notonecta indica Linnaeus	2041
Notonecta irrorata Uhler	2042
Notonecta sp.	2044
Notonecta uhieri Kirkaldy	2043
Nyctiophylax sp.	2206
Ochrotrichia sp.	2220
Oecetis inconspicua ? (Walker)	2239
Oecetis nr. cinerascens (Hagen)	2237
Oecetis prob. eddlestoni Ross	2238

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ACCESSION NO.

GENUS & SPECIES:

Oecetis sp.	2240
Omophron americanum Dejean	2054
Ophidonaïs serpentina (Muller)	1739
Ophidonaïs sp(p?).	1740
Orthocladius nr. carlatus Roback	2286
Orthocladius nr. doreanus Roback	2287
Orthocladius nr. obumbratus Johannsen	2288
Orthocladius spp.	2289
Oxus sp.	1903
Oxyethira sp.	2221
Pachydiplax longipennis (Burmeister)	1954
Palaemonetes kadiakensis Rathbun	1881
Palaemonetes paludosus (Gibbes)	1882
Palaemonetes spp.	1883
Palmaeoxixa buenoi Abbott	2045
Paludicella articulata (Ehrenberg)	1671
Parachironomus nr. abortiva (Malloch)	2317
Parachironomus tenuicaudata (Malloch)	2316
Paracymus digestus (Leconte)	2157
Paracymus spp.	2160
Paracymus subcupreus (Say)	2158
Paracymus suturalis (Leconte)	2159
Paragnetina kansensis Banks	2000
Paragnetina media (Walker)	2001
Paragnetina nr. kansensis Banks	2002
Paragnetina sp.	2003
Paraleptophlebia sp.	1971
Parapoynx sp.	2243
Paratanytarsus spp.	2338
Pectinatella magnifica Leidy	1675
Pelocoris femoratus Palisot de Beauvais	2037
Peloscolex multisetosus Smith	1722
Peloscolex spp.	1724
Peloscolex variegatus Leidy	1723
Peltodytes (Cnemidotus) sp.	2067
Peltodytes festivus (Wehncke)	2059
Peltodytes floridensis ? Sanderson	2065
Peltodytes littoralis Matheson	2060
Peltodytes muticus (Leconte)	2061

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ACCESSION NO.

GENUS & SPECIES:

<i>Peltodytes pedunculatus</i> Blatchley	2062
<i>Peltodytes sexmaculatus</i> Roberts	2063
<i>Peltodytes simplex</i> (Leconte)	2064
<i>Peltodytes</i> sp.	2066
<i>Periscolex?</i> sp.	1749
<i>Perithemis tenera</i> (Say)	1955
<i>Perlesta placida</i> (Hagen)	2004
<i>Perlinella drymo?</i> (Newman)	2005
<i>Phaenonotum estriatum</i> (Say)	2169
<i>Phaenonotum</i> spp.	2170
<i>Phaenopsectra</i> spp.	2318
<i>Phagocata velata</i> (Stringer)	1585
<i>Phanocerus</i> sp.	2185
<i>Phasganophora capitata</i> Pictet	2006
<i>Phylocentropus</i> sp.	2207
<i>Physa crocata</i> Lea	1793
<i>Physa gyrina</i> Say	1794
<i>Physa heterostropha</i> (Say)	1795
<i>Physa</i> sp(p?).	1796
<i>Pilaria</i> sp.	2248
<i>Piona</i> sp.	1905
<i>Pisidium aequilaterale</i> Prime	1848
<i>Pisidium amnicum</i> Muller	1849
<i>Pisidium casertanum</i> (Poli)	1850
<i>Pisidium compressum</i> Prime	1851
<i>Pisidium dubium</i> (Say)	1852
<i>Pisidium milium</i> Held	1853
<i>Pisidium nitidum</i> Jenyns	1854
<i>Pisidium obtusale</i> Pfeiffer	1855
<i>Pisidium</i> sp(p?).	1857
<i>Pisidium walkeri</i> Sterki	1856
<i>Placobdella carolinensis</i> Sawyer & Shelley	1694
<i>Placobdella montifera</i> Moore	1695
<i>Placobdella papillifera</i> (Verrill)	1696
<i>Placobdella parasitica</i> (Say)	1697
<i>Placobdella picta</i> (Verrill)	1698
<i>Placobdella rugosa</i> (Verrill)	1699
<i>Planorbula</i> sp.	1792
<i>Plathemis lydia</i> (Drury)	1956

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NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Platyias patulus</i> (Muller)	1652
<i>Plumatella emarginata</i> Allman?	1677
<i>Plumatella repens</i> Linnaeus	1678
<i>Plumatella</i> sp(p?).	1679
<i>Polyarthra remata</i> (Skorikov)	1625
<i>Polyarthra trigla</i> Ehrenberg	1626
<i>Polyarthra vulgaris</i> (Carlin)	1627
<i>Polycentropus</i> sp.	2208
<i>Polypedilum fallax</i> (Johannsen)	2319
<i>Polypedilum halterale</i> (Coquillett)	2320
<i>Polypedilum illinoense</i> (Malloch)	2321
<i>Polypedilum</i> poss. <i>convictum</i> (Walker)	2323
<i>Polypedilum scalaenum</i> (Schrank)	2322
<i>Polypedilum</i> sp. nr. <i>fallax</i> (Johannsen)	2324
<i>Polypedilum</i> spp.	2325
<i>Pomatiopsis lapidaria</i> (Say)	1771
<i>Potamyia flava</i> (Hagen)	2218
<i>Pottsiella erecta</i> (Potts)	1672
<i>Pristina longiseta leidyi</i> Smith	1742
<i>Pristina</i> sp(p?).	1743
<i>Procambarus hirsutus</i> Hobbs	1885
<i>Procambarus pubescens</i> (Faxon)	1886
<i>Procambarus troglodytes</i> (Le Conte)	1887
<i>Procladius</i> (<i>Procladius</i>) sp.	2263
<i>Procladius</i> (<i>Psilotanypus</i>) <i>bellus</i> (Loew)	2262
<i>Progomphus obscurus</i> Rambur	1938
<i>Prostoma rubrum</i> (Leidy)	1592
<i>Psectrocladius elatus</i> ? Roback	2290
<i>Psectrocladius</i> nr. <i>simulans</i> Johannsen	2292
<i>Psectrocladius</i> poss. <i>nigrus</i> Roback	2291
<i>Psectrocladius</i> sp. 3 Roback	2293
<i>Psectrocladius</i> spp.	2294
<i>Pseudiron</i> sp.	1969
<i>Pseudochironomus</i> nr. <i>richardsoni</i> (Malloch)	2326
<i>Pseudocloeon</i> sp.	1964
<i>Pseudosuccinea columella</i> (Say)	1781
<i>Pteronarcys dorsata</i> (Say)	1990
<i>Pteronarcys</i> spp.	1991
<i>Pycnopsyche</i> sp.	2225

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NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Quadrula sp.	1838
Ranatra australis Hungerford	2032
Ranatra buenoii Hungerford	2033
Ranatra fusca Hungerford	2034
Ranatra kirkaldyi Bueno	2035
Ranatra nigra Herr.-Sch.	2036
Rhagovelia obesa Uhler	2025
Rhanthus bistrigatus Borgstrasser	2102
Rheorthocladus spp.	2295
Rheotanytarsus exigua ? Johanssen	2339
Rheumatobates hungerfordi ? Wiley	2019
Rheumatobates rileyi Bergroth	2017
Rheumatobates sp.	2020
Rheumatobates tenuipes Meinert	2018
Rhynchelmis elrodi (?) Smith & Dickey	1713
Rhynchelmis sp(?)	1715
Rhynchelmis tetratheca (Michaelsen)	1714
Robackia clavigera Townes	2327
Scaridium longicaudum (Muller)	1610
Sialis sp.	2051
Sida crystallina (Muller)	1863
Sigara sp.	2046
Simulium jenningsi (Malloch)	2342
Simulium poss. decorum (Walker)	2343
Simulium spp.	2344
Sinantherina semibullata (Thorpe)	1664
Siphlonurus sp. nr. marshalli Traver.	1957
Slavina appendiculata (d'Udekem)	1744
Somatogyrus virginicus Walker	1772
Sparganophilus sp.	1750
Sphaerium lacustre (Muller)	1858
Sphaerium securis Prime	1859
Sphaerium spp.	1861
Sphaerium transversum (Say)	1860
Spongilla aspinosa Potts	1579
Spongilla fragilis Leidy	1580
Spongilla lacustris (Linnaeus)	1581
Stenacron spp.	1967
Stenelmis antennalis Sanderson	2186

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ACCESSION NO.

GENUS & SPECIES:

<i>Stenelmis beameri</i> Sanderson	2187
<i>Stenelmis bicarinata</i> Leconte	2188
<i>Stenelmis convexula</i> Sanderson	2189
<i>Stenelmis crenata</i> (Say)	2190
<i>Stenelmis decorata</i> Sanderson	2191
<i>Stenelmis douglasensis</i> Sanderson	2192
<i>Stenelmis fuscata</i> (Blatchey)	2193
<i>Stenelmis grossa</i> Sanderson	2194
<i>Stenelmis hungerfordi</i> Sanderson	2195
<i>Stenelmis musgravei</i> Sanderson	2196
<i>Stenelmis parva</i> Sanderson	2197
<i>Stenelmis quadrimaculatus</i> Horn	2198
<i>Stenelmis sinuata</i> Leconte	2199
<i>Stenelmis</i> spp.	2200
<i>Stenochironomus</i> sensu Johannsen	2329
<i>Stenochironomus</i> sp.	2330
<i>Stenochironomus</i> spp.	2328
<i>Stenonema</i> spp.	1968
<i>Stenostomum</i> sp.	1587
<i>Stictochironomus devinctus</i> (Say)	2331
<i>Strophitus undulatus</i> (Say)	1804
<i>Stylaria fossularis</i> Leidy	1745
<i>Stylaria lacustris</i> (Linnaeus)	1746
<i>Suphisellus bicolor</i> (Say)	2069
<i>Suphisellus gibbulus</i> (Aube)	2070
<i>Synchaeta oblonga</i> Ehrenberg	1628
<i>Synchaeta</i> sp.	1629
<i>Synorthocladius semivirens</i> (Kieffer)	2296
<i>Tabanus</i> sp.	2347
<i>Taeniopteryx</i> prob. <i>lonicera</i> Ricker & Ross	1988
<i>Tanypus punctipennis</i> Meigen	2260
<i>Tanypus stellatus</i> Coquillett	2261
<i>Tanytarsus</i> poss. <i>guerla</i> (Roback)	2340
<i>Tanytarsus</i> spp.	2341
<i>Testudinella patina</i> (Hermann)	1663
<i>Tetragoneuria cynosura</i> (Say)	1950
<i>Throscinus</i> spp.	2175
<i>Tipula</i> sp.	2249
<i>Tortopus</i> poss. <i>incertus</i> Traver	1985

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

Tortopus sp.	1986
Trepobates inermis Esaki	2021
Trepobates sp.	2023
Trepobates subnitidus Esaki	2022
Triaenodes sp.	2242
Triaenodes tarda ? Milne	2241
Tribelos jucundus (Walker)	2332
Trichocerca dixonnuttalli (Jennings)	1617
Trichocerca rattus (Muller)	1618
Trichocerca similis Wierzejski	1619
Trichocerca sp.	1622
Trichocerca tenuior (Gosse)	1620
Trichocerca uncinata (Voigt)	1621
Trichocladius sp.	2297
Trichocorixa calva (Say)	2047
Trichocorixa kanza Sailer	2048
Trichocorixa naias Kirkaldy	2049
Trichotria similis (Stenroos)	1653
Tricorythodes spp.	1981
Trochospongilla horrida (Weltner)	1582
Trochospongilla leidyi (Bowerbank)	1583
Tropisternus glaber Herbst	2161
Tropisternus lateralis (Fabricius)	2162
Tropisternus mixtus Leconte	2163
Tropisternus spp.	2166
Tropisternus striolatus Leconte	2164
Tropisternus sublaevis Leconte	2165
Tubifex sp(p?).	1726
Tubifex tubifex (Muller)	1725
Tyrellia sp.	1902
Undet. Genus (? Eriopterini tribe) sp.	2245
Undet. Genus ("Palpomyia group") spp.	2256
Undet. Genus ("Thienemannimyia group") spp.	2272
Undet. Genus nr. Microtendipes sp.	2335
Undet. Genus sp. a Roback	2334
Uniomerus obesus (Lea)	1839
Uniomerus tetralasmus (Say)	1840
Unionicola sp(p?).	1908
Valvata tricarinata (Say)	1766

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

3. MACRO-INVERTEBRATES

NOMENCLATURE

GENUS & SPECIES:

Velia brachialis ? Stal
Villosa delumbis (Conrad)
Villosa vibex (Conrad)
Xenochironomus xenolabis (Kieffer)

ACCESSION NO.

2026
1817
1818
2333

Synonyms of the Phylum Mollusca according to Sam Fuller

<u>Synonym</u>	<u>ID. No.</u>	<u>Computer Nomenclature</u>
Campeloma sp.	1763	Campeloma decisum (Say)
	1764	C. lima Anthony
Amnicola limosa (Say)	1768	Amnicola n. sp. a
Amnicola (Lyogyrus) sp.	1769	A. n. sp. b
Goniobasis caternia (Say)	1775	Goniobasis sp.
Lymnaea humilis Say	1777	Galba humilis modicella (Say)
	1780	Lymnaea sp.
Lymnaea columella Say	1781	Pseudosuccinea columella (Say)
Laevapex fuscus (C. B. Adams)	1783	Ferrissia fusca (C. B. Adams)
Physa sp(p?).	1793	Physa crocata Lea
	1794	P. gyrina Say
	1795	P. heterostopha (Say)
Alasmidonta triangulata (Lea)	1797	Alasmidonta arcula Lea
Anodonta cataracta Say	1800	Anodonta nr. cataracta Say
	1802	A. hallenbackii Lea
Carunculina pulla (Conrad)	1805	Carunculina parva (Barnes)
	1806	C. patrickae Bates
Lampsilis cariosa (Say)	1809	Lampsilis dolabraeformis (Lea)
Lampsilis radiata splendida (Lea)	1810	L. radiata (Lea)
	1813	L. splendida (Lea)
Villosa delumbis (Conrad)	1815	Micromya ogeecheensis (Conrad)
Villosa vibex (Conrad)	1816	M. vibrex (Conrad)
Elliptio lanceolata (Lea)	1819	Elliptio arctatus (Conrad)
	1826	E. fisherianus Lea
	1835	E. productus (Conrad)
Elliptio complanata (Lightfoot)	1821	E. complanatus (Solander)
	1822	E. compalatus northamptonensis (Lea)
	1829	E. hopetonensis Lea
	1836	E. subinflatus (Conrad)
Elliptio congarea (Lea)	1824	E. crassidens (Lamarack)
	1825	E. crassidens congareus Lea
	1827	E. forbesiana (Lea)
	1831	E. incrassatus Lea
	1833	E. lecontianus (Lea)
Elliptio icterina (Conrad)	1834	E. obnubilus (Lea)
	1837	E. tuomeyi (Lea)
Uniomerus obesus (Lea)	1840	Uniomerus tetralasmus (Say)

<u>Synonym</u>	<u>ID. No.</u>	<u>Computer Nomenclature</u>
<i>Corbicula fluminea</i> (Muller)	1842	<i>Corbicula manilensis</i> (Philippi)
<i>Eypera cubensis</i> (Prime)	1844	<i>Eupera singleyi</i> Pilsbry
	1845	<i>E. spp.</i>
<i>Musculium transversum</i> (Say)	1847	<i>Musculium spp.</i>
	1858	<i>Sphaerium lacustre</i> (Muller)
	1859	<i>S. securis</i> Prime
	1860	<i>S. transversum</i> (Say)
	1861	<i>S. spp.</i>
<i>Pisidium compressum</i> Prime	1848	<i>Pisidium aequilaterale</i> Prime
<i>Pisidium amnicum</i> Muller	1850	<i>P. casertanum</i> (Poli)
	1853	<i>P. milium</i> Held
	1854	<i>P. nitidum</i> Jenyns
	1855	<i>P. obtusale</i> Pfeiffer
	1856	<i>P. walkeri</i> Sterki

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

PHYLUM:

Protozoa	3221
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CLASS:

Ciliata	3423
Mastigophora	3222
Sarcodina	3351
Suctorria	3600

ORDER:

Amoebida	3357
Chloromonadida	3300
Chrysomonadida	3223
Cryptomonadida	3243
Dinoflagellida	3304
Euglenoidida	3271
Gymnostomatida	3424
Heliozoida	3403
Heterotrichida	3520
Hymenostomatida	3485
Hypotrichida	3546
Odontostomatida	3576
Oligotrichida	3536
Peritrichida	3581
Phytomonadida	3250
Polymastigida	3344
Proteomyxida	3352
Protomonadida	3323
Rhizomastigida	3315
Suctorida	3601
Testacida	3371
Tintinnida	3542
Trichostomatida	3474

FAMILY:

Acanthocystidae	3404
Acinetidae	3602

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

FAMILY:

<i>Actinophryidae</i>	3409
<i>Amoebidae</i>	3358
<i>Amphileptidae</i>	3425
<i>Amphimonadidae</i>	3324
<i>Anisonemidae</i>	3272
<i>Arcellidae</i>	3372
<i>Aspidiscidae</i>	3547
<i>Astasiidae</i>	3283
<i>Bicosoecidae</i>	3327
<i>Bodonidae</i>	3330
<i>Bursariidae</i>	3521
<i>Carteriidae</i>	3251
<i>Chlamydodontidae</i>	3429
<i>Chlamydomonadidae</i>	3255
<i>Chloromonadidae</i>	3301
<i>Chromulinidae</i>	3224
<i>Ciliophryidae</i>	3412
<i>Clathrellidae</i>	3414
<i>Clathrulinidae</i>	3416
<i>Codosigidae</i>	3337
<i>Coelosomidiidae</i>	3475
<i>Cohnilembidae</i>	3486
<i>Colepidae</i>	3433
<i>Colpodidae</i>	3477
<i>Condylostomidae</i>	3525
<i>Cryptomonadidae</i>	3244
<i>Cystodiniidae</i>	3305
<i>Dendrosomatidae</i>	3608
<i>Didiniidae</i>	3435
<i>Difflugiidae</i>	3381
<i>Dysteriidae</i>	3439
<i>Epalkellidae</i>	3577
<i>Epistylidae</i>	3582
<i>Euglenidae</i>	3291
<i>Euglyphidae</i>	3393
<i>Euplotidae</i>	3549
<i>Frontoniidae</i>	3489
<i>Gromiidae</i>	3398
<i>Gymnodiniidae</i>	3307

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

FAMILY:

Halteriidae	3537
Heterophryidae	3418
Hexamitidae	3345
Holophryidae	3441
Lithocollidae	3420
Loxodidae	3456
Mastigamoebidae	3316
Mayorellidae (=Paramoebidae, in Page)	3367
Metopidae	3527
Microthoracidae	3479
Monadidae	3340
Multiciliidae	3321
Naegleriidae	3369
Nassulidae	3458
Ochromonadidae	3230
Oikomonadidae	3342
Ophrydiidae	3589
Ophryoglenidae	3501
Oxytrichidae	3551
Parameciidae	3503
Peridiniidae	3312
Phacotidae	3262
Philasteridae	3505
Pleuronematidae	3507
Podophryidae	3610
Spathidiidae	3463
Spirostomatidae	3530
Spirozonidae	3483
Stentoridae	3533
Strobilidiidae	3540
Syncryptidae	3240
Tetrahymenidae	3511
Tetramitidae	3347
Tintinnidae	3543
Tracheliididae	3470
Trimastigidae	3349
Vaginicolidae	3591
Vampyrellidae	3353
Volvocidae	3264

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

FAMILY:

Vorticellidae

3596

GENUS & SPECIES:

<i>Acanthamoeba castellanii</i> (Douglas)	1114
<i>Acanthamoeba hyalina</i> (Dob. & O'Conn.)	1115
<i>Acanthamoeba</i> sp.	1116
<i>Acanthocystis aculeata</i> Hart. & Less.	1201
<i>Acanthocystis turfacea</i> (Cart.)	1202
<i>Acineta</i> spp.	1560
<i>Actinophrys sol</i> Ehr.	1207
<i>Actinophrys</i> spp.	1209
<i>Actinophrys vesiculata</i> Pen.	1208
<i>Actinosphaerium eichhorni</i> Ehr.	1210
<i>Amoeba discoidea</i> Schaeff.	1117
<i>Amoeba dubia</i> Schaeff.	1118
<i>Amoeba gorgonia</i> Pen.	1119
<i>Amoeba guttula</i> Duj.	1120
<i>Amoeba limicola</i> Rhumb.	1121
<i>Amoeba proteus</i> (Pallas)	1122
<i>Amoeba radiosa</i> Ehr.	1123
<i>Amoeba</i> spp.	1129
<i>Amoeba spumosa</i> Gruber	1128
<i>Amoeba striata</i> Pen.	1124
<i>Amoeba verrucosa</i> Ehr.	1125
<i>Amoeba vespertilio</i> Pen.	1126
<i>Amoeba villosa</i> Wall. (= <i>Trichamoeba villosa</i> Wall., in Page)	1127
<i>Amphidinium</i> sp.	1046
<i>Amphileptus claparedei</i> Stein	1217
<i>Amphisicella oblonga</i> Schew.	1447
<i>Amphisicella</i> sp.	1448
<i>Anazma brevis</i> Goodr. & Jahn	1561
<i>Ancyromonas contorta</i> (Klebs)	1105
<i>Anisonema acinus</i> Duj.	907
<i>Anisonema emarginatum</i> Stokes	908
<i>Anisonema ovale</i> Klebs	909
<i>Anisonema pusillum</i> Stokes	910
<i>Anisonema truncatum</i> Stein	911
<i>Anthophysis steinii</i> Senn	838

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Anthophysis vegetans</i> (Mull.)	839
<i>Arcella arenaria</i> Greeff	1143
<i>Arcella catinus</i> Pen.	1144
<i>Arcella dentata</i> Ehr.	1145
<i>Arcella discooides</i> Ehr.	1146
<i>Arcella hemisphaerica</i> Perty	1147
<i>Arcella mitrata</i> Leidy	1148
<i>Arcella polypora</i> Pen..	1149
<i>Arcella</i> spp.	1151
<i>Arcella vulgaris</i> Ehr.	1150
<i>Artodiscus saltans</i> Pen.	1197
<i>Ascoglena</i> sp.	948
<i>Aspidisca costata</i> (Duj.)	1430
<i>Aspidisca lynceus</i> Ehr.	1431
<i>Aspidisca marsupialis</i> Pen.	1432
<i>Aspidisca</i> spp.	1435
<i>Aspidisca steini</i> Buddenbrook	1433
<i>Aspidisca sulcata</i> Kahl	1434
<i>Astasia curvata</i> Klebs	930
<i>Astasia dangeardii</i> Lemm.	931
<i>Astasia inflata</i> Duj.	932
<i>Astasia klebsi</i> Lemm.	933
<i>Astasia mobilis</i> (Rehberg)	934
<i>Astasia ocellata</i> Khawkins	935
<i>Astasia</i> spp.	936
<i>Astramoeba</i> sp.	1130
<i>Astrodisculus</i> sp.	1215
<i>Blepharisma coeruleum</i> Gaj.	1402
<i>Blepharisma lateritium</i> (Ehr.)	1403
<i>Bodo alexeieffii</i> Lemm.	1075
<i>Bodo amoebinus</i> Lemm.	1076
<i>Bodo angustus</i> (Duj.)	1077
<i>Bodo caudatus</i> (Duj.)	1078
<i>Bodo celer</i> Klebs	1079
<i>Bodo compressus</i> Lemm.	1080
<i>Bodo edax</i> Klebs	1081
<i>Bodo fusiformis</i> (Stokes)	1082
<i>Bodo globosus</i> Stein	1083
<i>Bodo lens</i> (Mull.)	1084

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Bodo minimus</i> Klebs	1085
<i>Bodo mutabilis</i> Klebs	1086
<i>Bodo obovatus</i> Lemm.	1087
<i>Bodo parvus</i> (Naegler)	1088
<i>Bodo putrinus</i> Lemm.	1089
<i>Bodo repens</i> Klebs	1090
<i>Bodo rostratus</i> (Kent)	1091
<i>Bodo saltans</i> Ehr.	1092
<i>Bodo spp.</i>	1094
<i>Bodo triangularis</i> (Stokes)	1093
<i>Bullinula</i> sp.	1160
<i>Bursaria</i> sp.	1393
<i>Bursaria truncatella</i> Mull.	1392
<i>Bursaridium pseudobursaria</i> Faure-Fr.	1394
<i>Caenomorpha medusula</i> Perty	1397
<i>Campanella</i> sp.	1517
<i>Campanella umbellaria</i> (L.)	1516
<i>Campascus</i> sp.	1161
<i>Carchesium polypinum</i> (L.)	1538
<i>Carteria cordiformis</i> Cart.	874
<i>Carteria ellipsoidalis</i> Bold	875
<i>Carteria globosa</i> Korsch.	876
<i>Carteria</i> spp.	877
<i>Centropyxis aculeata</i> Stein	1162
<i>Centropyxis ecornis</i> (Ehr.)	1163
<i>Ceratium hirundinella</i> (Mull.)	1054
<i>Cercobodo laciniaegegens</i> Krassil.	1058
<i>Cercobodo longicauda</i> (Stein)	1059
<i>Cercobodo</i> sp.	1060
<i>Cercomonas crassicauda</i> Duj.	1095
<i>Cercomonas longicauda</i> Duj.	1096
<i>Chaetospira remex</i> Hudson	1449
<i>Chilodonella caudata</i> (Stokes)	1227
<i>Chilodonella cucullulus</i> (Mull.)	1228
<i>Chilodonella fluviatilis</i> (Stokes)	1229
<i>Chilodonella</i> spp.	1231
<i>Chilodonella uncinata</i> (Ehr.)	1230
<i>Chilodontopsis muscorum</i> Kahl	1280
<i>Chilodontopsis</i> sp.	1282

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

GENUS & SPECIES:

	<u>ACCESSION NO.</u>
<i>Chilodontopsis vorax</i> (Stokes)	1281
<i>Chilomonas paramecium</i> Ehr.	862
<i>Chilophrya labiata</i> (Edmondson)	1247
<i>Chlamydodon</i> sp.	1232
<i>Chlamydomonas angulosa</i> Dill.	880
<i>Chlamydomonas compressa</i> Pasch.	881
<i>Chlamydomonas conica</i> Dang.	882
<i>Chlamydomonas depauperata</i> Pasch.	883
<i>Chlamydomonas globosa</i> Snow	884
<i>Chlamydomonas gracilis</i> Snow	885
<i>Chlamydomonas monadina</i> Stein	886
<i>Chlamydomonas pomiformis</i> Pasch.	887
<i>Chlamydomonas regularis</i> Korsch.	888
<i>Chlamydomonas sectilis</i> Korsch.	889
<i>Chlamydomonas snowiae</i> Printz	890
<i>Chlamydomonas</i> spp.	891
<i>Chlamydomyxa montana</i> Lankester	1109
<i>Chlamydophrys stercorea</i> Cienk.	1152
<i>Chlorogonium elegans</i> Playf.	892
<i>Chlorogonium</i> sp.	893
<i>Chromulina ovalis</i> Klebs	824
<i>Chromulina rosanoffii</i> Butschli	825
<i>Chromulina</i> spp.	826
<i>Chroomonas norstedtii</i> Hansgirg	863
<i>Chrysosphaerella</i> spp.	827
<i>Ciliophrys infusionum</i> Cienk.	1211
<i>Cinetochilum margaritaceum</i> Perty	1328
<i>Cladomonas fruticulosa</i> Stein	1068
<i>Cladonema pauperum</i> Pasch.	1562
<i>Clathrella foreli</i> Pen.	1212
<i>Clathrulina elegans</i> (Leidy)	1213
<i>Climacostomum</i> sp.	1411
<i>Clypeolina marginata</i> Pen.	1164
<i>Coccoonas orbicularis</i> Stein	894
<i>Cochliopodium bilimbosum</i> (Auerbach)	1153
<i>Cochliopodium digitatum</i> (Greeff)	1154
<i>Codonella cratera</i> (Leidy)	1427
<i>Codonoeca</i> sp.	1071
<i>Cohnilembus fusiformis</i> (Cohn)	1326

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Coleps bicuspis</i> Noland	1234
<i>Coleps elongatus</i> Ehr.	1235
<i>Coleps hirtus</i> (Mull.)	1236
<i>Coleps octospinus</i> Noland	1237
<i>Coleps</i> spp.	1239
<i>Coleps striatus</i> Smith	1238
<i>Colpidium campylum</i> (Stokes)	1373
<i>Colpidium colpoda</i> (Ehr.)	1374
<i>Colpoda aspera</i> Kahl	1314
<i>Colpoda cucullus</i> Mull.	1315
<i>Colpoda henneguyi</i> Fabre-Dom.	1316
<i>Colpoda inflata</i> (Stokes)	1317
<i>Colponema loxodes</i> Stein	1097
<i>Colponema</i> sp.	1098
<i>Condyllostoma</i> sp.	1396
<i>Copromonas subtilis</i> Dob.	937
<i>Cothurnia annulata</i> Stokes	1525
<i>Cothurnia ovata</i> Fromentel	1526
<i>Cothurnia pupa</i> Eichwald	1527
<i>Cothurnia</i> sp.	1528
<i>Cranotheridium taeniatum</i> Schew.	1290
<i>Cristigera phoenix</i> Pen.	1359
<i>Croblylura pelagica</i> Andre	1248
<i>Cryptochrysis commutata</i> Pasch.	864
<i>Cryptochrysis polychrysis</i> Pasch.	865
<i>Cryptoglena pigra</i> Ehr.	949
<i>Cryptoglena</i> spp.	950
<i>Cryptomonas compressa</i> Pasch.	866
<i>Cryptomonas erosa</i> Ehr.	867
<i>Cryptomonas nasuta</i> Pasch.	868
<i>Cryptomonas obovoidea</i> Pasch.	869
<i>Cryptomonas ovata</i> Ehr.	870
<i>Cryptomonas reflexa</i> Marsson	871
<i>Cryptomonas</i> spp.	872
<i>Cucurbitella mespiliformis</i> Pen.	1165
<i>Cyathomonas truncata</i> Ehr.	873
<i>Cyclidium brandoni</i> Kahl	1360
<i>Cyclidium citrullus</i> Cohn	1361
<i>Cyclidium elongatum</i> Schew.	1362

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Cyclidium glaucoma</i> Mull.	1363
<i>Cyclidium granulosum</i> Kahl	1364
<i>Cyclidium instabile</i> Clark	1365
<i>Cyclidium litomesum</i> Stokes	1366
<i>Cyclidium musicola</i> Kahl	1367
<i>Cyclidium pellucidum</i> Kahl	1368
<i>Cyclidium spp.</i>	1370
<i>Cyclidium versatile</i> Pen.	1369
<i>Cyclogramma</i> sp.	1284
<i>Cyclogramma trichocystis</i> (Stokes)	1283
<i>Cyclonexis annularis</i> Stokes	840
<i>Cyclotrichium sphaericum</i> Faure-Fr.	1240
<i>Cyphoderia ampulla</i> (Ehr.)	1185
<i>Cyrtolophosis elongata</i> Schew.	1329
<i>Cyrtolophosis mucicola</i> Stokes	1330
<i>Dallingeria drysdali</i> Kent	1108
<i>Dendomonas virgaria</i> (Weisse)	841
<i>Diaphoropodon</i> sp.	1166
<i>Dichilum cuneiforme</i> Schew.	1375
<i>Didinium balbianii</i> (Fabre-Dom.)	1241
<i>Didinium nasutum</i> (Mull.)	1242
<i>Diffugia areolata</i> Ehr.	1167
<i>Diffugia constricta</i> Ehr.	1168
<i>Diffugia corona</i> Wall.	1169
<i>Diffugia globulosa</i> Duj.	1170
<i>Diffugia globulus</i> (Ehr.)	1171
<i>Diffugia gramen</i> Pen.	1172
<i>Diffugia lanceolata</i> Pen.	1173
<i>Diffugia oblonga</i> Ehr.	1174
<i>Diffugia spiralis</i> Leclerc	1175
<i>Diffugia</i> spp.	1178
<i>Diffugia tuberculata</i> Wall.	1176
<i>Diffugia urceolata</i> Cart.	1177
<i>Dileptus americanus</i> Kahl	1303
<i>Dileptus anser</i> Mull.	1304
<i>Dileptus bivacuolatus</i> da Cunha	1305
<i>Dileptus cygnus</i> (Clap. & Lachm.)	1306
<i>Dileptus gracilis</i> Kahl	1307
<i>Dileptus monilatus</i> (Stokes)	1308

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GENUS & SPECIES:

Dileptus sp.	1309
Dinamoeba mirabilis L.	1131
Dinamoeba sp.	1132
Dinema griseolum Perty	912
Dinobryon sertularia Ehr.	842
Dinomonas sp.	1070
Dinomonas vorax Kent	1069
Diplophrys sp.	1198
Disematosstoma sp.	1331
Enchelydium fusidens Kahl	1291
Enchelydium virens Kahl	1292
Enchelys arcuata Clap. & Lachm.	1249
Enchelys gasterosteus Kahl	1250
Enchelys variabilis Svec	1251
Entosiphon ovatum Stokes	913
Entosiphon sulcatum (Duj.)	914
Epalkysis (=Epalkella, in Kudo) mirabilis (Roux)	1513
Epistylus plicatilis Ehr.	1518
Epistylus sp.	1519
Espejoia musicola Pen.	1332
Espejoia sp.	1333
Eudorina elegans Ehr.	899
Eudorina sp.	900
Euglena acus Ehr.	951
Euglena acutissima Lemm.	952
Euglena agilis Cart.	953
Euglena caudata Hubner	954
Euglena chlamydophora Mainx	955
Euglena clavata Skuja	956
Euglena deses Ehr.	957
Euglena ehrenbergi Klebs	958
Euglena fusca (Klebs)	959
Euglena geniculata (Duj.)	960
Euglena gracilis Klebs	961
Euglena granulata Lemm.	962
Euglena haematodes Ehr.	963
Euglena intermedia Schmitz	964
Euglena klebsi Mainx	965
Euglena limnophila Lemm.	966

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ACCESSION NO.

GENUS & SPECIES:

<i>Euglena minima</i> France	967
<i>Euglena minuta</i> Prescott	968
<i>Euglena mutabilis</i> mainxi Gojd.	970
<i>Euglena mutabilis</i> Schmitz	969
<i>Euglena oxyuris</i> Schmarda	971
<i>Euglena pisciformis</i> Klebs	972
<i>Euglena platydesma</i> Skuja	973
<i>Euglena proxima</i> Dang.	974
<i>Euglena rubra</i> Hardy	975
<i>Euglena sanguinea</i> Ehr.	976
<i>Euglena sociabilis</i> Dang.	977
<i>Euglena spirogyra</i> Ehr.	978
<i>Euglena splendens</i> Dang.	979
<i>Euglena</i> spp.	985
<i>Euglena terricola</i> (Dang.)	980
<i>Euglena torta</i> Korsch.	981
<i>Euglena tripteris</i> (Duj.)	982
<i>Euglena truncata</i> Walton	983
<i>Euglena viridis</i> Ehr.	984
<i>Euglenopsis vorax</i> Klebs	915
<i>Euglypha acanthophora</i> Ehr.	1186
<i>Euglypha cristata</i> Leidy	1187
<i>Euglypha elongata</i> Leidy	1188
<i>Euglypha mucronata</i> Leidy	1189
<i>Euglypha scutigera</i> Pen.	1190
<i>Euglypha</i> spp.	1191
<i>Euplotes aediculatus</i> Pierson	1436
<i>Euplotes affinis</i> Duj.	1437
<i>Euplotes carinatus</i> Stokes	1438
<i>Euplotes eurystomus</i> Wrzes.	1439
<i>Euplotes leticiensis</i> Bovee	1440
<i>Euplotes moebiusi</i> Kahl	1441
<i>Euplotes muscicola</i> Kahl	1442
<i>Euplotes patella</i> (Mull.)	1443
<i>Euplotes plumipes</i> Stokes	1444
<i>Euplotes</i> spp.	1446
<i>Euplotes taylori</i> Garnjobst	1445
<i>Eutreptia viridis</i> Perty	986
<i>Flabellula velata</i> Jahn	1133

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ACCESSION NO.

GENUS & SPECIES:

<i>Frontonia acuminata</i> (Ehr.)	1334
<i>Frontonia acuminata angusta</i> Kahl	1335
<i>Frontonia depressa</i> (Stokes)	1336
<i>Frontonia elliptica</i> Beardsley	1337
<i>Frontonia leucas</i> Ehr.	1338
<i>Frontonia</i> spp.	1339
<i>Frontonialla complanata</i> Wetzel	1340
<i>Gastrostyla muscorum</i> Kahl	1450
<i>Gastrostyla steini</i> Engelmann	1451
<i>Gigantochloris</i> sp.	895
<i>Glaucoma avellana</i> Kahl	1376
<i>Glaucoma reniformis</i> Schew.	1377
<i>Glaucoma scintillans</i> Ehr.	1378
<i>Glaucoma setosa</i> Schew.	1379
<i>Glaucoma</i> spp.	1380
<i>Glenodinium cinctum</i> Ehr.	1044
<i>Glenodinium neglectum</i> Schilling	1045
<i>Gonium formosum</i> Pasch.	901
<i>Gonium pectorale</i> Mull.	902
<i>Gonostomum affine</i> (Stein)	1452
<i>Gonostomum strenuum</i> (Engelmann)	1453
<i>Gonyostomum semen</i> Diesing	1041
<i>Gonyostomum</i> sp.	1042
<i>Gromia nigricans</i> (Pen.)	1199
<i>Gymnodinium aeruginosum</i> Stein	1047
<i>Gymnodinium fuscum</i> Ehr.	1048
<i>Gymnodinium palustre</i> Schilling	1049
<i>Gymnodinium rotundatum</i> Klebs	1050
<i>Gymnodinium</i> spp.	1051
<i>Gyrodinium hyalinum</i> Schilling	1052
<i>Hallezia brachypoda</i> (Stokes)	1563
<i>Halteria grandinella</i> (Mull.)	1422
<i>Hartmannella hyalina</i> Dang.	1134
<i>Heleopera</i> sp.	1179
<i>Hemicyclostyla</i> sp.	1455
<i>Hemicyclostyla sphagni</i> Stokes	1454
<i>Heterochromonas</i> (=Monas, in Pascher) sp.	1104
<i>Heteronema acus</i> (Ehr.)	916
<i>Heteronema acutissimum</i> Lemm.	917

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GENUS & SPECIES:

<i>Heteronema klebsii</i> Senn	918
<i>Heteronema mutabile</i> (Stokes)	919
<i>Heteronema</i> spp.	921
<i>Heteronema tremulum</i> Zach.	920
<i>Heterophrys myriopoda</i> Archer	1214
<i>Histrio</i> (=Oxytricha, in Kudo) <i>histrio</i> (Mull.)	1456
<i>Holophrya simplex</i> Schew.	1252
<i>Holophrya</i> spp.	1253
<i>Holosticha discocephalus</i> Kahl	1457
<i>Holosticha kessleri</i> (Wrzes.)	1458
<i>Holosticha</i> spp.	1461
<i>Holosticha vernalis</i> Stokes	1459
<i>Holosticha violacea</i> Kahl	1460
<i>Homalozoon vermiculare</i> Stokes	1293
<i>Hyalobryon</i> sp.	843
<i>Hyalosphenia</i> sp.	1155
<i>Kahlia acrobates</i> Horv.	1462
<i>Keronopsis flavicans</i> Kahl	1463
<i>Keronopsis muscorum</i> Kahl	1464
<i>Keronopsis similis</i> (Stokes)	1465
<i>Keronopsis</i> sp.	1466
<i>Khawinea halli</i> Jahn & McKib.	987
<i>Khawinea</i> sp.	988
<i>Lacrymaria olor</i> (Mull.)	1254
<i>Lacrymaria</i> sp.	1255
<i>Lagynophrya mutans</i> Kahl	1256
<i>Lagynophrya simplex</i> Kahl	1257
<i>Lembadion bullinum</i> Perty	1341
<i>Lembadion lucens</i> (Mast)	1342
<i>Lembadion magnum</i> (Stokes)	1343
<i>Lembadion</i> spp.	1344
<i>Lepocinclus acicularis</i> France	989
<i>Lepocinclus capito</i> Wehrle	990
<i>Lepocinclus caudata</i> Da Cunha	991
<i>Lepocinclus fusiformis</i> (Cart.)	992
<i>Lepocinclus ovum</i> (Ehr.)	993
<i>Lepocinclus</i> sp.	997
<i>Lepocinclus sphagnophila</i> Lemm.	994
<i>Lepocinclus steinii</i> Lemm.	995

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GENUS & SPECIES:

<i>Lepocinclus texta</i> (Duj.)	996
<i>Leptopharynx</i> spp. (includes <i>Trichopelma</i>)	1318
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) <i>armillatus</i> Pen.	1218
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) <i>cygnus</i> (Mull.)	1219
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) <i>fasciola</i> (Ehr.)	1220
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) spp. (includes <i>Hemiophrys</i>)	1223
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) <i>trichocystis</i> Stokes	1221
<i>Lionotus</i> (= <i>Litonotus</i> , in Kudo) <i>triqueter</i> Pen.	1222
<i>Lithocolla globosa</i> Schulze	1216
<i>Lobomonas</i> sp.	896
<i>Loxocephalus plagioides</i> (Stokes)	1381
<i>Loxodes magnus</i> Stokes	1275
<i>Loxodes rostrum</i> Ehr.	1276
<i>Loxodes</i> sp.	1279
<i>Loxodes striatus</i> Pen.	1277
<i>Loxodes vorax</i> Stokes	1278
<i>Loxophyllum meleagris</i> Duj.	1224
<i>Loxophyllum</i> spp.	1226
<i>Loxophyllum utriculariae</i> (Pen.)	1225
<i>Malacophrys rotans</i> Kahl	1345
<i>Mallomonas acaroides</i> Perty	828
<i>Mallomonas litomosa</i> Stokes	829
<i>Mallomonas</i> sp.	830
<i>Marsupiogaster</i> sp.	922
<i>Massartia musei</i> (Danysz)	1053
<i>Mastigamoeba longifilum</i> Stokes	1061
<i>Mastigamoeba radiospora</i> Lackey	1062
<i>Mastigamoeba trichophora</i> Laut.	1063
<i>Mastigella radicula</i> (Moroff)	1064
<i>Mastigella simplex</i> (Kent)	1065
<i>Mastigosphaera gobii</i> Schew.	903
<i>Mayorella</i> sp.	1141
<i>Menodium incurvum</i> (Fres.)	938
<i>Menodium pellucidum</i> Perty	939
<i>Mesodinium acarus</i> Stein	1243
<i>Mesodinium pulex</i> (Clap. & Lachm.)	1244
<i>Mesodinium</i> spp.	1245
<i>Metacineta mystacina</i> (Ehr.)	1567
<i>Metanema variabile</i> Klebs	923

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GENUS & SPECIES:

Metopus es Mull.	1398
Metopus fuscus Kahl	1399
Metopus mucicola Kahl	1400
Metopus striatus McMurrich	1401
Microgromia socialis (Archer)	1200
Microregma auduboni (Smith)	1258
Microthorax elegans Kahl	1319
Microthorax sp.	1321
Microthorax tridentatus Pen.	1320
Monas amoebina Meyer	845
Monas arhabdomonas (Fisch)	844
Monas dangeardii Lemm.	846
Monas elongata (Stokes)	847
Monas guttula Ehr.	848
Monas sociabilis Meyer	849
Monas socialis (Kent)	850
Monas spp.	853
Monas vestita (Stokes)	851
Monas vulgaris (Cienk.)	852
Monochilum frontatum Schew.	1382
Monochilum ovale (Schew.)	1383
Monochilum tesselatum Pen.	1384
Monosiga ovata Kent	1102
Multicilia lacustris Laut.	1067
Naegleria spp.	1142
Nassula aurea Ehr.	1285
Nassula gracilis Kahl	1286
Nassula ornata Ehr.	1287
Nassula spp.	1288
Nebela caudata Leidy	1192
Nebela collaris (Ehr.)	1193
Nebela spp.	1195
Nebela vitrea Wailes	1194
Notosolenus apocamptus Stokes	924
Notosolenus sinuatus Stokes	925
Nuclearia delicatula (Cash)	1110
Nuclearia simplex Cienk.	1111
Ochromonas crenata Klebs	854
Ochromonas mutabilis Klebs	855

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GENUS & SPECIES:

Ochromonas spp.	856
Oikomonas mutabilis Kent	831
Oikomonas obliqua Kent	832
Oikomonas quadrata Kent	833
Oikomonas socialis Moroff	834
Oikomonas spp.	836
Oikomonas termo (Ehr.)	835
Onychodromopsis flexilis Maupas	1467
Onychodromus grandis Stein	1468
Opercularia plicatilis Stokes	1520
Ophrydium vernalis (Stokes)	1524
Ophryoglena atra Lieberkuhn	1350
Opisthostyla pusilla Stokes	1521
Opisthotricha (=Oxytricha, in Kudo) procera Kahl	1469
Opisthotricha (=Oxytricha, in Kudo) similis Engelmann	1470
Orthodonella sp.	1289
Oxytricha bifaria Stokes	1471
Oxytricha chlorelligera Kahl	1472
Oxytricha fallax Stein	1473
Oxytricha furcata Smith	1474
Oxytricha hymenostoma Stokes	1475
Oxytricha ludibunda Stokes	1476
Oxytricha minor Kahl	1477
Oxytricha setigera Stokes	1478
Oxytricha sp.	1480
Oxytricha tricornis Milne	1479
Pamphagus mutabilis Bailey	1156
Pandorina morum (Mull.)	904
Paracineta spp.	1568
Paradileptus robustus Wenrich	1310
Paraholosticha herbicola Kahl	1481
Paramastix conifera Skuja	1106
Paramecium aurelia Ehr.	1351
Paramecium bursaria (Ehr.)	1352
Paramecium calkinsi Woodruff	1353
Paramecium caudatum Ehr.	1354
Paramecium multimicronucleatum Powers & Mitchell	1355
Paramecium putrinum Clap. & Lachm.	1356
Paramecium trichium Stokes	1357

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ACCESSION NO.

GENUS & SPECIES:

<i>Paruroleptus</i> sp.	1482
<i>Pelodinium reniforme</i> Laut.	1514
<i>Pelomyxa carolinensis</i> Wilson	1135
<i>Pelomyxa palustris</i> Greeff	1136
<i>Pelomyxa</i> spp.	1137
<i>Penardiella crassa</i> (Pen.)	1294
<i>Peranema curvicauda</i> Skuja	926
<i>Peranema granulifera</i> Pen.	927
<i>Peranema trichophorum</i> (Ehr.)	928
<i>Peridinium cinctum</i> (Mull.)	1055
<i>Peridinium</i> sp.	1057
<i>Peridinium tabulatum</i> (Clap. & Lachm.)	1056
<i>Petalomonas curvata</i> Skuja	940
<i>Petalomonas mediocanellata</i> Stein	941
<i>Petalomonas sexlobata</i> Klebs	942
<i>Petalomonas sinuata</i> Stein	943
<i>Petalomonas steinii</i> Klebs	944
<i>Phacotus lenticularis</i> (Ehr.)	898
<i>Phacus acuminata</i> Stokes	998
<i>Phacus alata</i> Klebs	999
<i>Phacus anacoelus</i> Stokes	1000
<i>Phacus brevicaudata</i> (Klebs)	1001
<i>Phacus caudata</i> Hubner	1002
<i>Phacus clavata</i> Dang.	1003
<i>Phacus helicoides</i> Poch.	1004
<i>Phacus longicauda</i> (Ehr.)	1005
<i>Phacus norstedtii</i> Lemm.	1006
<i>Phacus orbicularis</i> Hubner	1007
<i>Phacus oscillans</i> Klebs	1008
<i>Phacus pleuronectes</i> Mull.	1009
<i>Phacus pyrum</i> (Ehr.)	1010
<i>Phacus</i> spp.	1016
<i>Phacus stokesii</i> Lemm.	1011
<i>Phacus striata</i> France	1012
<i>Phacus torta</i> Lemm.	1013
<i>Phacus tripteris</i> (Duj.)	1014
<i>Phacus triqueter</i> (Ehr.)	1015
<i>Phascolodon</i> sp.	1233
<i>Philaster armata</i> (Kahl)	1358

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ACCESSION NO.

GENUS & SPECIES:

<i>Phryganella paradoxa</i> Pen.	1180
<i>Phyllomitus amylophagus</i> Klebs	1099
<i>Pithothorax simplex</i> Kahl	1259
<i>Placus ovum</i> Kahl	1260
<i>Placus</i> sp.	1262
<i>Placus striatus</i> France	1261
<i>Plagiophrys parvipunctata</i> Pen.	1157
<i>Platycola</i> sp.	1529
<i>Platydorina caudata</i> Kofoid	905
<i>Platynematum sociale</i> (Pen.)	1346
<i>Platynematum</i> sp.	1347
<i>Platyophrya lata</i> Kahl	1263
<i>Pleodorina</i> sp.	906
<i>Pleuromonas jaculans</i> (Perty)	1100
<i>Pleuronema crassum</i> Duj.	1371
<i>Pleuronema</i> sp.	1372
<i>Pleurotricha grandis</i> Stein	1483
<i>Pleurotricha lanceolata</i> (Ehr.)	1484
<i>Podophrya fixa</i> Mull.	1569
<i>Podophrya</i> sp.	1570
<i>Polytomella</i> sp.	878
<i>Pompholyxophrys punicea</i> Archer	1203
<i>Pontigulasia elisa</i> Schouteden	1181
<i>Prorodon discolor</i> Ehr.	1264
<i>Prorodon griseus</i> (Clap. & Lachm.)	1265
<i>Prorodon margaritifer</i> Clap. & Lachm.	1266
<i>Prorodon minutus</i> Kahl	1267
<i>Prorodon ovum</i> Ehr.	1268
<i>Prorodon</i> sp.	1269
<i>Pseudochlamys</i> sp.	1158
<i>Pseudodiffugia fulva</i> Archer	1182
<i>Pseudodiffugia gracilis</i> Schlum.	1183
<i>Pseudodiffugia</i> spp.	1184
<i>Pseudoglaucoma muscorum</i> Kahl	1385
<i>Pseudomicrothorax agilis</i> Mermod	1322
<i>Pseudomicrothorax dubius</i> (Maupas)	1323
<i>Pseudomicrothorax</i> sp.	1324
<i>Pseudoprorodon farctus</i> (Clap. & Lachm.)	1313
<i>Pteridomonas</i> sp.	1066

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ACCESSION NO.

GENUS & SPECIES:

<i>Pyxidicula operculata</i> (Agardh)	1159
<i>Pyxidium</i> sp.	1522
<i>Raphidiophrys pallida</i> Schulze	1204
<i>Raphidiophrys viridis</i> (Leidy)	1205
<i>Raphidocystis</i> sp.	1206
<i>Rhabdostyla</i> sp.	1523
<i>Rhynchomonas nasuta</i> (Stokes)	1101
<i>Salpingoeca buetschlii</i> Lemm.	1072
<i>Salpingoeca frequentissima</i> Lemm.	1073
<i>Salpingoeca</i> spp.	1074
<i>Saprodinium dentatum</i> Laut.	1515
<i>Saprophilus muscorum</i> Kahl	1386
<i>Scourfieldia complanata</i> West	897
<i>Scytononas pusilla</i> (Stein)	945
<i>Spathidiooides sulcata</i> Brodsky	1295
<i>Spathidium brunneum</i> Kahl	1296
<i>Spathidium caudatum</i> Wetzel	1297
<i>Spathidium faurei</i> Kahl	1298
<i>Spathidium gibbum</i> Kahl	1299
<i>Spathidium</i> sp.	1302
<i>Spathidium</i> sp. (nr. <i>frontinale</i> Pen.)	1301
<i>Spathidium spathula</i> Mull.	1300
<i>Spermatozopsis exultans</i> Korsch.	879
<i>Sphaeroeca</i> sp.	1103
<i>Sphaerophrya magna</i> Maupas	1571
<i>Sphaerophrya soliformis</i> Laut.	1572
<i>Sphaerophrya</i> sp.	1574
<i>Sphaerophrya stentoris</i> Maupas	1573
<i>Sphaleromantis</i> sp.	837
<i>Sphenomonas teres</i> (Stein)	946
<i>Spirostomum ambiguum</i> Ehr.	1404
<i>Spirostomum filum</i> (Ehr.)	1405
<i>Spirostomum intermedium</i> Kahl	1406
<i>Spirostomum loxodes</i> Stokes	1407
<i>Spirostomum minus</i> Roux	1408
<i>Spirostomum</i> sp.	1410
<i>Spirostomum teres</i> Clap. & Lachm.	1409
<i>Spirozoa caudata</i> Kahl	1325
<i>Steinia (=Oxytricha, in Kudo) gracilis</i> Kahl	1485

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ACCESSION NO.

GENUS & SPECIES:

Stentor amethystinus Leidy	1412
Stentor coeruleus Ehr.	1413
Stentor igneus Ehr.	1414
Stentor mulleri (Borg)	1415
Stentor multiformis (Mull.)	1416
Stentor niger (Mull.)	1417
Stentor polymorphus (Mull.)	1418
Stentor pyriformis Johns.	1419
Stentor roeseli Ehr.	1420
Stentor sp.	1421
Stichotricha aculeata Wrzes.	1486
Stichotricha intermedia Froud	1487
Stichotricha secunda Perty	1488
Stichotricha spp.	1489
Stokesiella sp.	857
Strobilidium gyrans (Stokes)	1425
Strobilidium velox Faure-Fr.	1426
Strombidium fallax (Zach.)	1423
Strombidium spp.	1424
Strongylidium crassum Sterki	1490
Styloynchia curvata Kahl	1491
Styloynchia mytilus (Mull.)	1492
Styloynchia notophora Stokes	1493
Styloynchia pustulata (Ehr.)	1494
Styloynchia putrina Stokes	1495
Styloynchia sp.	1496
Stylopysis mucicola Bolochonzew	858
Syncrypta sp.	859
Synura adamsi Smith	860
Synura uvella Ehr.	861
Tachysoma pellionella (Mull.)	1497
Tetrahymena geleii Furg.	1387
Tetrahymena patula (Ehr.)	1388
Tetrahymena pyriformis Ehr.	1389
Tetrahymena sp.	1391
Tetrahymena vorax (Kid., Lilly & Claff)	1390
Tetramitus rostratus Perty	1107
Thurcicola folliculata (Mull.)	1530
Thylacidium truncatum Schew.	1395

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Tintinnopsis cylindrata</i> (Entz)	1428
<i>Tintinnopsis</i> sp.	1429
<i>Tokophrya infusionum</i> (Stein)	1564
<i>Tokophrya</i> sp.	1565
<i>Trachelius ovum</i> Ehr.	1311
<i>Trachelius</i> sp.	1312
<i>Trachelocerca phoenicopterus</i> Cohn	1270
<i>Trachelocerca</i> sp.	1271
<i>Trachelomonas acanthostoma</i> Stokes	1017
<i>Trachelomonas acuminata</i> (Schmarda)	1018
<i>Trachelomonas annulata</i> Daday	1019
<i>Trachelomonas armata</i> (Ehr.)	1020
<i>Trachelomonas crebea</i> Kellicott	1021
<i>Trachelomonas dubia</i> Deflandre	1022
<i>Trachelomonas ensifera</i> Daday	1023
<i>Trachelomonas hispida</i> (Perty)	1024
<i>Trachelomonas horrida</i> Palmer	1025
<i>Trachelomonas intermedia</i> Dang.	1026
<i>Trachelomonas oblonga</i> Lemm.	1027
<i>Trachelomonas perforata</i> Awerinzew	1028
<i>Trachelomonas piscatoris</i> (Fisch.)	1029
<i>Trachelomonas raciborskii</i> Wolosz.	1030
<i>Trachelomonas rugulosa</i> Stein	1031
<i>Trachelomonas saccata</i> Lemm.	1032
<i>Trachelomonas setosa</i> Zytkoff	1033
<i>Trachelomonas spiculifera</i> Palmer	1034
<i>Trachelomonas</i> spp.	1040
<i>Trachelomonas urceolata</i> Stokes	1035
<i>Trachelomonas vermiculosa</i> Palmer	1036
<i>Trachelomonas verrucosa</i> Stokes	1037
<i>Trachelomonas volvocina</i> Ehr.	1038
<i>Trachelomonas zmiewika</i> Swir.	1039
<i>Trachelophyllum clavatum</i> Stokes	1272
<i>Trachelophyllum pusillum</i> (Clap. & Lachm.)	1273
<i>Trentonia flagellata</i> Stokes	1043
<i>Trichophyra epistylidis</i> Clap. & Lachm.	1566
<i>Trinema enchelys</i> (Ehr.)	1196
<i>Trochilia palustris</i> Stein	1246
<i>Tropidoscaphus octocostatus</i> Stein	929

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Urceolus sabulosus</i> (Stokes)	947
<i>Urocentrum turbo</i> (Mull.)	1348
<i>Uroleptus caudatus</i> (Clap. & Lachm.)	1498
<i>Uroleptus dispar</i> Stokes	1499
<i>Uroleptus gibbus</i> (Clap. & Lachm.)	1500
<i>Uroleptus limnetis</i> Stokes	1501
<i>Uroleptus longicaudatus</i> Stokes	1502
<i>Uroleptus mobilis</i> Engelmann	1503
<i>Uroleptus muscorum</i> Kahl	1504
<i>Uroleptus piscis</i> (Mull.)	1505
<i>Uroleptus</i> sp.	1506
<i>Uronema</i> sp.	1327
<i>Urosoma acuminata</i> (Stokes)	1507
<i>Urosoma caudata</i> (Stokes)	1508
<i>Urosoma</i> spp.	1509
<i>Urostyla</i> spp.	1512
<i>Urostyla trichogaster</i> Kahl	1510
<i>Urostyla viridis</i> Stein	1511
<i>Urotricha agilis</i> (Stokes)	1274
<i>Urozona butschlii</i> Schew.	1349
<i>Vaginicola annulata</i> Stokes	1531
<i>Vaginicola inclinata</i> From.	1532
<i>Vaginicola ingenita</i> Mull.	1533
<i>Vaginicola leptosoma</i> Stokes	1534
<i>Vaginicola longipes</i> Stokes	1535
<i>Vaginicola</i> spp.	1537
<i>Vaginicola tincta</i> Ehr.	1536
<i>Vahlkampfia "limax type"</i>	1139
<i>Vahlkampfia limax</i> (Duj.)	1138
<i>Vahlkampfia</i> spp.	1140
<i>Vampyrella lateritia</i> (Fres.)	1112
<i>Vampyrella</i> sp.	1113
<i>Vorticella aequilata</i> Kahl	1539
<i>Vorticella campanula</i> Ehr.	1540
<i>Vorticella citrina</i> Mull.	1541
<i>Vorticella communis</i> From.	1542
<i>Vorticella convallaria</i> (L.)	1543
<i>Vorticella cupifera</i> Kahl	1544
<i>Vorticella extensa</i> Kahl	1545

APPENDIX A. SAVANNAH RIVER TAXONOMIC ACCESSION LIST

4. PROTOZOA

NOMENCLATURE

ACCESSION NO.

GENUS & SPECIES:

<i>Vorticella floridensis</i> Stokes	1546
<i>Vorticella kenti</i> Kahl	1547
<i>Vorticella limnetis</i> Stokes	1548
<i>Vorticella microstoma</i> Ehr.	1549
<i>Vorticella monilata</i> Tatem	1550
<i>Vorticella picta</i> (Ehr.)	1551
<i>Vorticella similis</i> Stokes	1552
<i>Vorticella sphaerica</i> d'Udekem	1553
<i>Vorticella spp.</i>	1557
<i>Vorticella submicrostoma</i> Ghosh	1554
<i>Vorticella utriculus</i> Stokes	1555
<i>Vorticella vestita</i> Stokes	1556
<i>Zoothamnium arbuscula</i> Ehr.	1558
<i>Zoothamnium simplex</i> Kent	1559

Appendix A. Savannah River Taxonomic Accession List

V. Unidentified Taxa*

<u>Lowest Identified Classification</u>	<u>Unidentified Taxa Accession No.</u>
(3878) CLASS CRUSTACEAE:	3892 3893 1866 1867
(3699) CLASS GASTROTRICHA:	3700 3701 1665
(3736) CLASS HIRUDINOIDIA:	3738 3739 1680
(3703) CLASS NEMATOD:	3704 3705 1666
(3760) CLASS OLIGOCHAETA:	3803 3804 1755 1756 1757 1758 1759 1760
(3650) CLASS ROTIFERA:	3651 3652 1593
(3624) CLASS TURBELLARIA:	3641 3642 1591
(3654) ORDER BDELLOIDEA:	3655 1594 1595 1596
(3881) ORDER CLADOCERA:	3882 1862

* All taxa are in macroinvertebrate subgroup.

(3711)	ORDER GORDIOIDEA:	1668
(3929)	ORDER HYDROCARINA:	3957 1909
(3811)	ORDER MESOGASTROPODA:	3812 1762
(3741)	ORDER RHYNCHOBELLAE:	3742 1681
(3838)	FAMILY ANCYLIDAE:	1786
(3930)	FAMILY ARRENURIDAE:	1890
(3923)	FAMILY ASTACIDAE:	1884 1888
(4229)	FAMILY CHIRONOMIDAE:	2264
(3898)	FAMILY CYCLOPIDAE:	1868
(3770)	FAMILY ENCHYTRAEIFIDAE:	1717
(3744)	FAMILY GLOSSIPHONIIDAE:	1700 1703
(3792)	FAMILY GLOSSOSCOLECIDAE:	1748
(3821)	FAMILY HYDROBIIDAE:	1773
(3796)	FAMILY LUMBRICIDAE:	1753
(3762)	FAMILY LUMBRICULIDAE:	1716
(3801)	FAMILY MEGASCOLECIDAE:	1754
(3708)	FAMILY MERMITHIDAE:	1667
(3778)	FAMILY NAIDIDAE	1747
(3950)	FAMILY PIONIDAE	1906
(3618)	FAMILY SPONGILLIDAE	1575 1576 1577

Appendix B. Sample Data Presentations

- B.1 Savannah River Taxonomic Data Summary.**
- B.2 Sample Presence/Absence Data for Two Diatom Species.**
- B.3 Sample Diatometer Data.**
- B.4 Savannah River Water Chemistry Summary.**

B.1. Savannah River Taxonomic Data

Taxonomic data from all 16 major river surveys have been summarized in Figures B.1.1-B.1.36. These data have been condensed into major taxonomic subgroups and graphed to show general trends over time at each survey station.

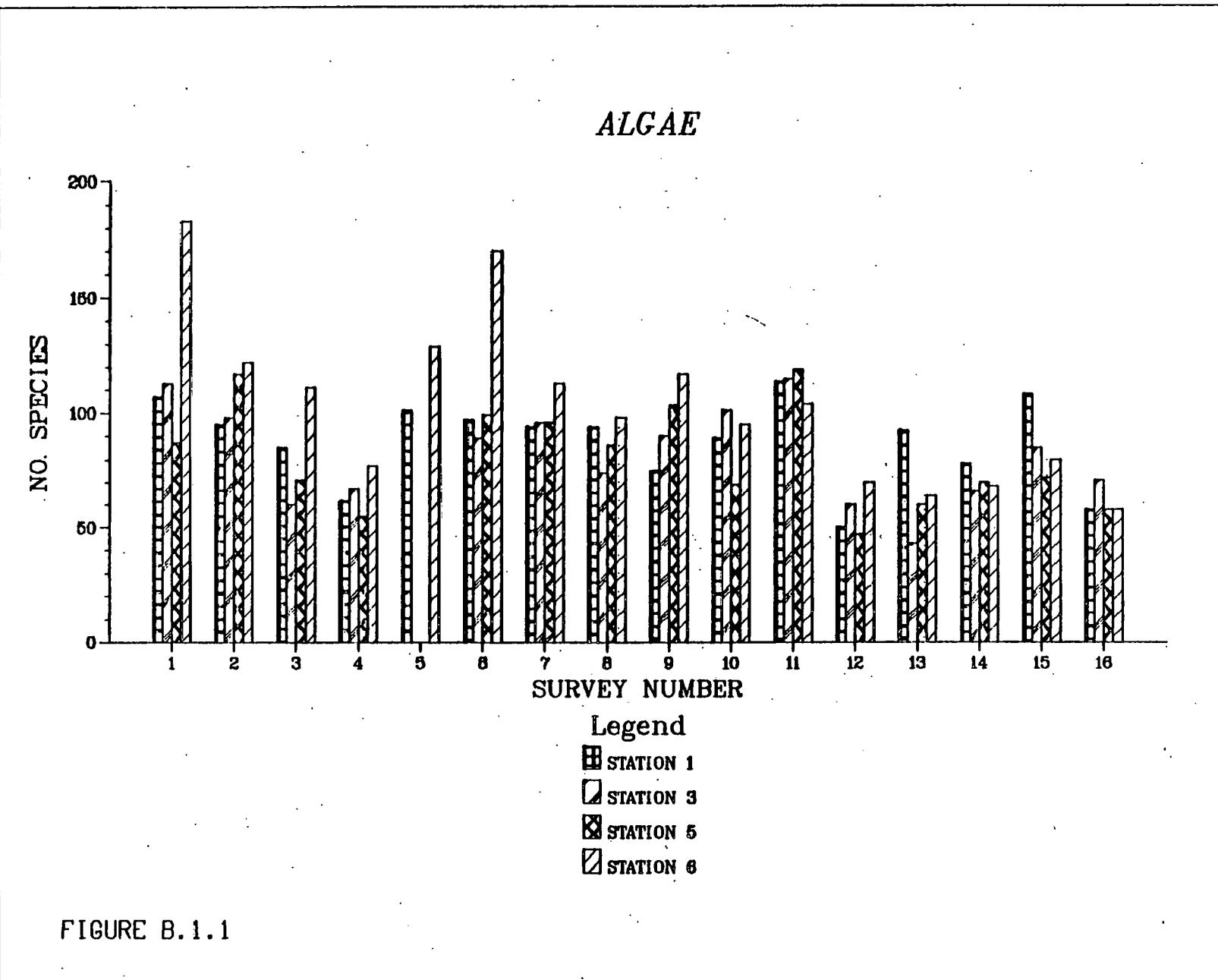


FIGURE B.1.1

DIVISION BACILLARIOPHYTA
(Diatoms)

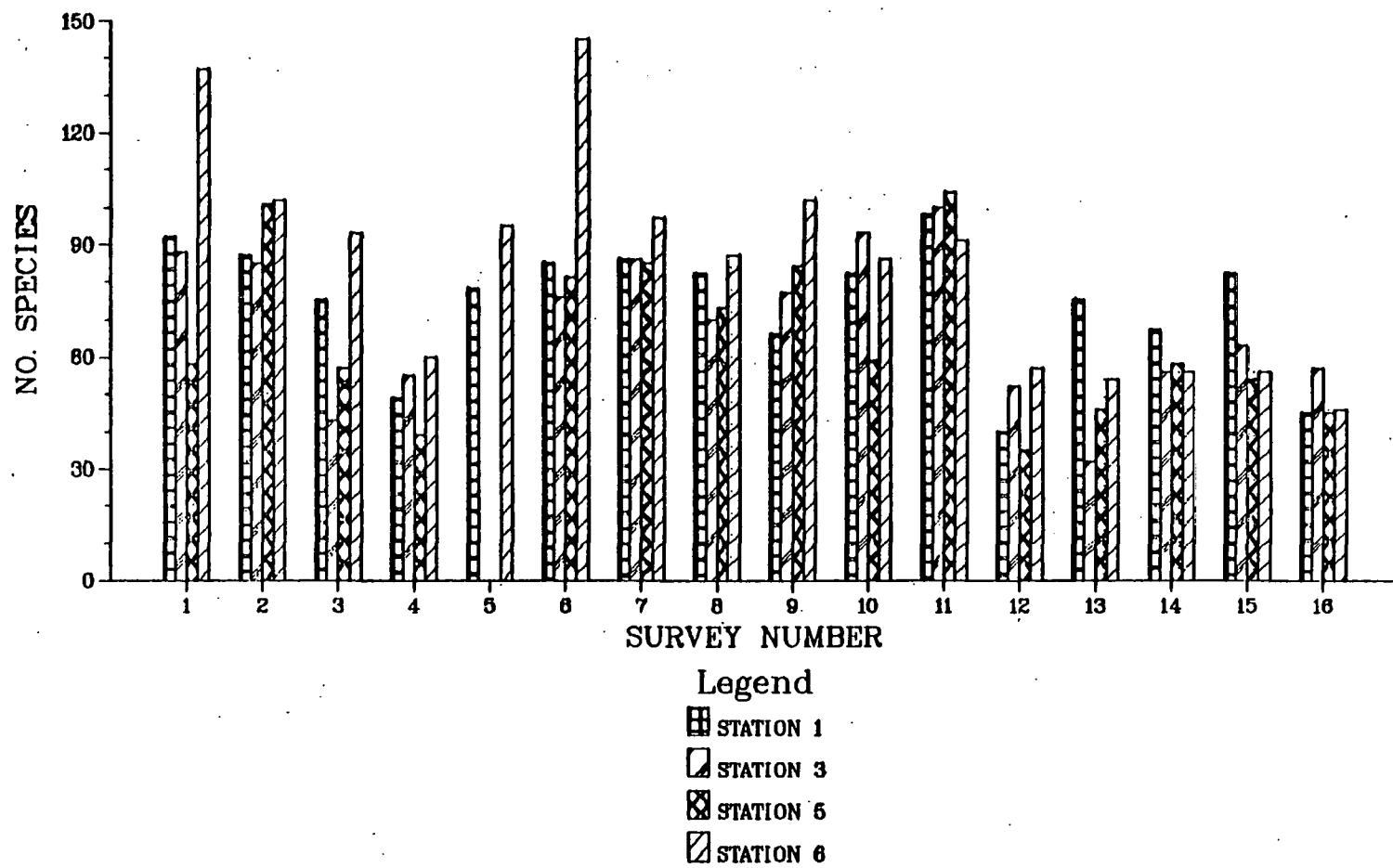


FIGURE B.1.2

*DIVISION CHLOROPHYTA
(Green Algae.)*

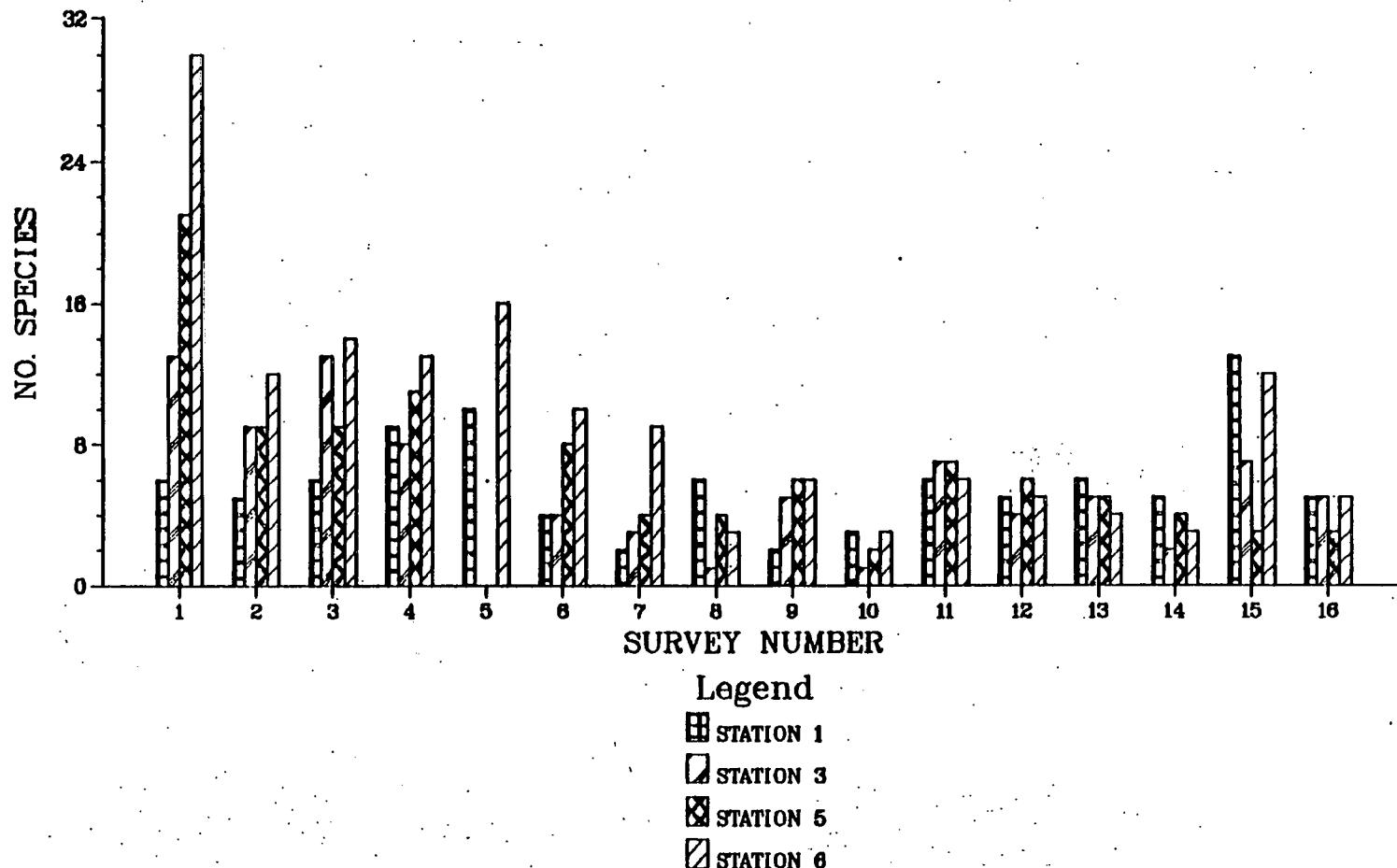


FIGURE B.1.3

*DIVISION CHROMOPHYTA
(Yellow-Green Algae)*

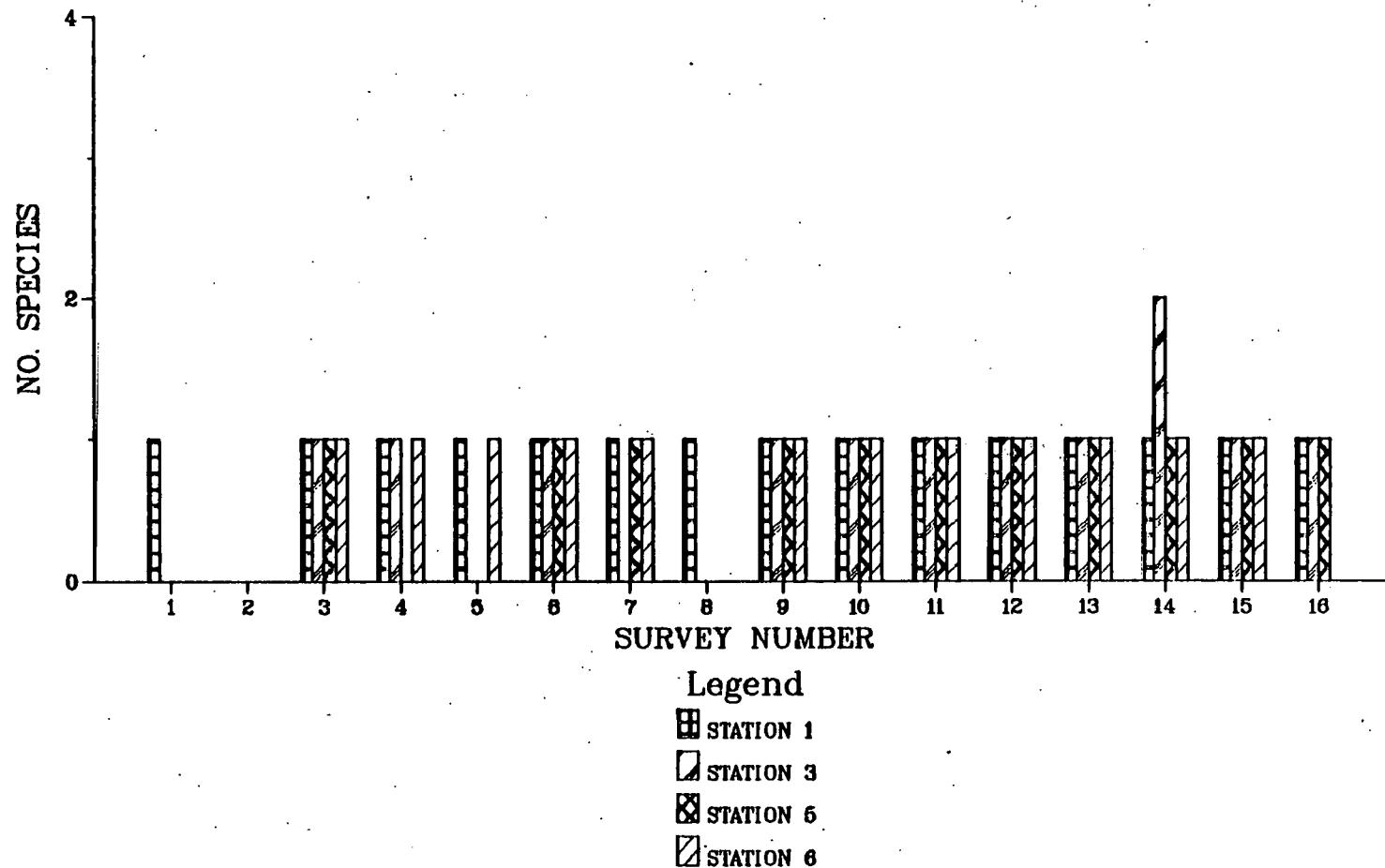


FIGURE B.1.4

*DIVISION CYANOPHYTA
(Blue-Green Algae)*

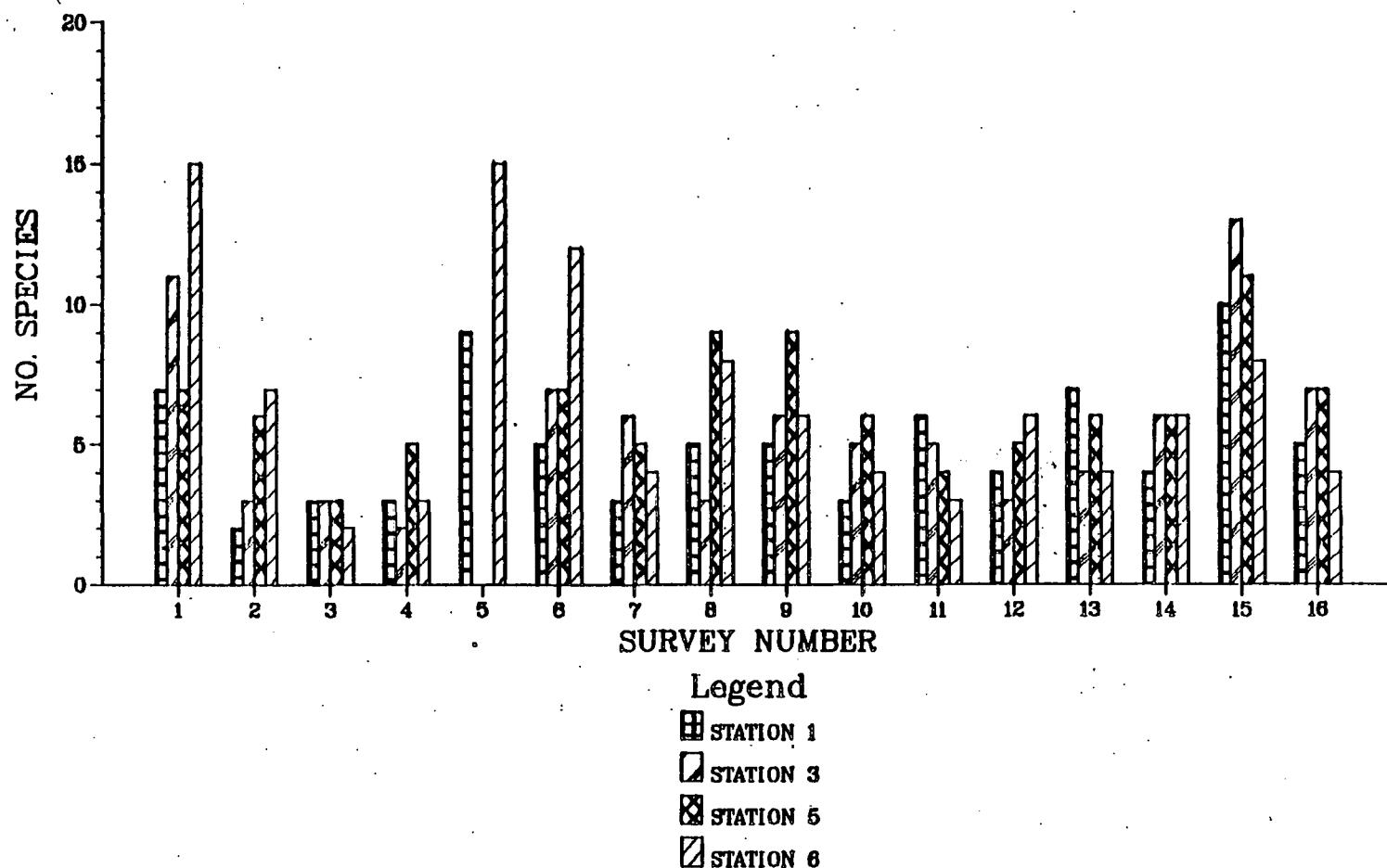


FIGURE B.1.5

*DIVISION RHODOPHYTA
(Red Algae)*

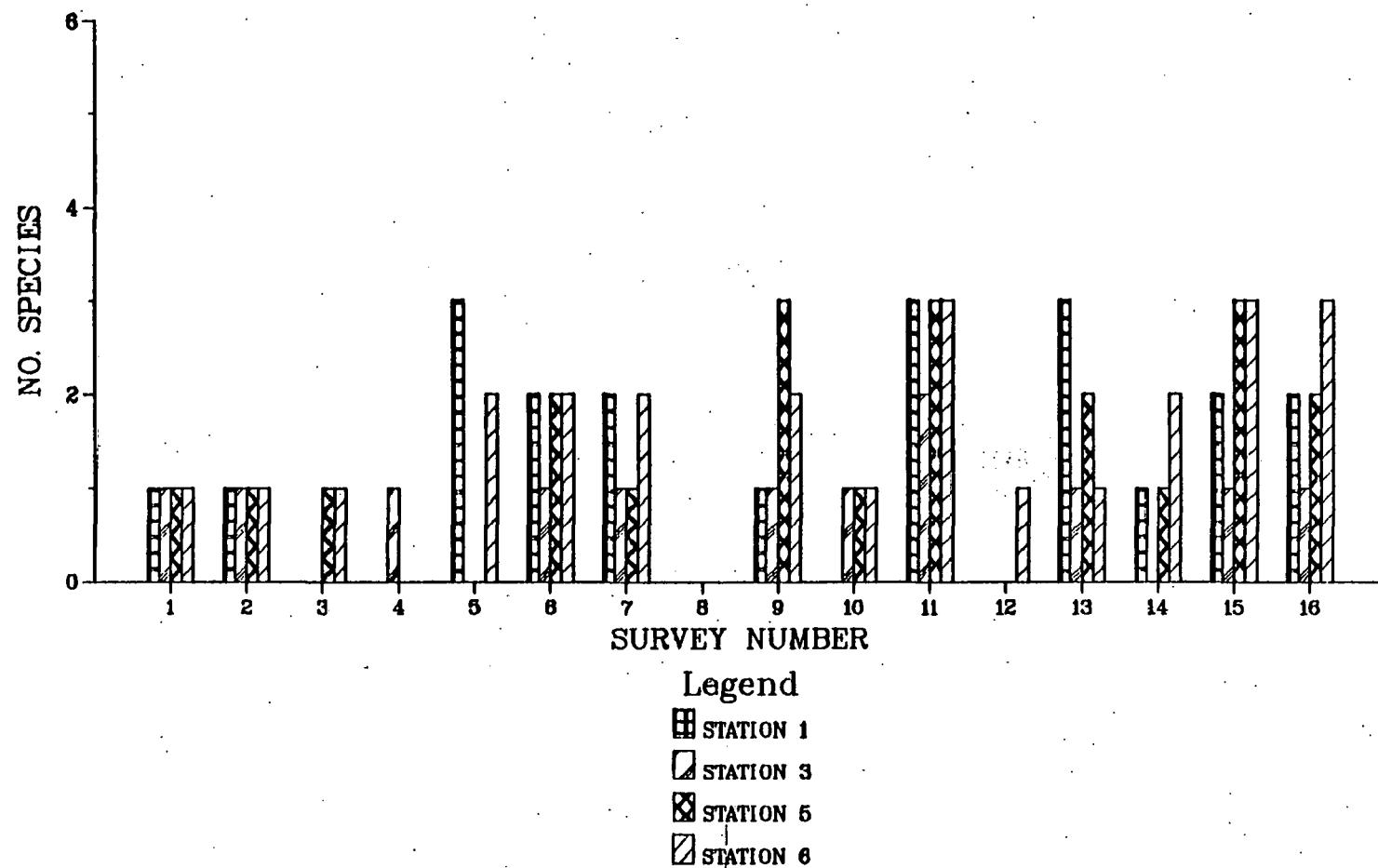


FIGURE B.1.6

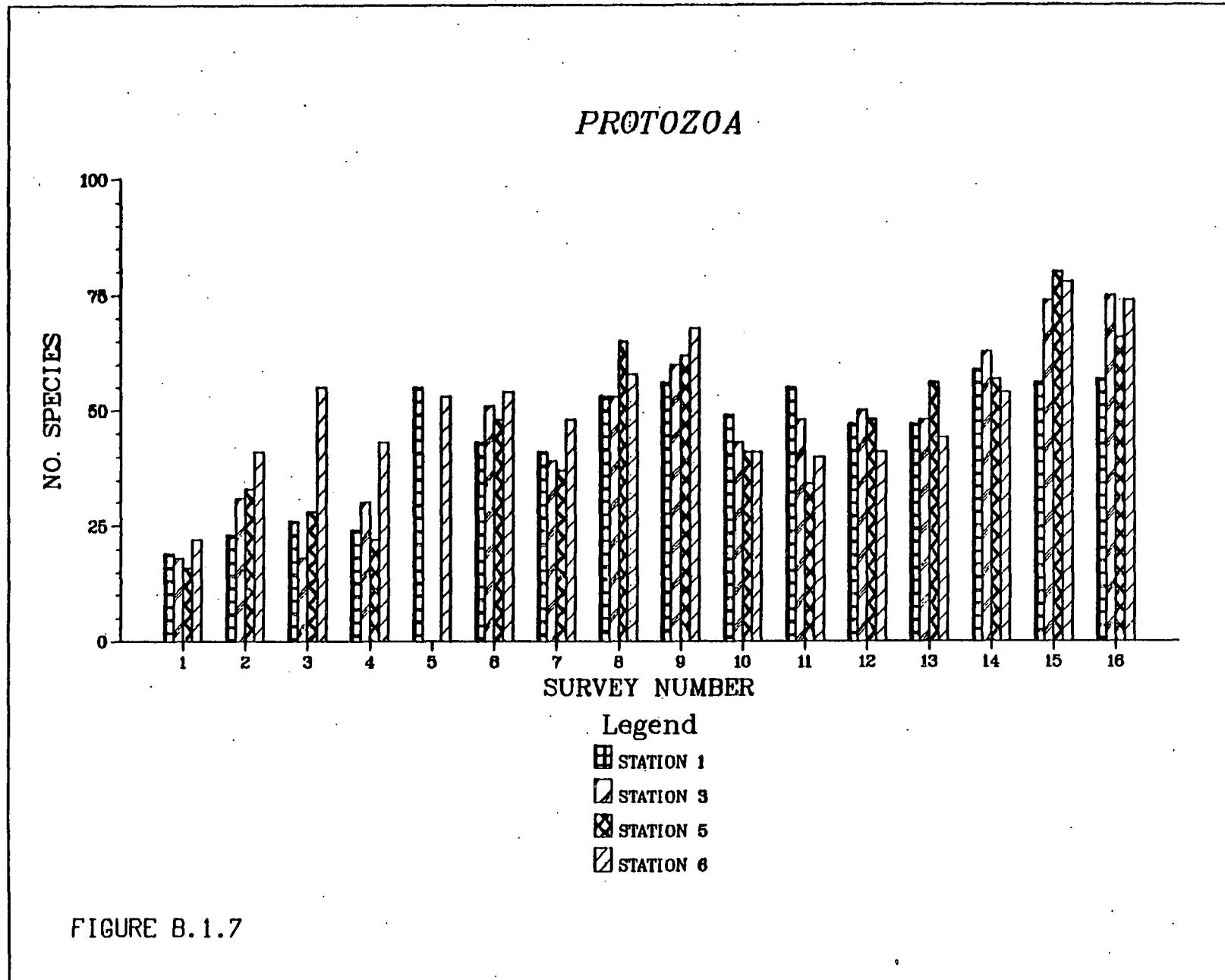


FIGURE B.1.7

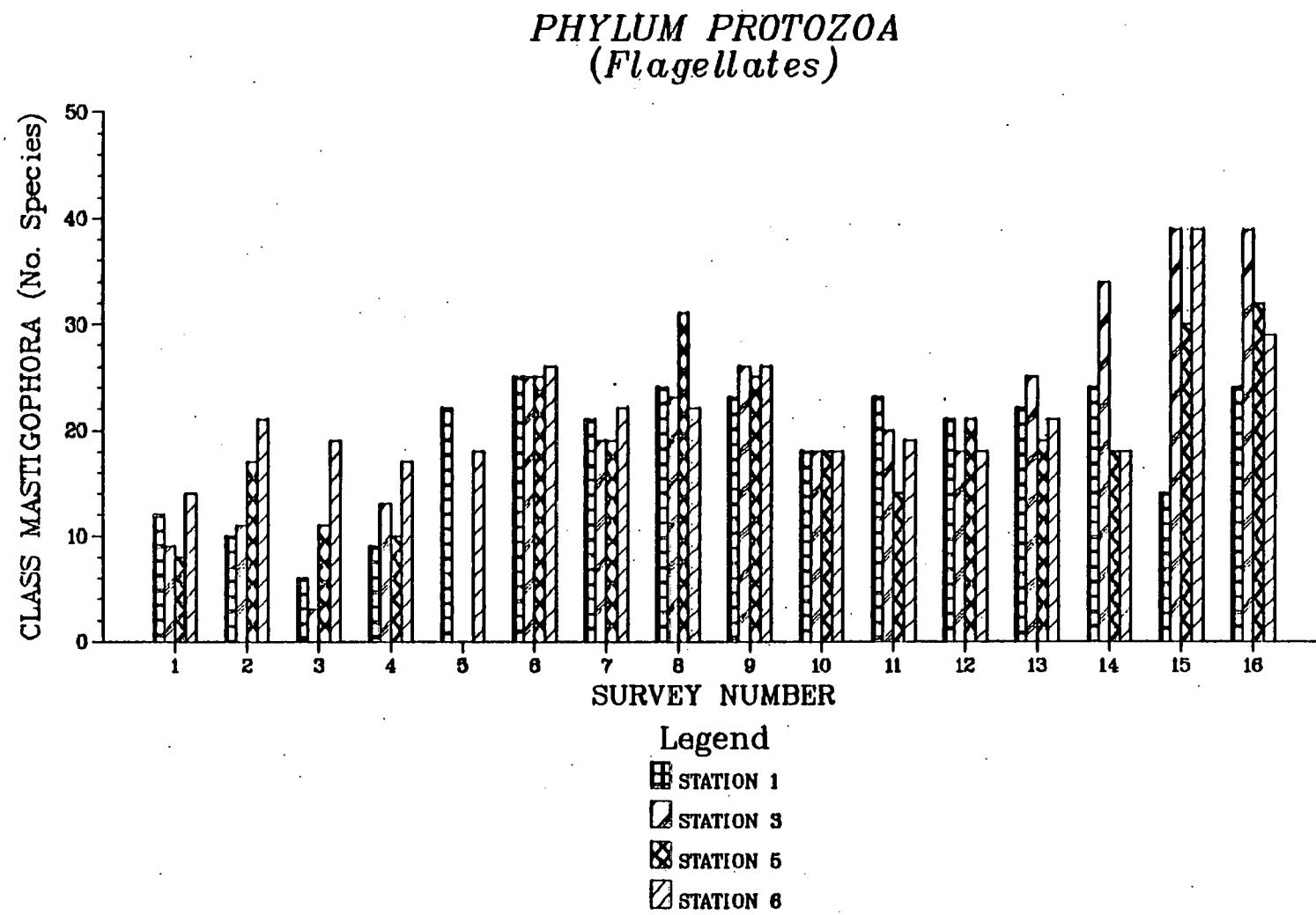


FIGURE B.1.8

PHYLUM PROTOZOA
(*Amoeba*)

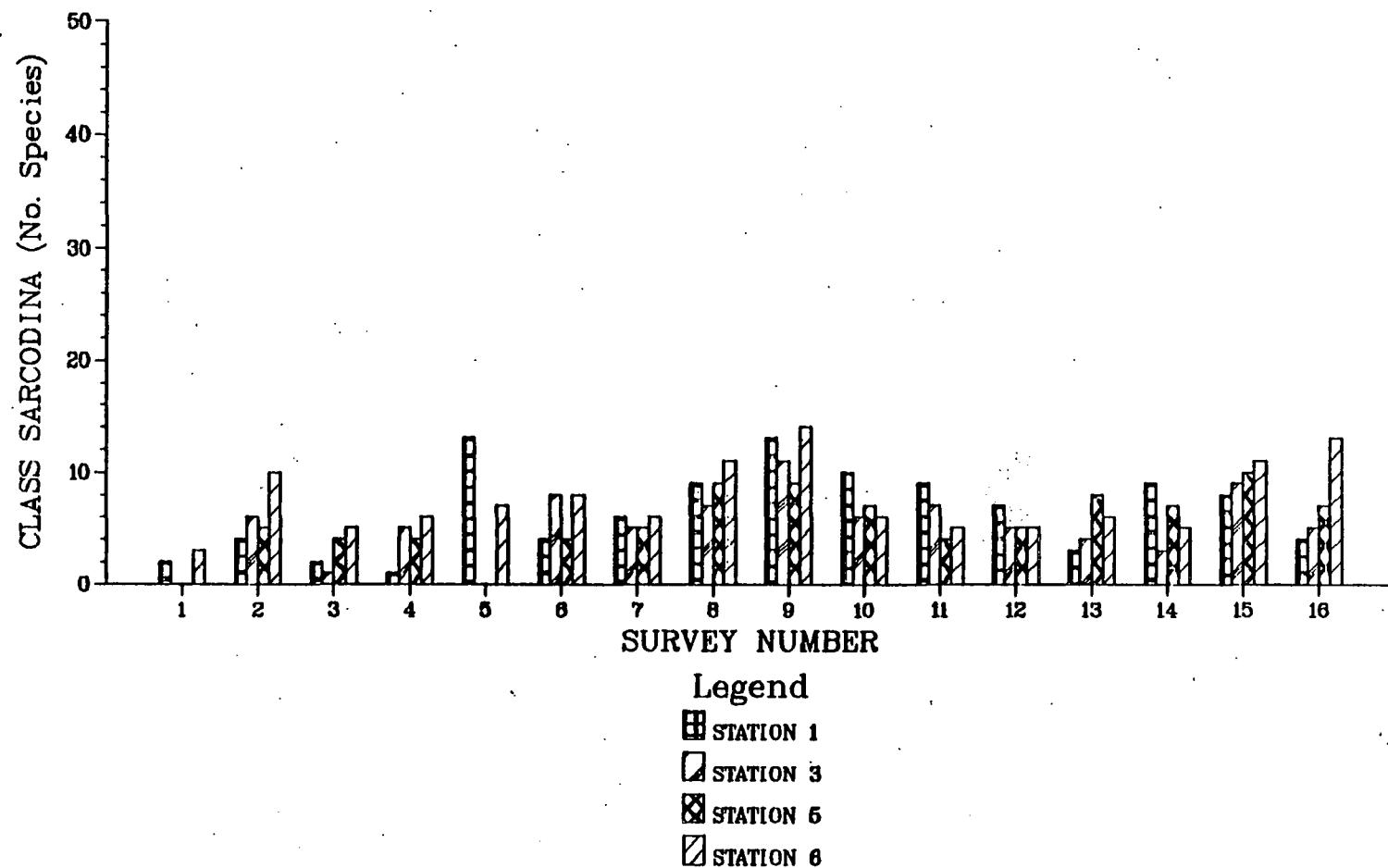


FIGURE B.1.9

*PHYLUM PROTOZOA
(Ciliates)*

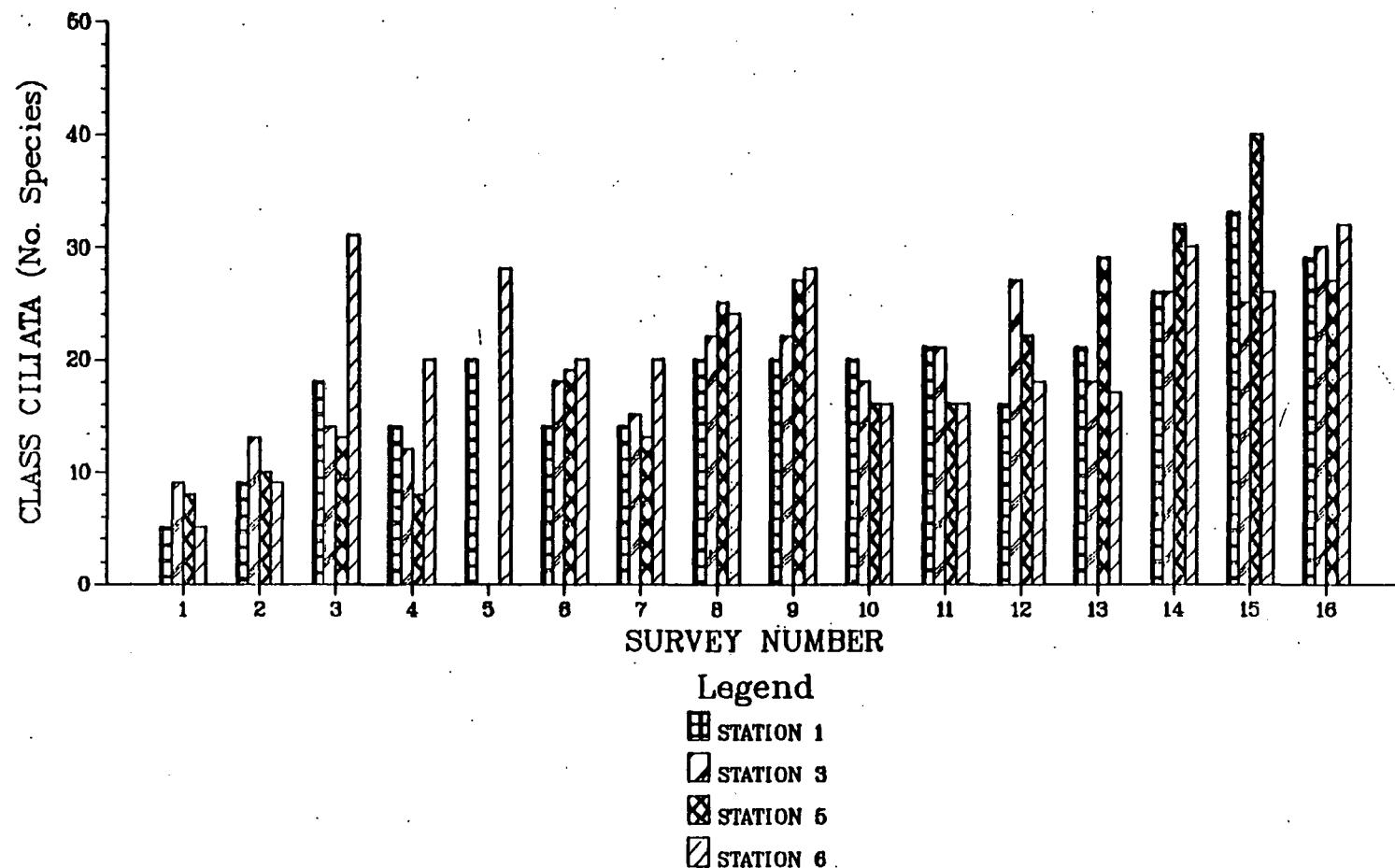


FIGURE B.1.10

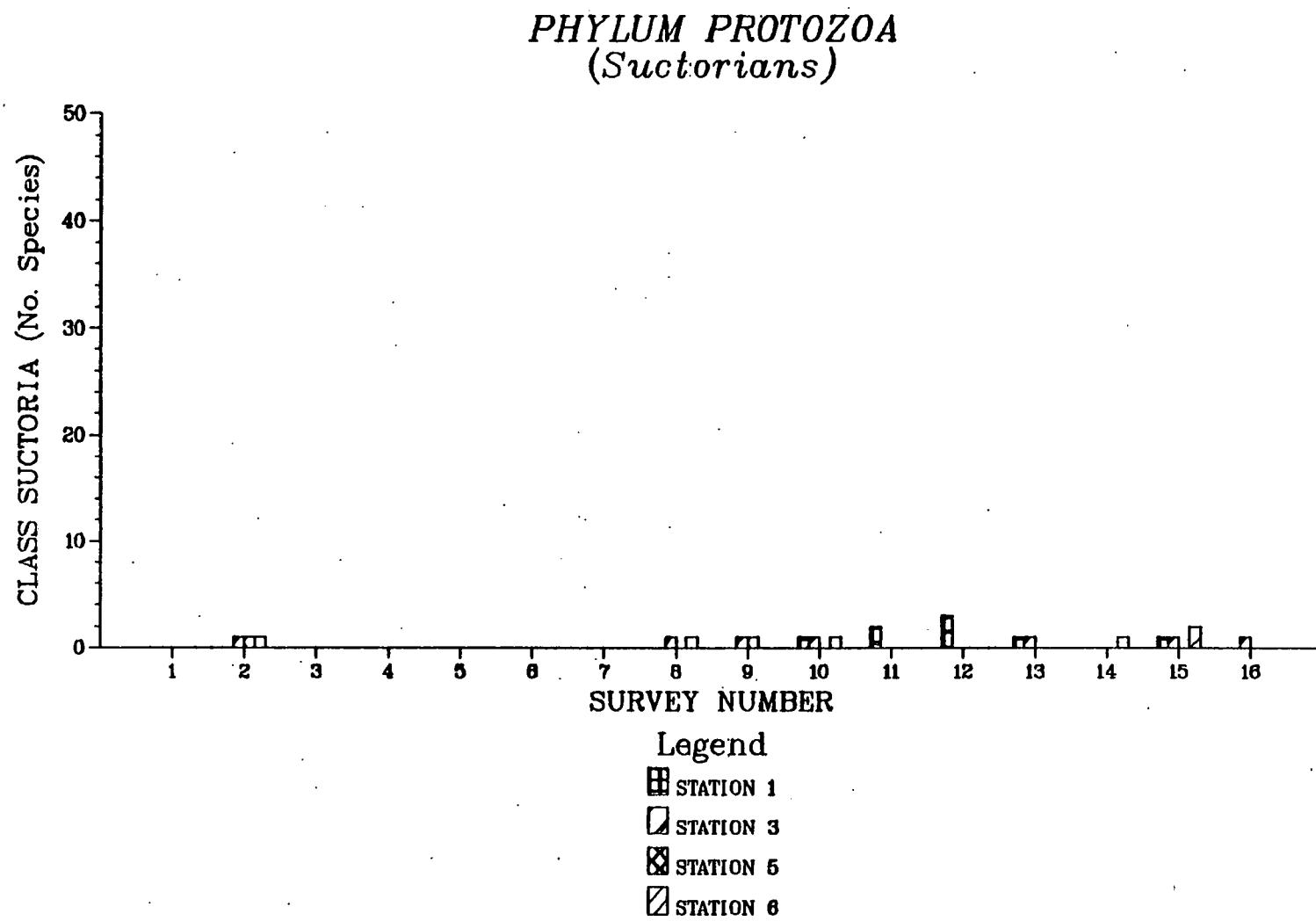
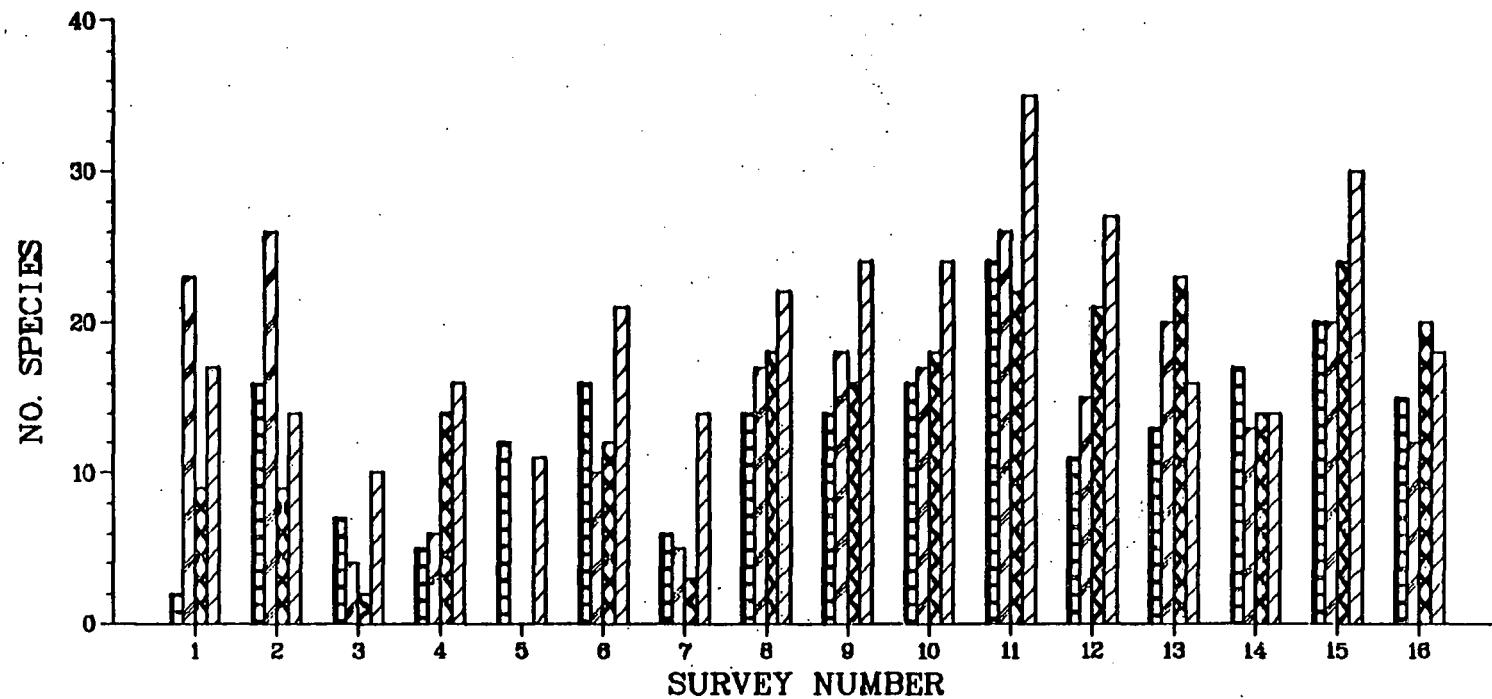


FIGURE B.1.11

PHYLUM MOLLUSCA
(Snails, Clams)



Legend

- STATION 1
- STATION 3
- ☒ STATION 5
- ▨ STATION 6

FIGURE B.1.12

PHYLUM ARTHROPODA
(Except Insects)

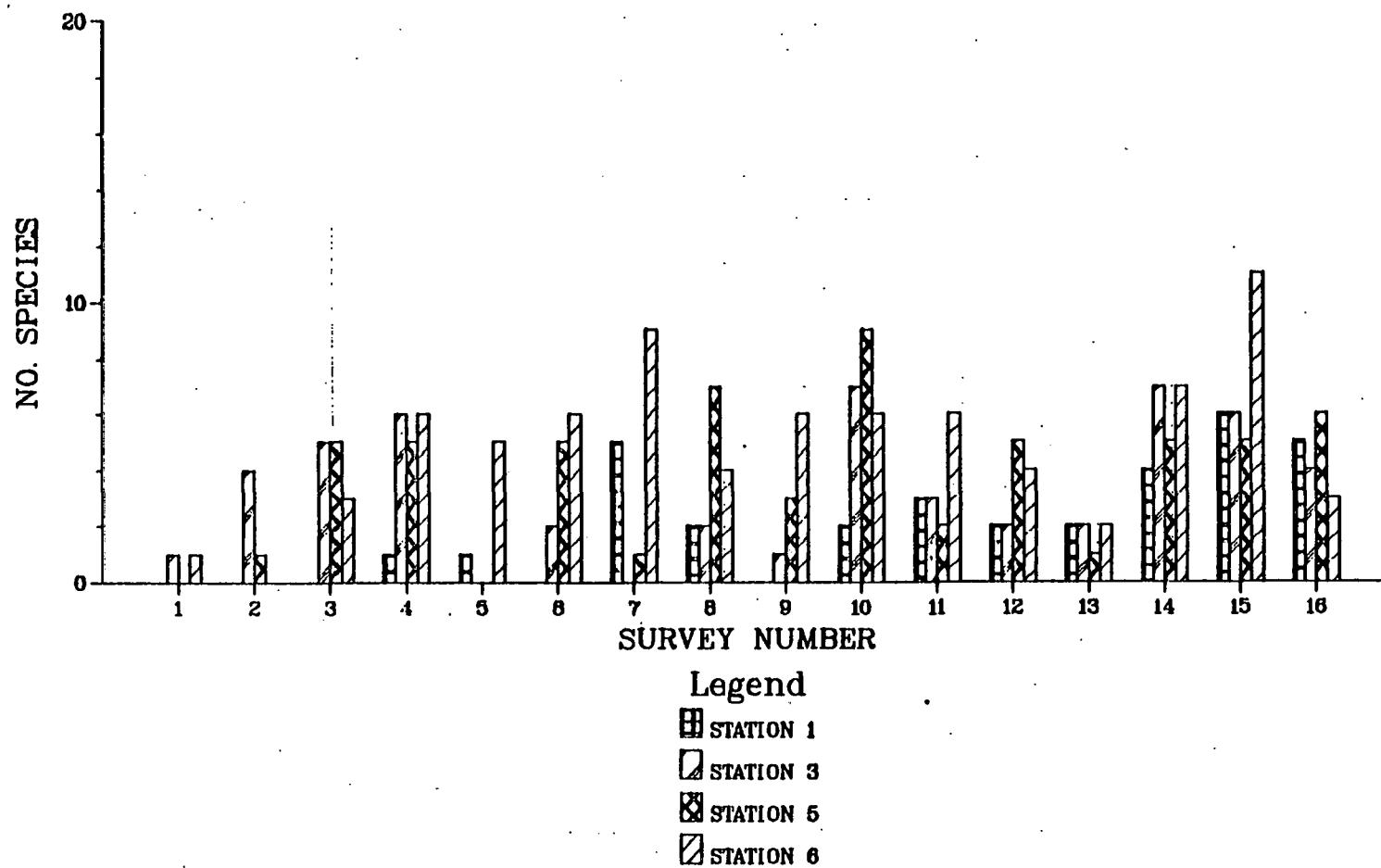


FIGURE B.1.13

PHYLUM ARTHROPODA, CLASS INSECTA

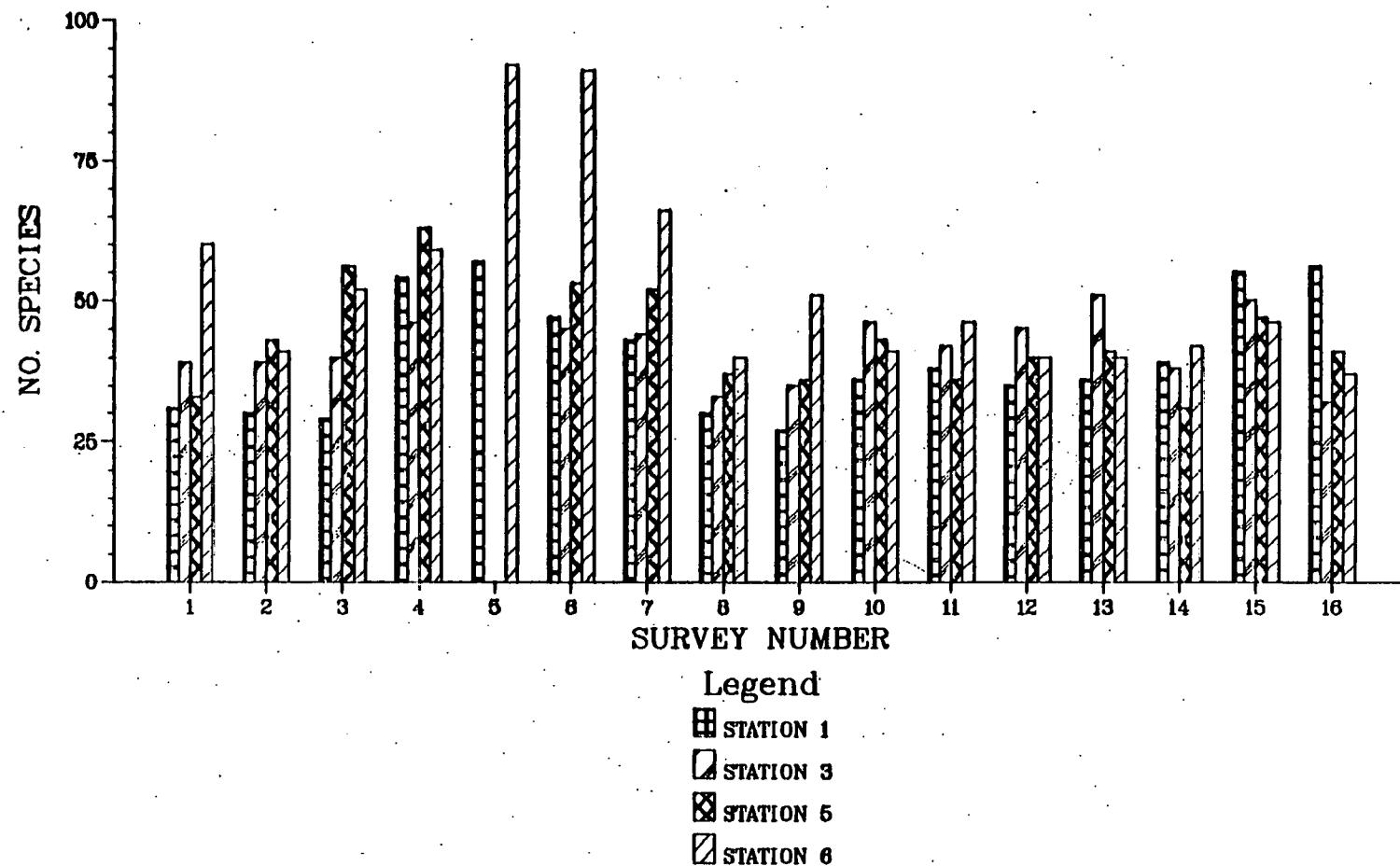


FIGURE B.1.14

*PHYLUM ARTHROPODA, CLASS INSECTA
(Dragon Flies)*

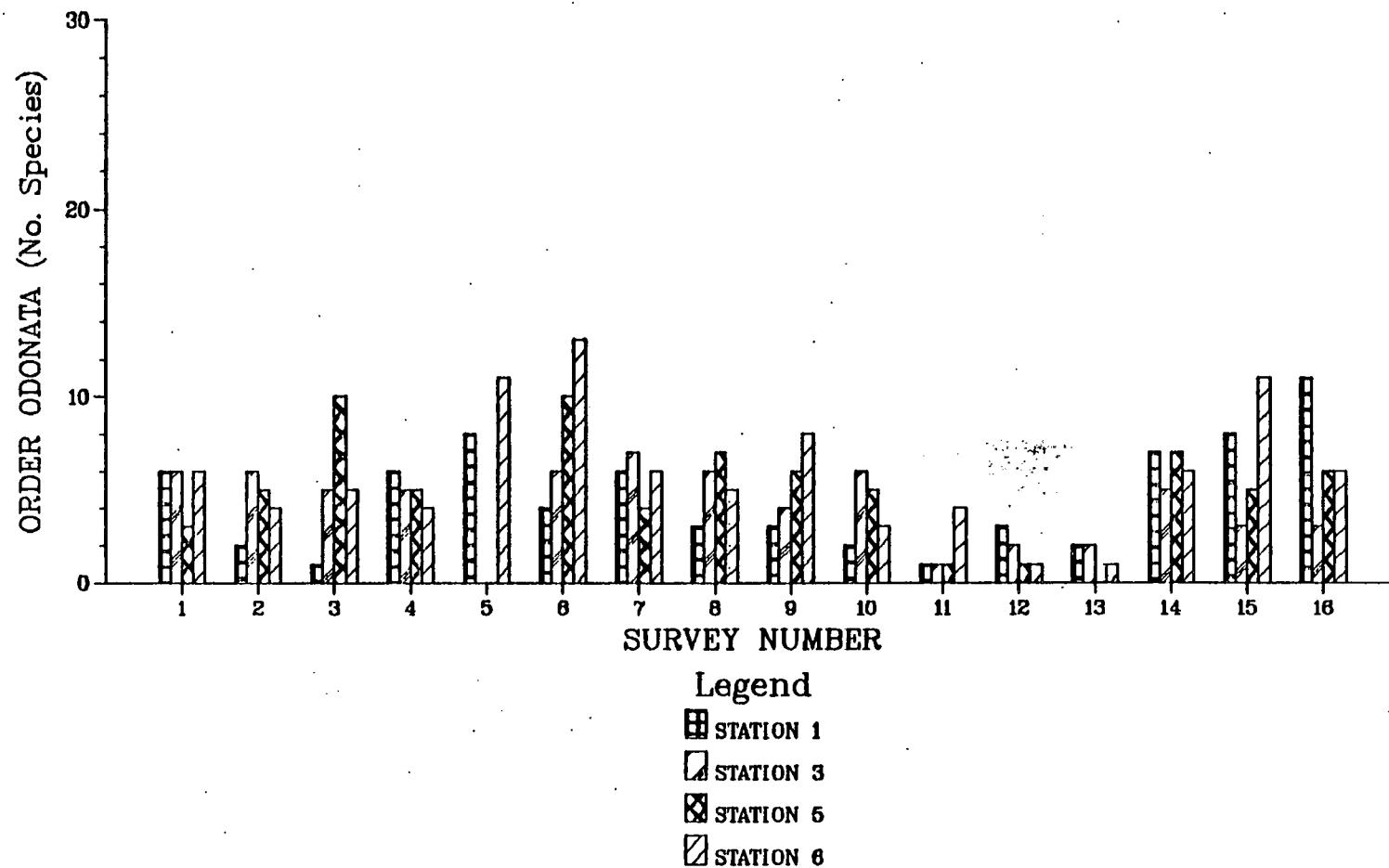


FIGURE B.1.15

PHYLUM ARTHROPODA, CLASS INSECTA
(Mayflies)

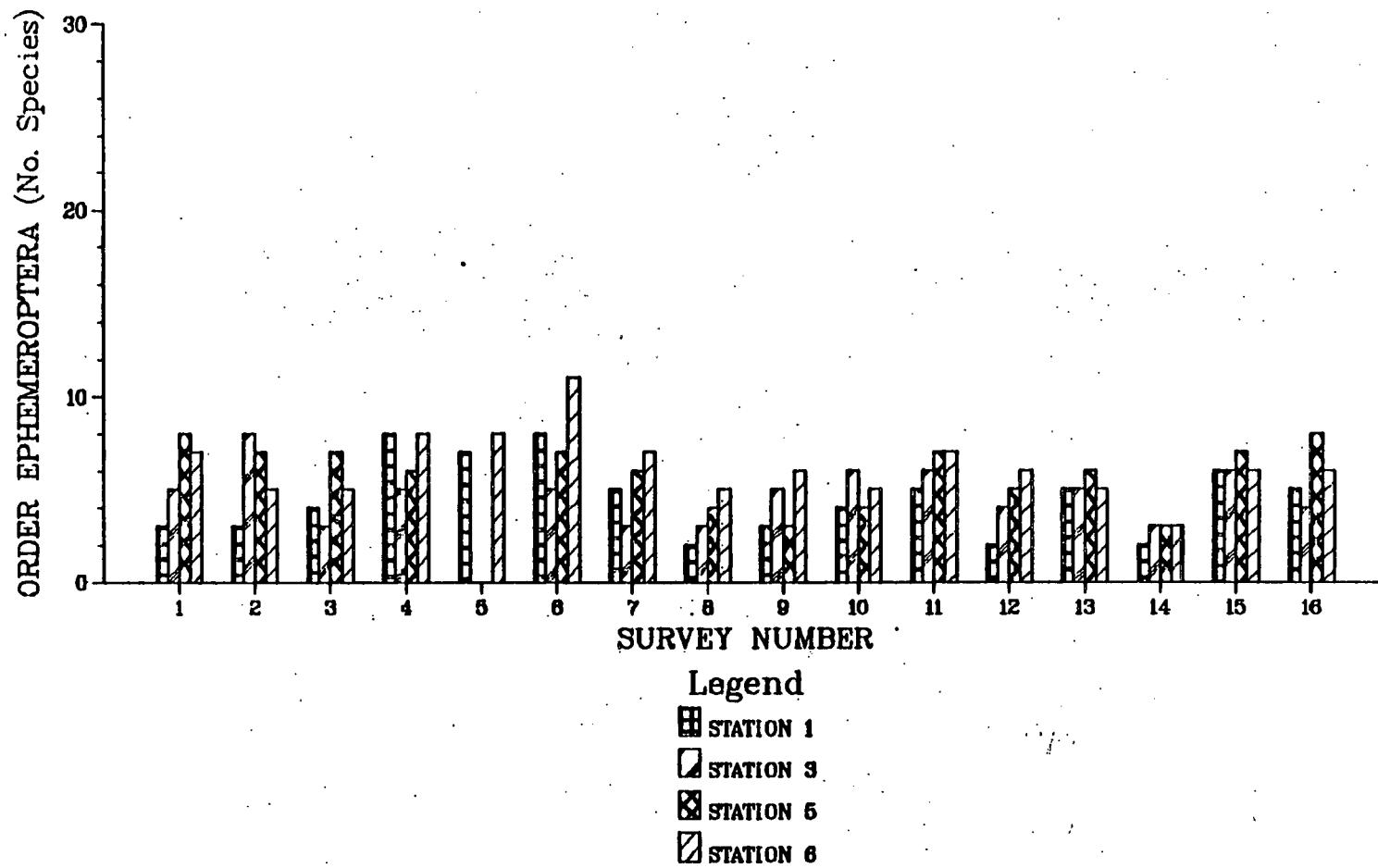


FIGURE B.1.16

PHYLUM ARTHROPODA, CLASS INSECTA
(Stoneflies)

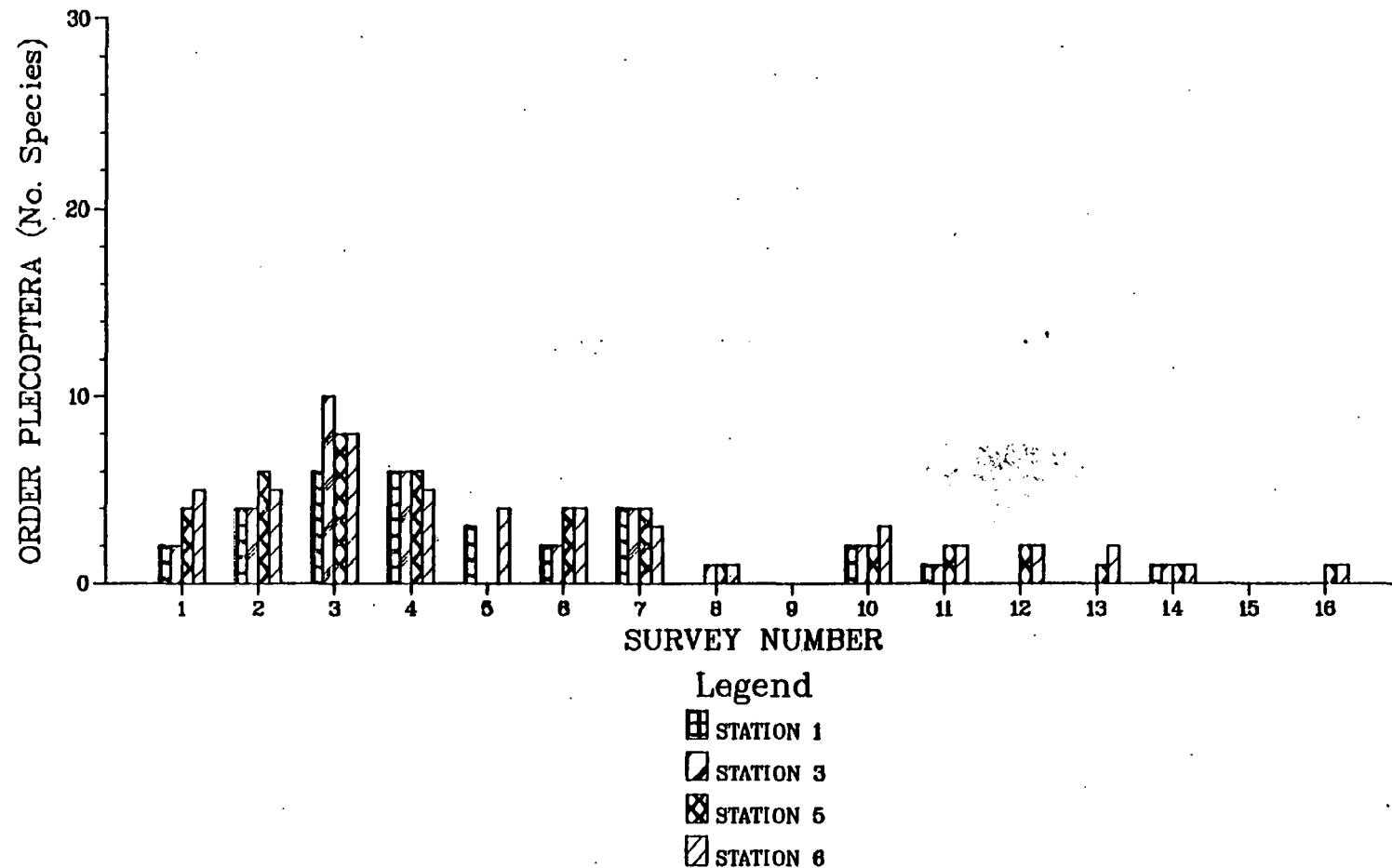


FIGURE B.1.17

PHYLUM ARTHROPODA, CLASS INSECTA
(True Bugs)

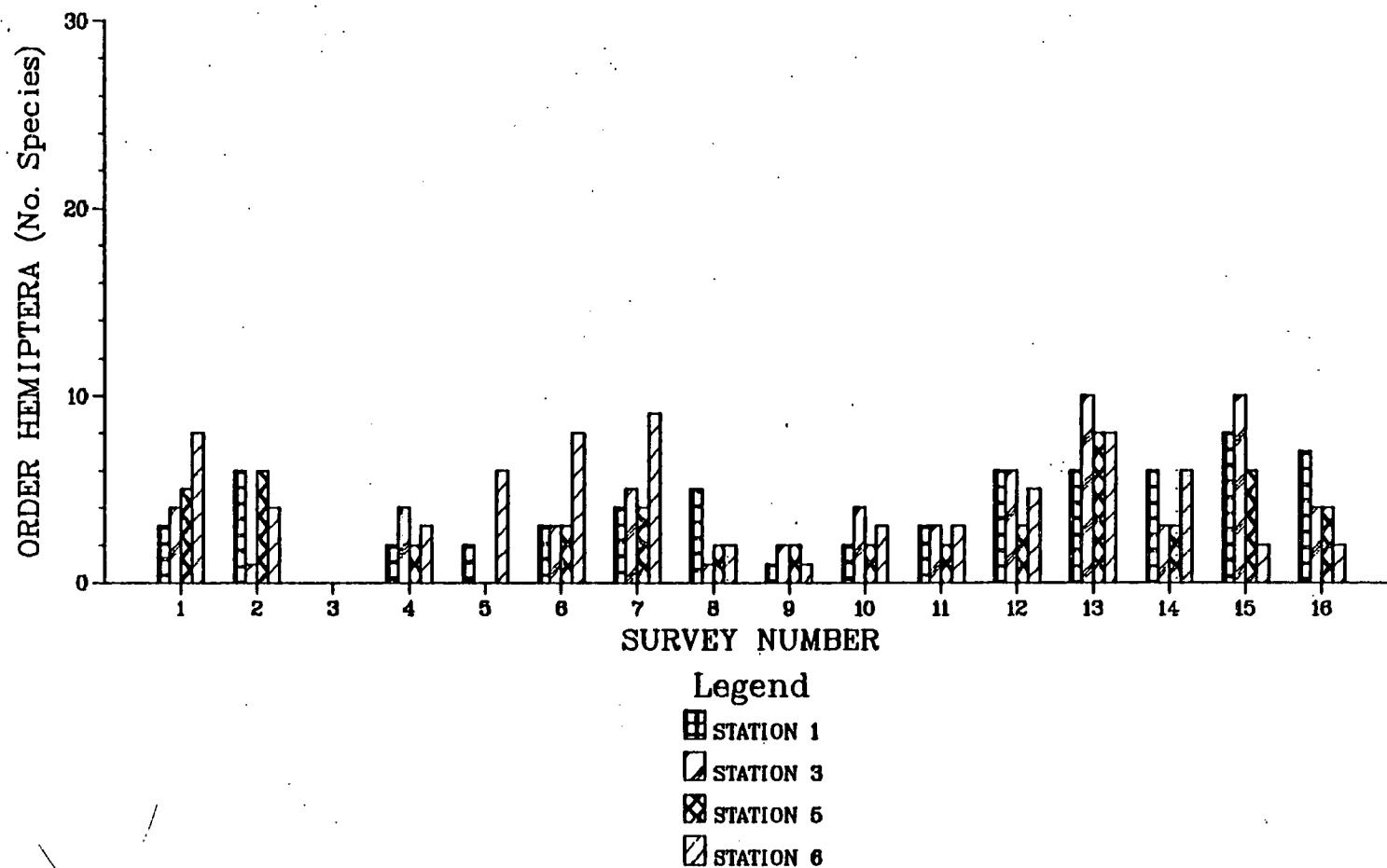


FIGURE B.1.18

PHYLUM ARTHROPODA, CLASS INSECTA
(Spongilla Flies)

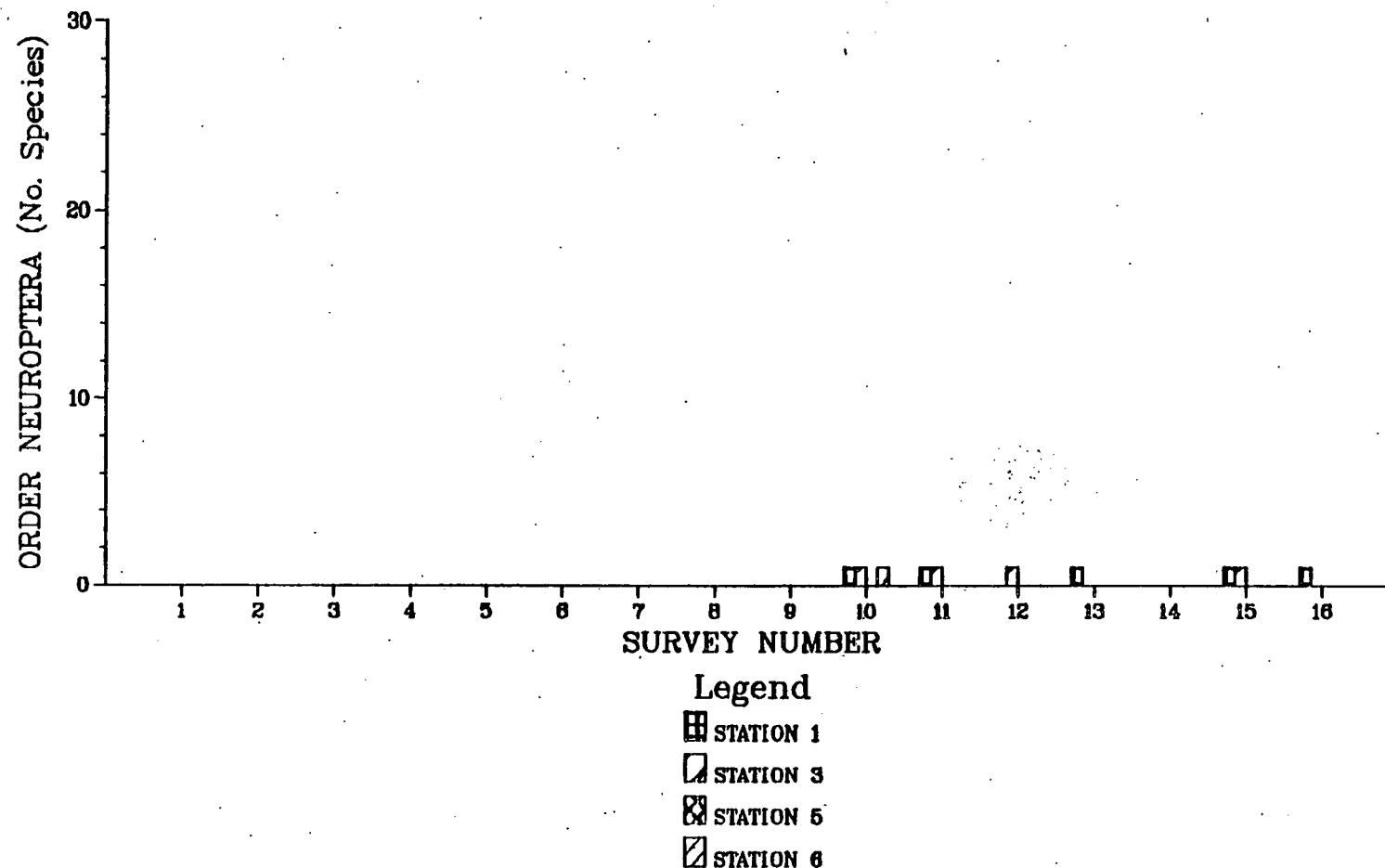


FIGURE B.1.19

PHYLUM ARTHROPODA, CLASS INSECTA
(Alderflies, Fishflies, Dobson Flies)

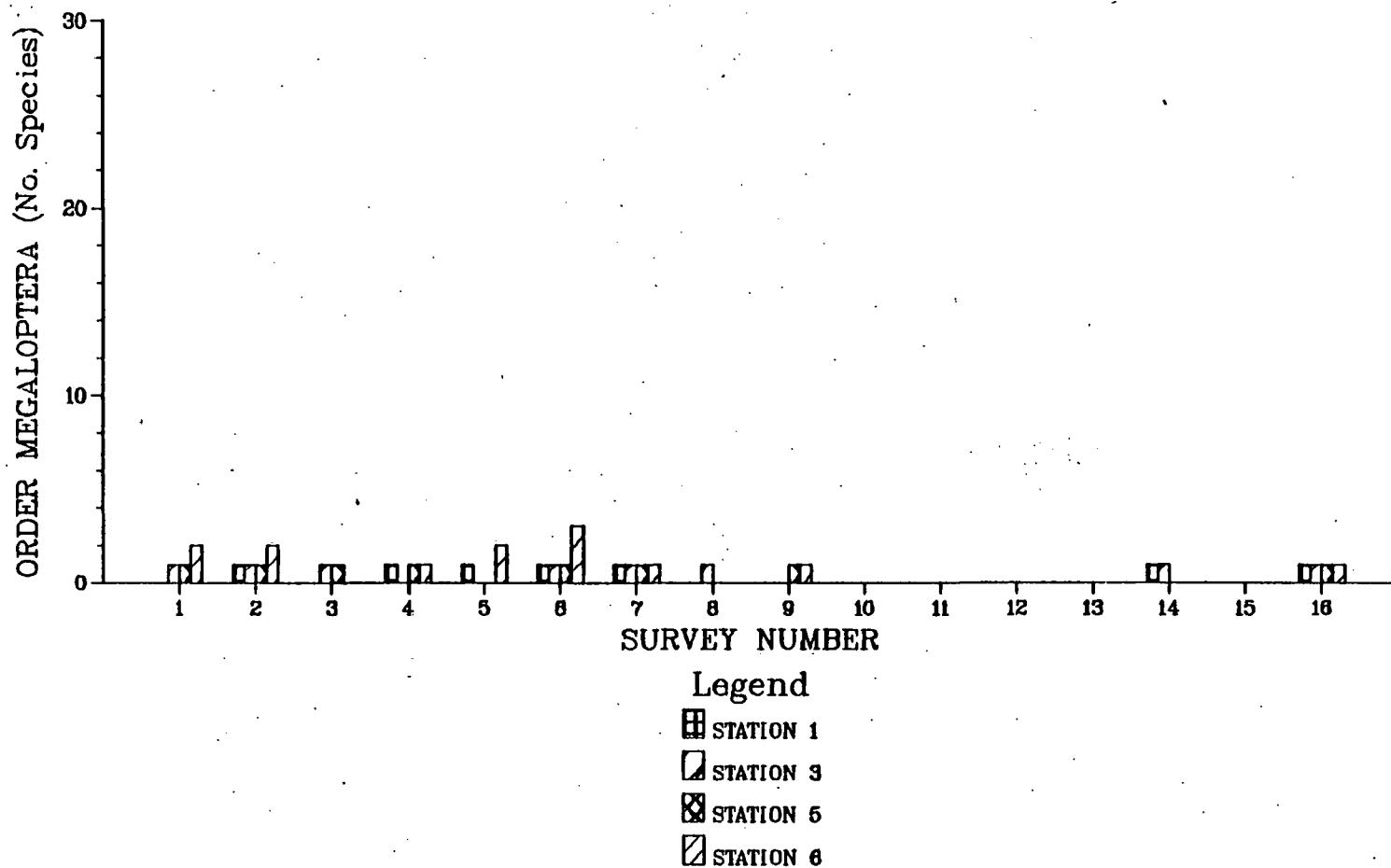


FIGURE B.1.20

**PHYLUM ARTHROPODA, CLASS INSECTA
(Beetles)**

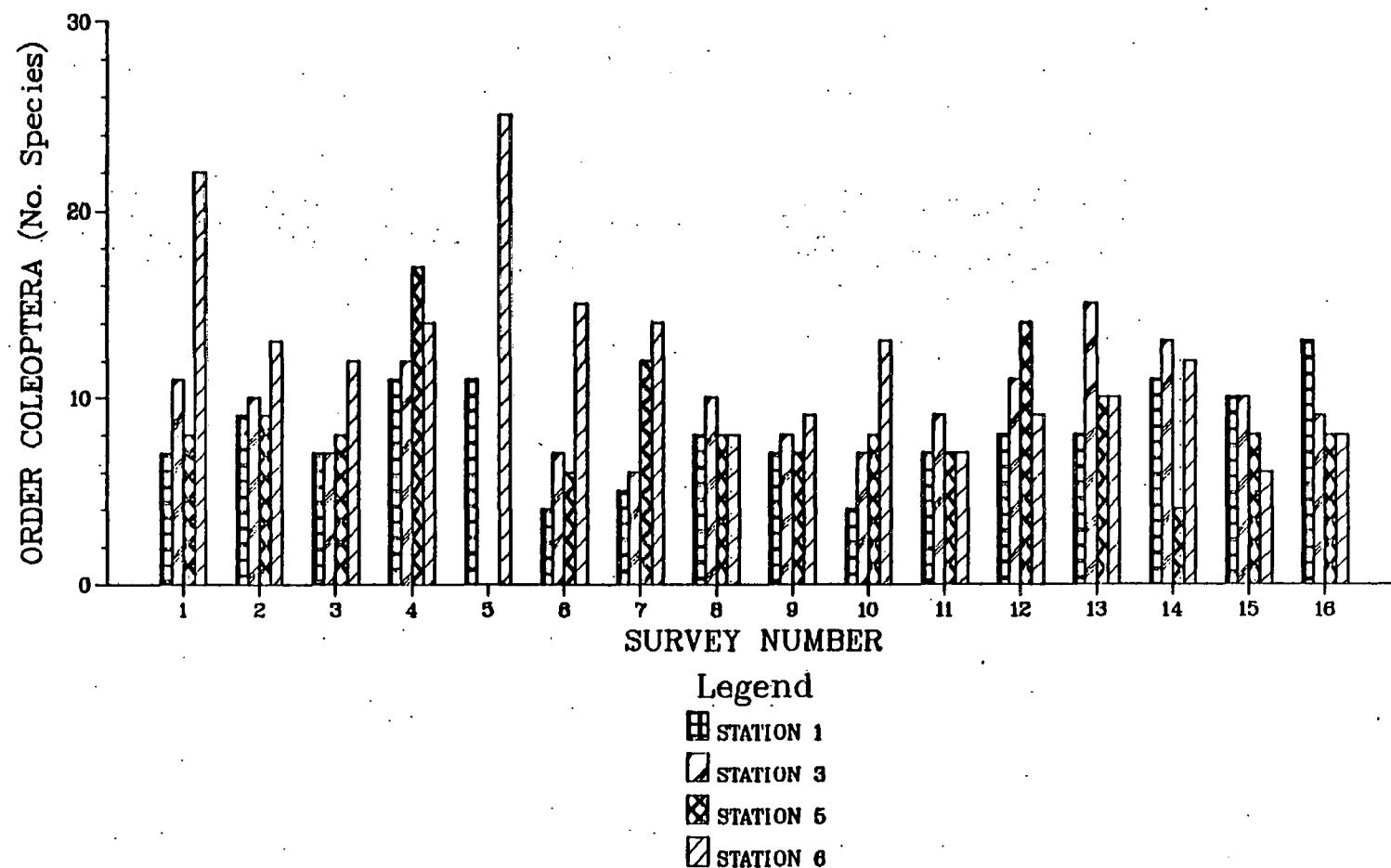


FIGURE B.1.21

PHYLUM ARTHROPODA, CLASS INSECTA
(Caddisflies)

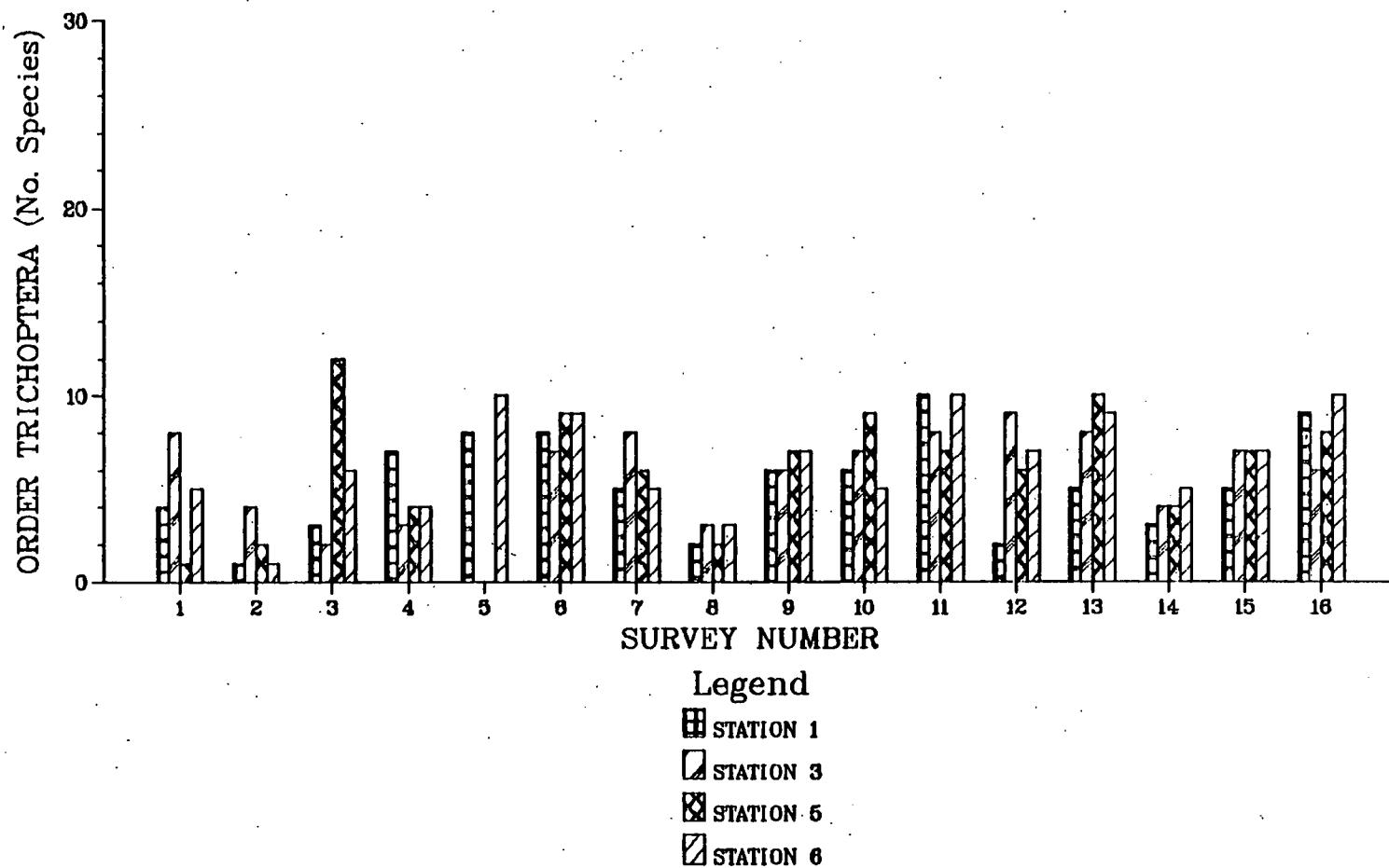


FIGURE B.1.22

PHYLUM ARTHROPODA, CLASS INSECTA
(Butterflies, Moths)

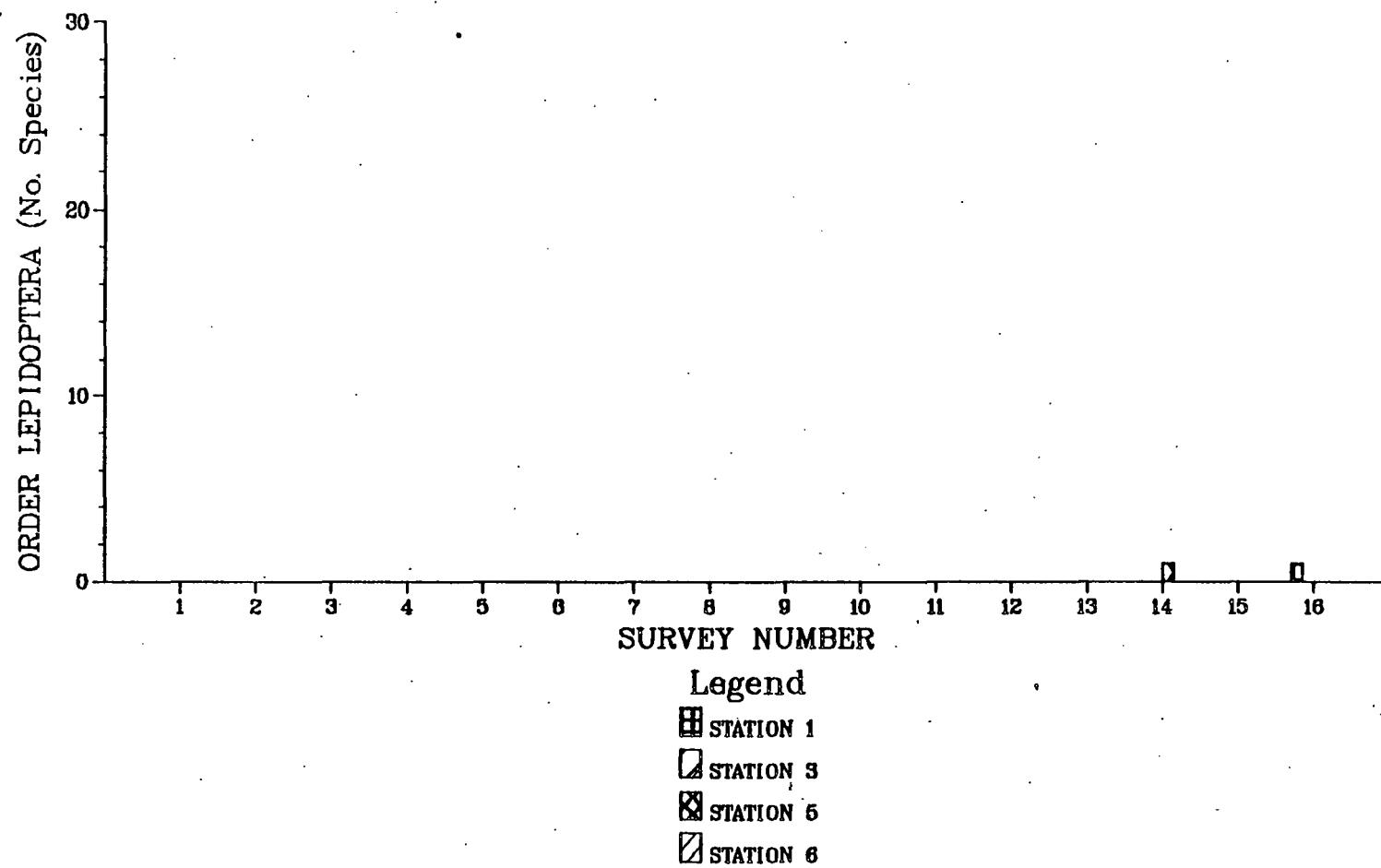


FIGURE B.1.23

OTHER INVERTEBRATES

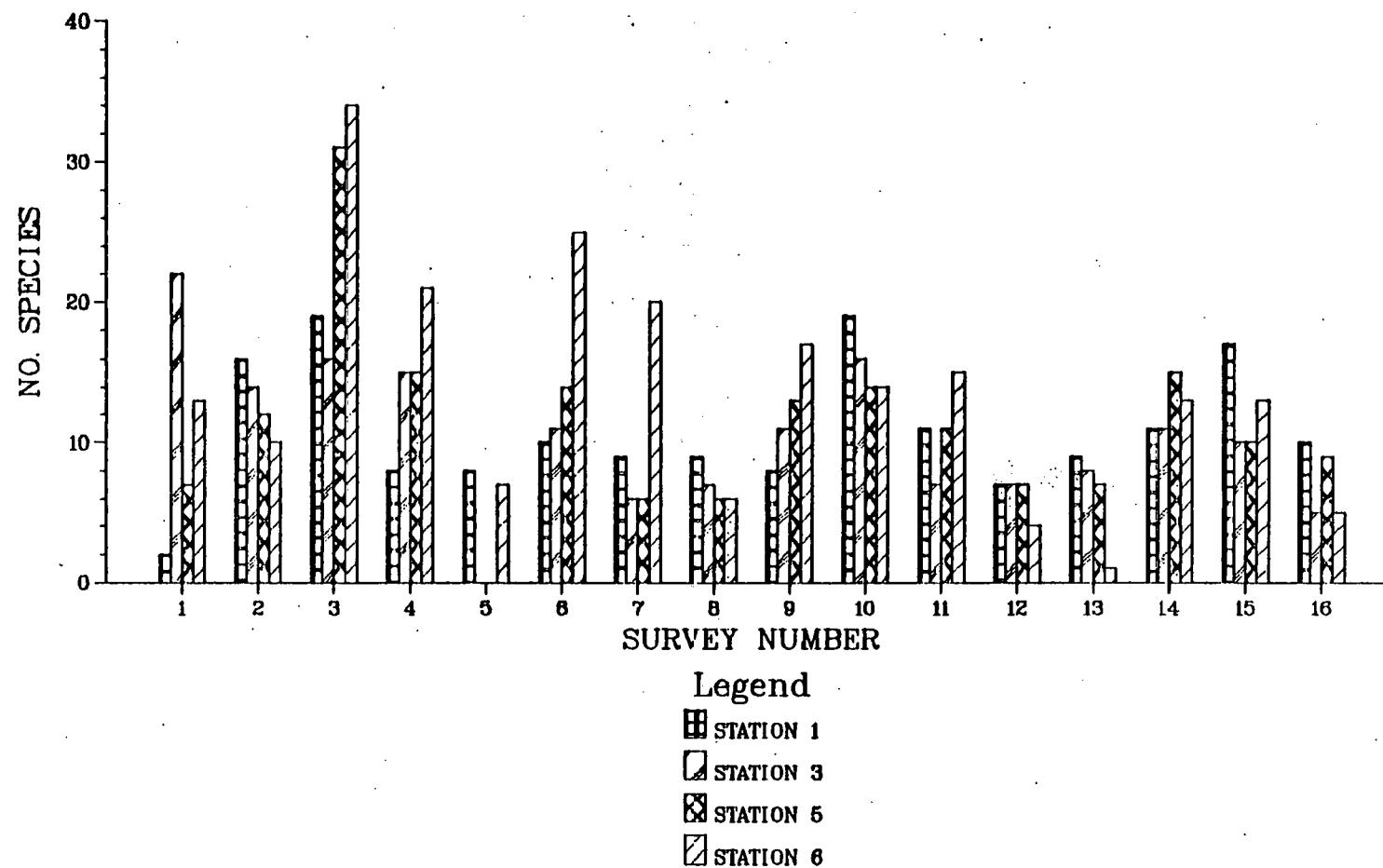


FIGURE B.1.24

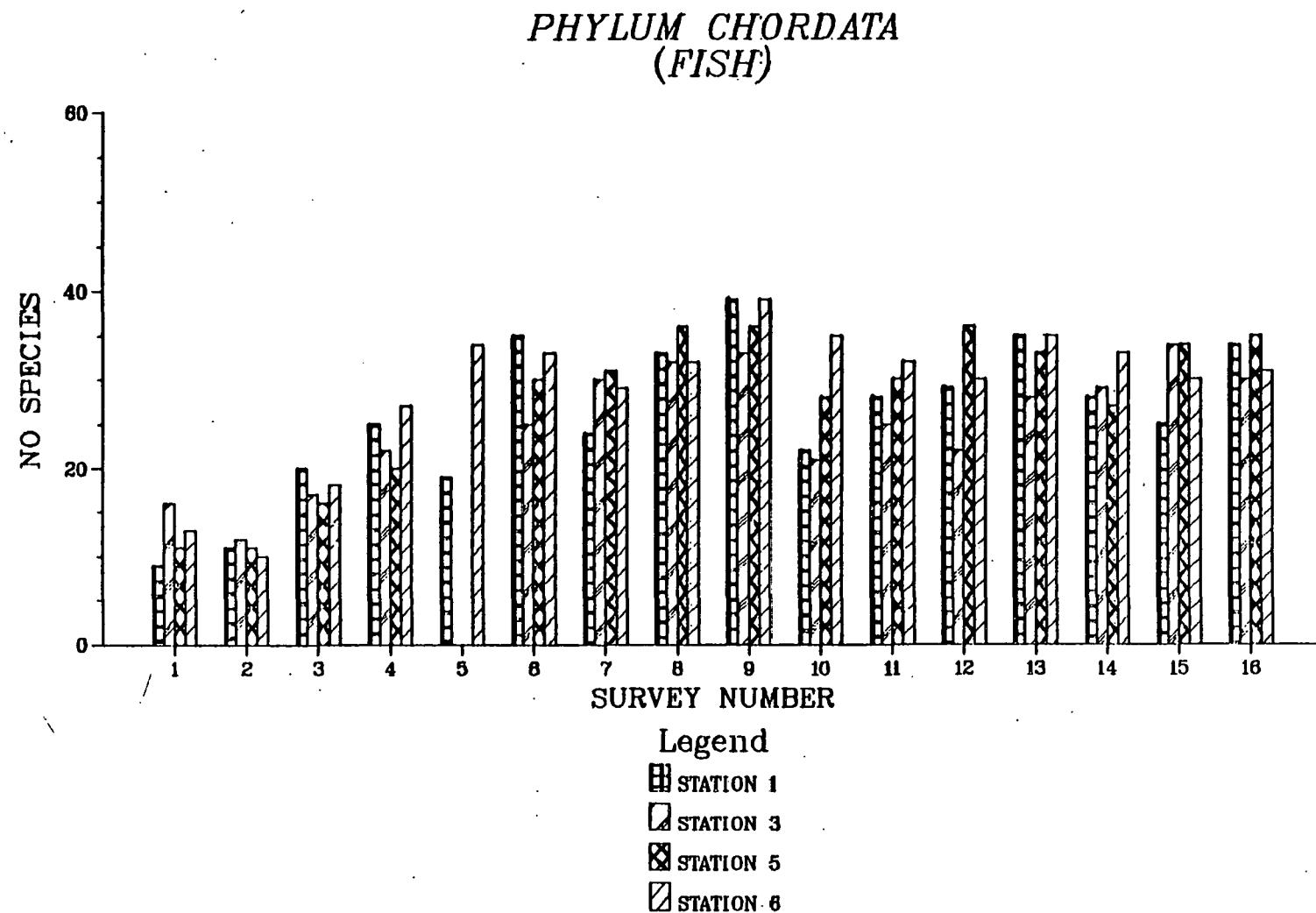


FIGURE B.1.25

PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Bowfin)

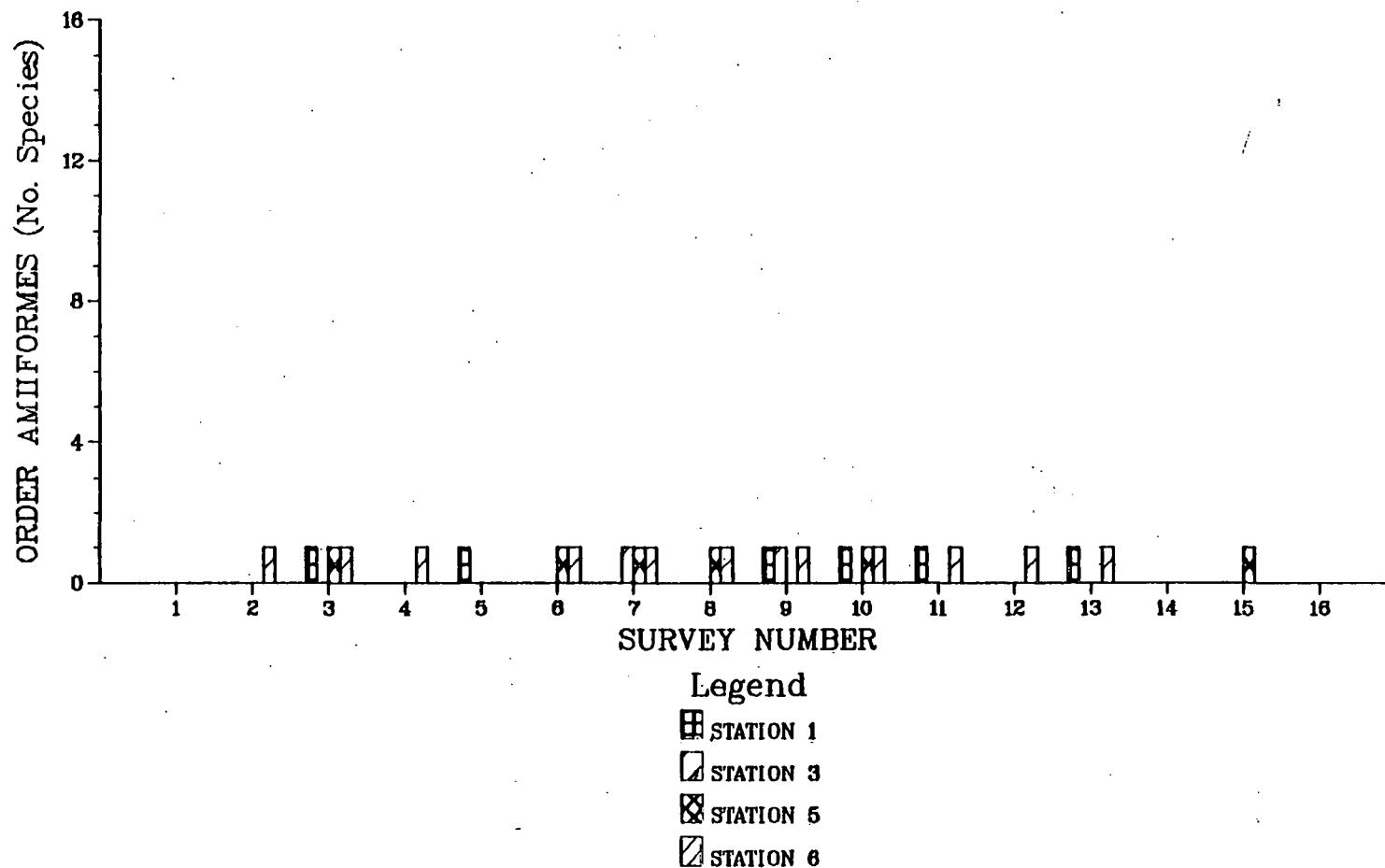


FIGURE B.1.26

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Eels)*

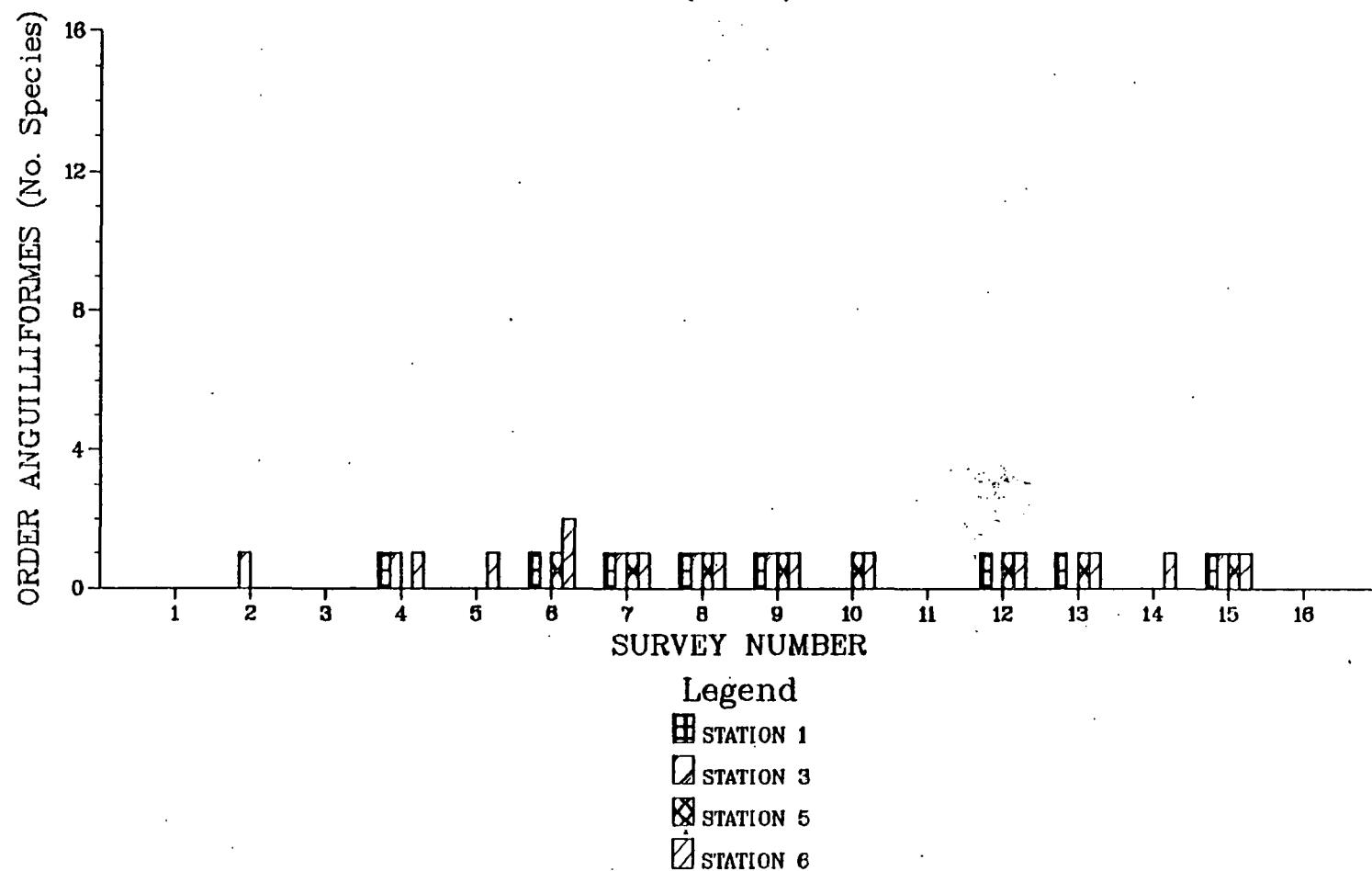


FIGURE B.1.27

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(*Silversides*)*

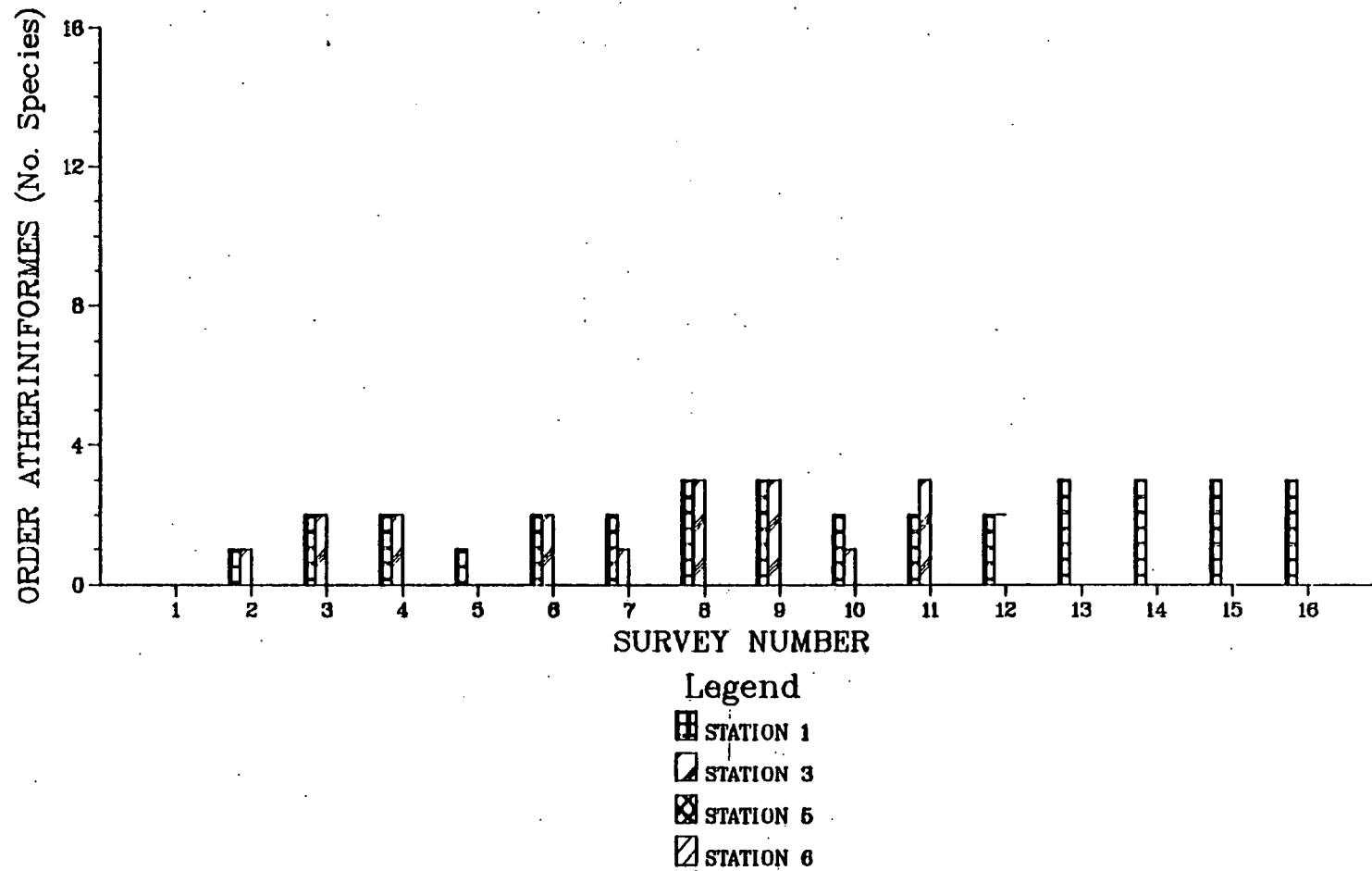


FIGURE B.1.28

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Herring)*

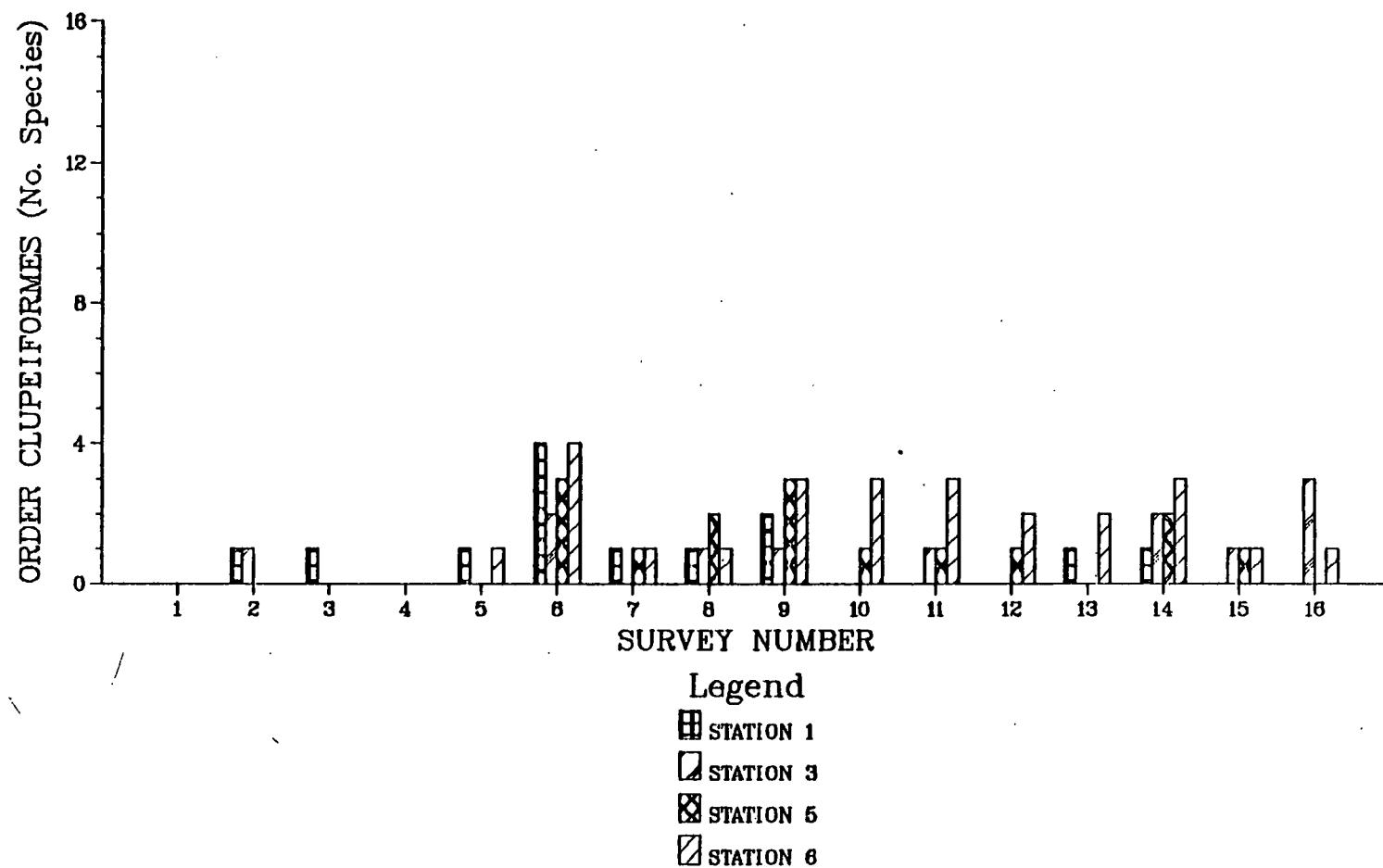


FIGURE B.1.29

PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Suckers, Minnows)

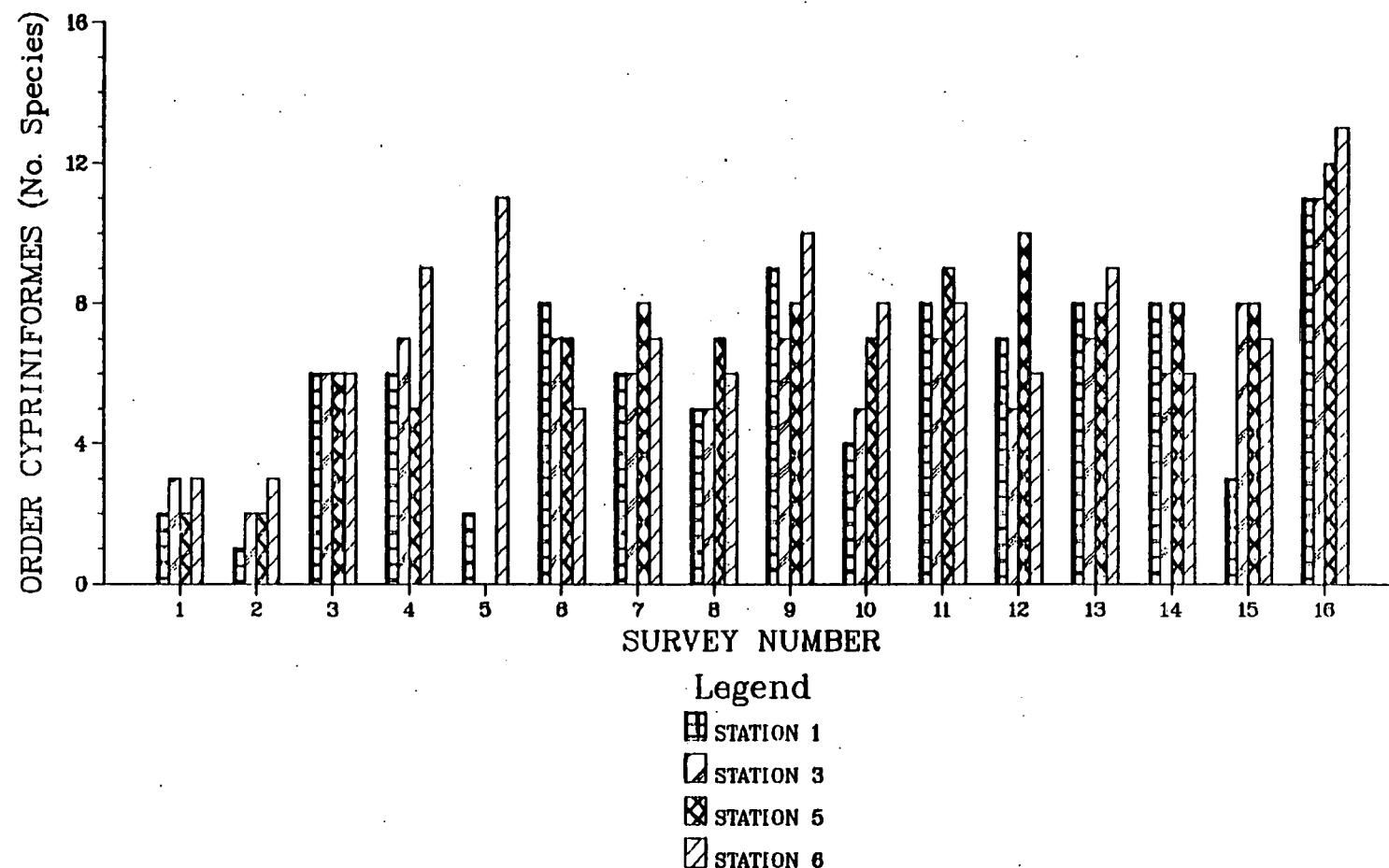


FIGURE B.1.30

PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Sunfish, Perch)

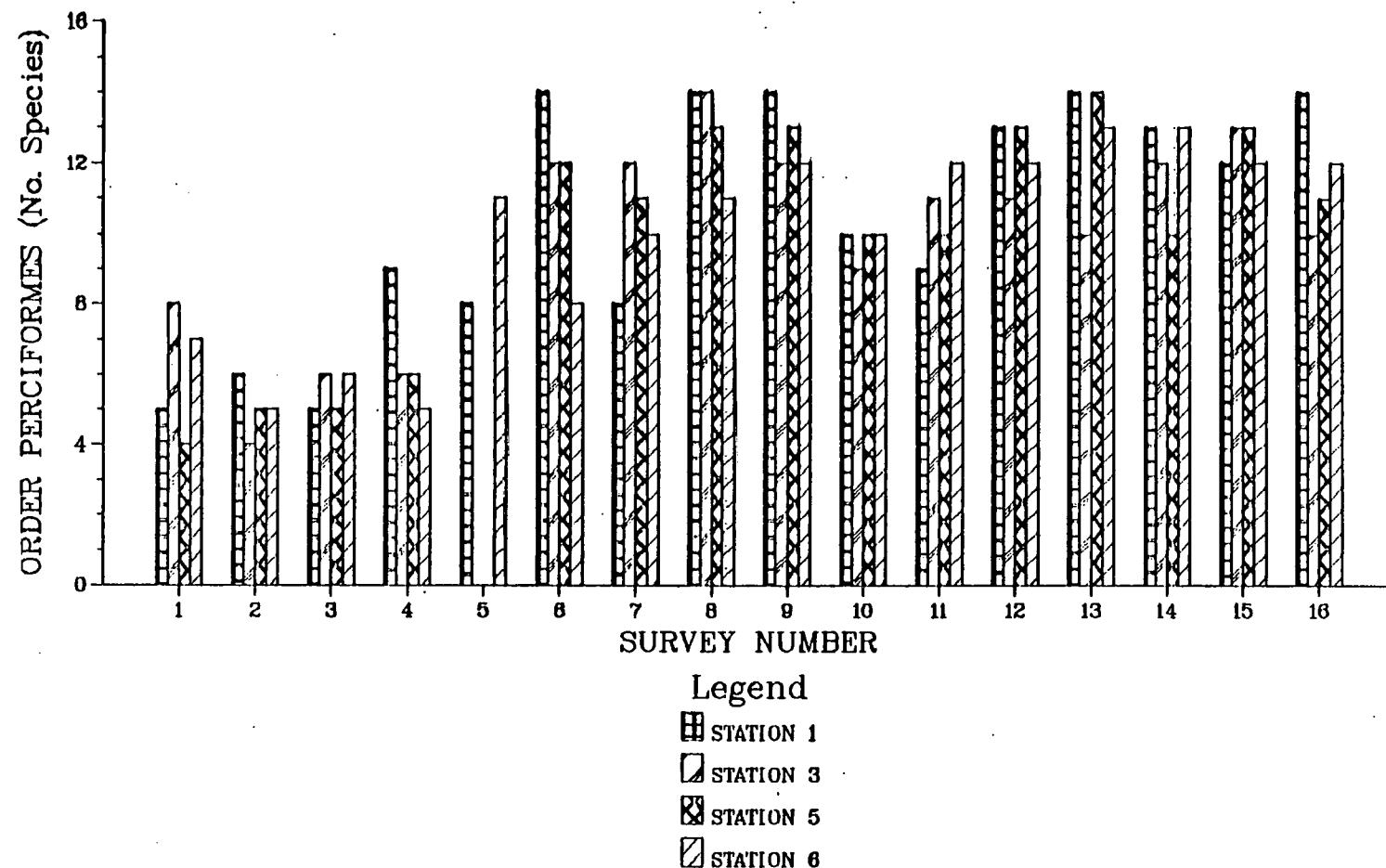


FIGURE B.1.31

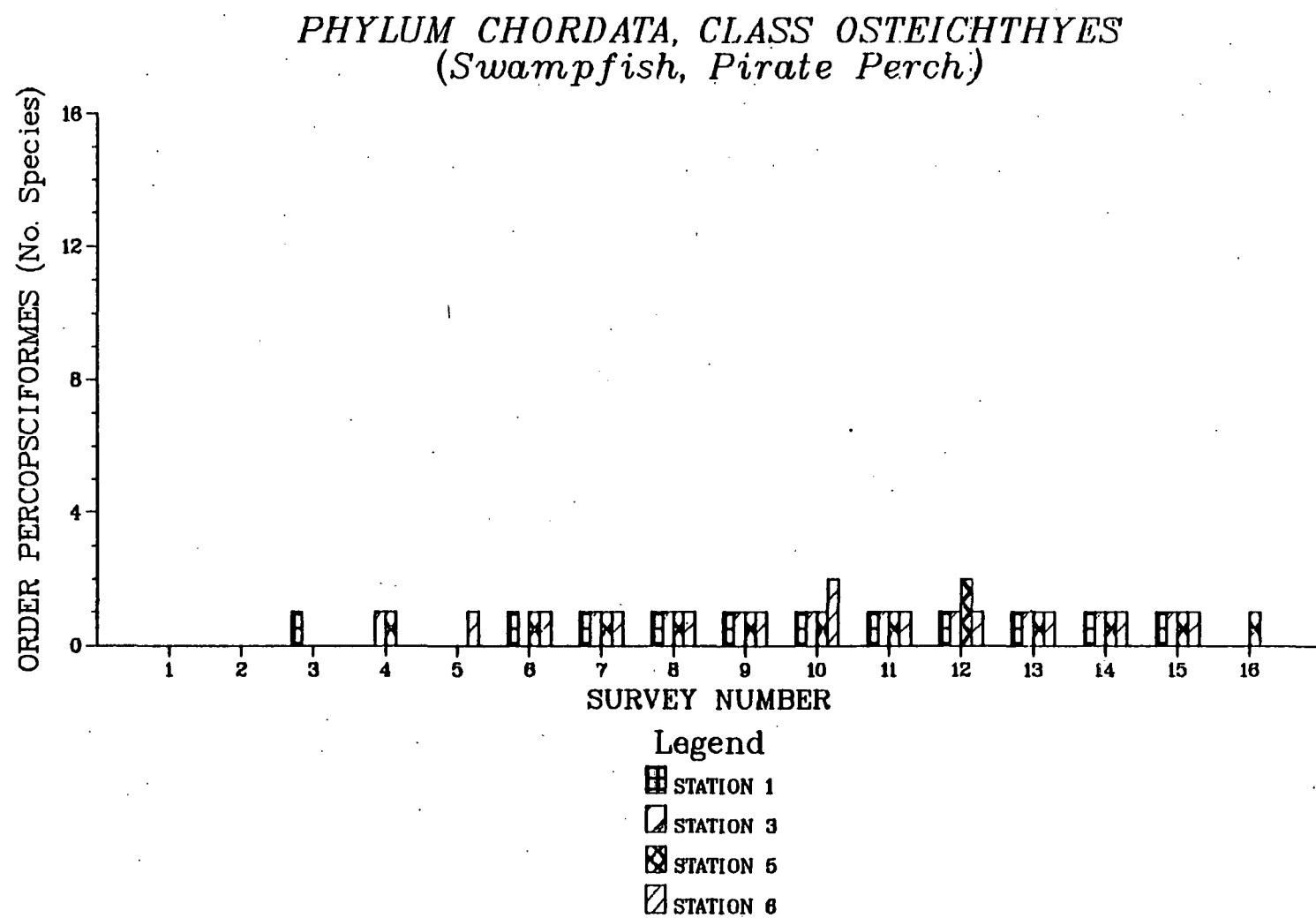


FIGURE B.1.32

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Flounders)*

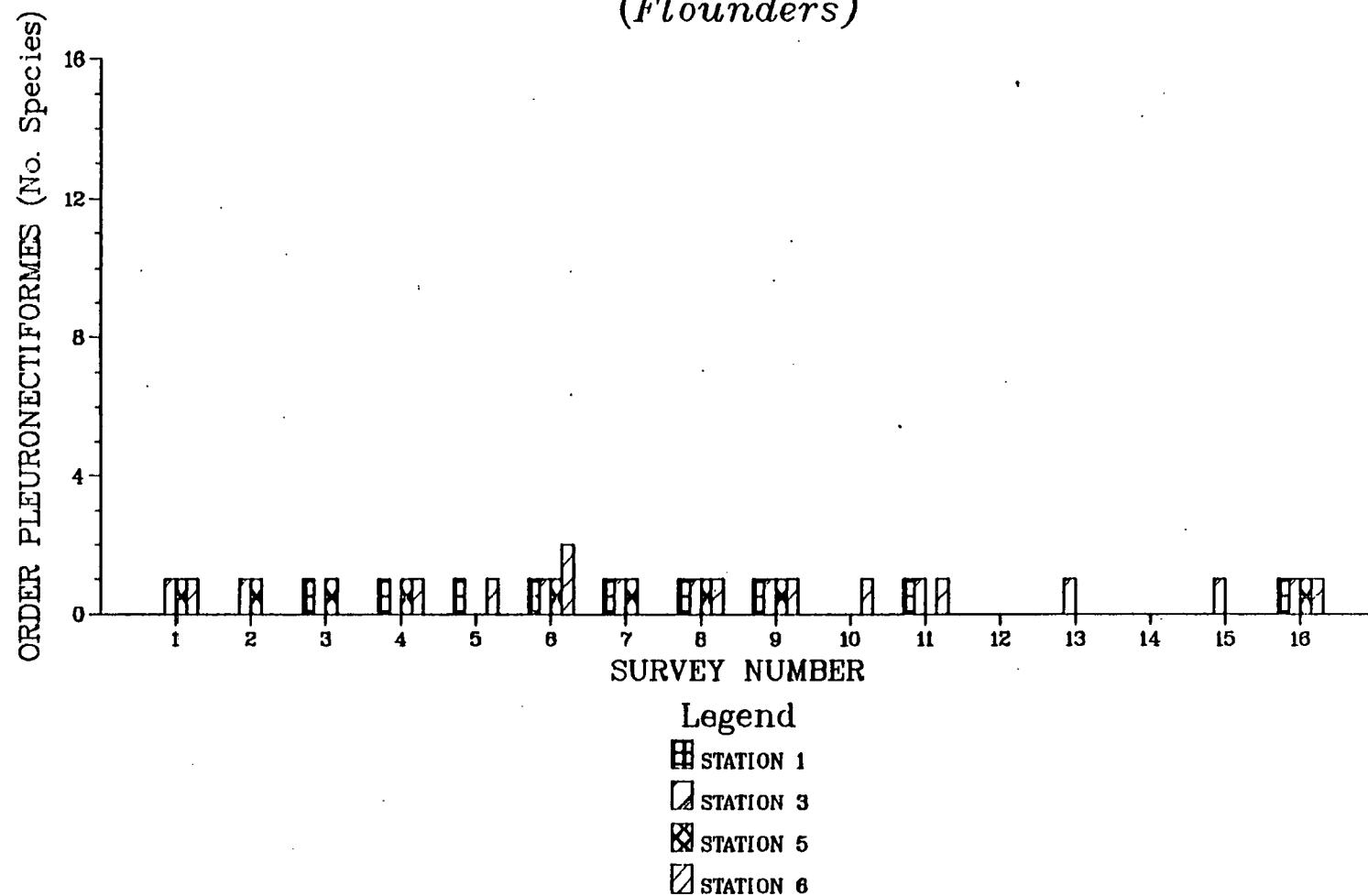


FIGURE B.1.33

PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Pike, Mudminnows.)

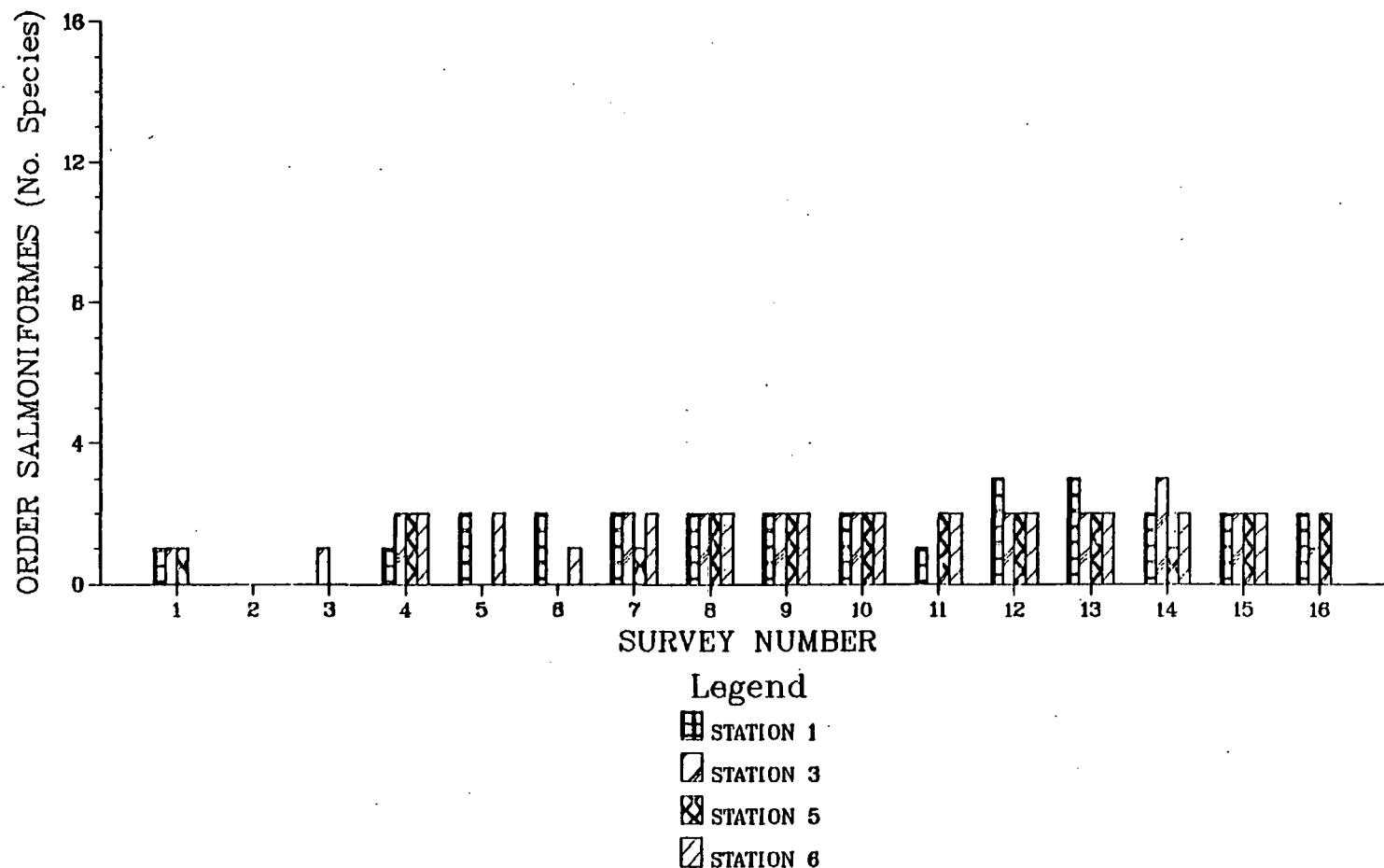


FIGURE B.1.34

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Gars)*

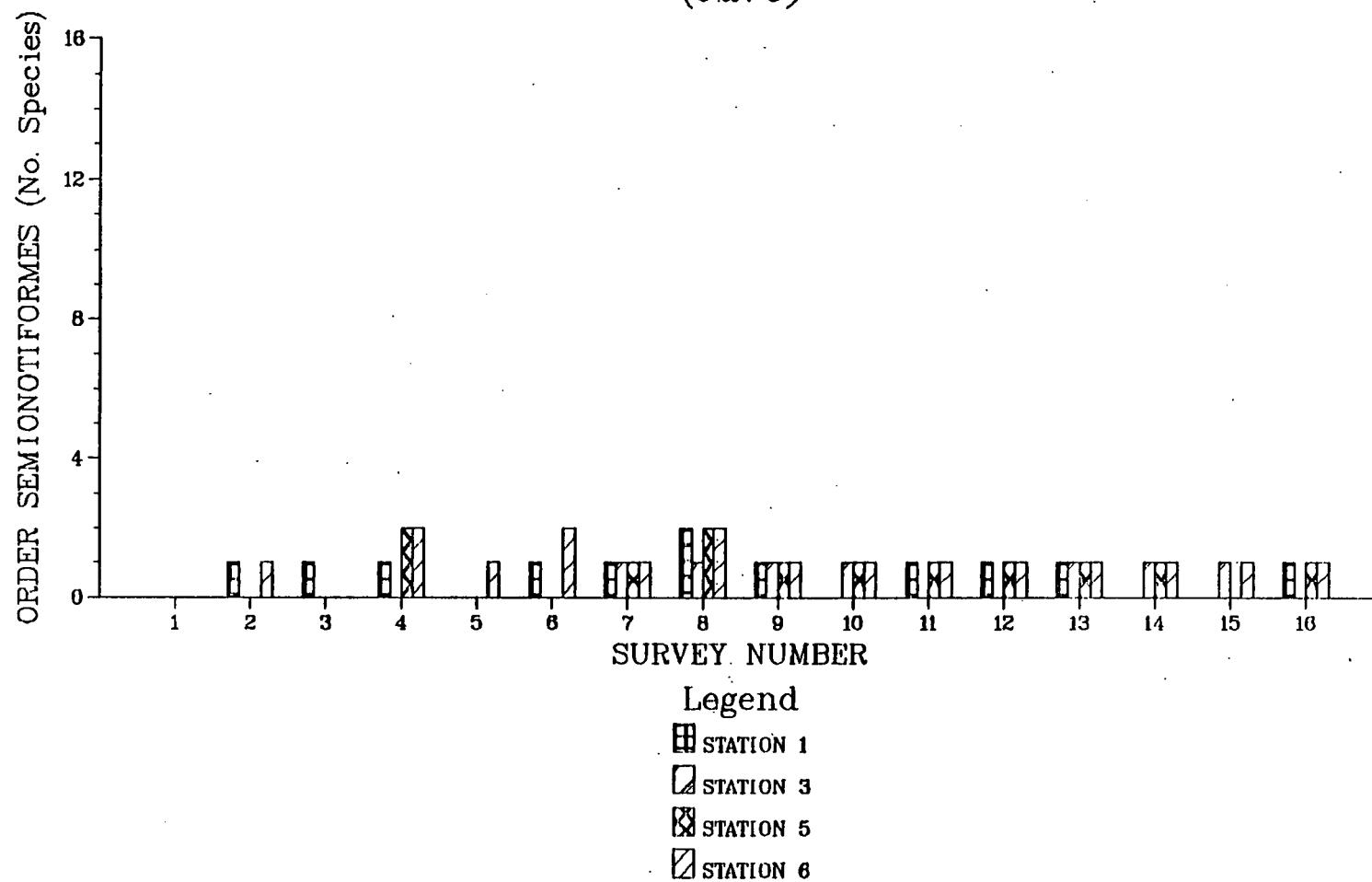


FIGURE B.1.35

*PHYLUM CHORDATA, CLASS OSTEICHTHYES
(Cat fish)*

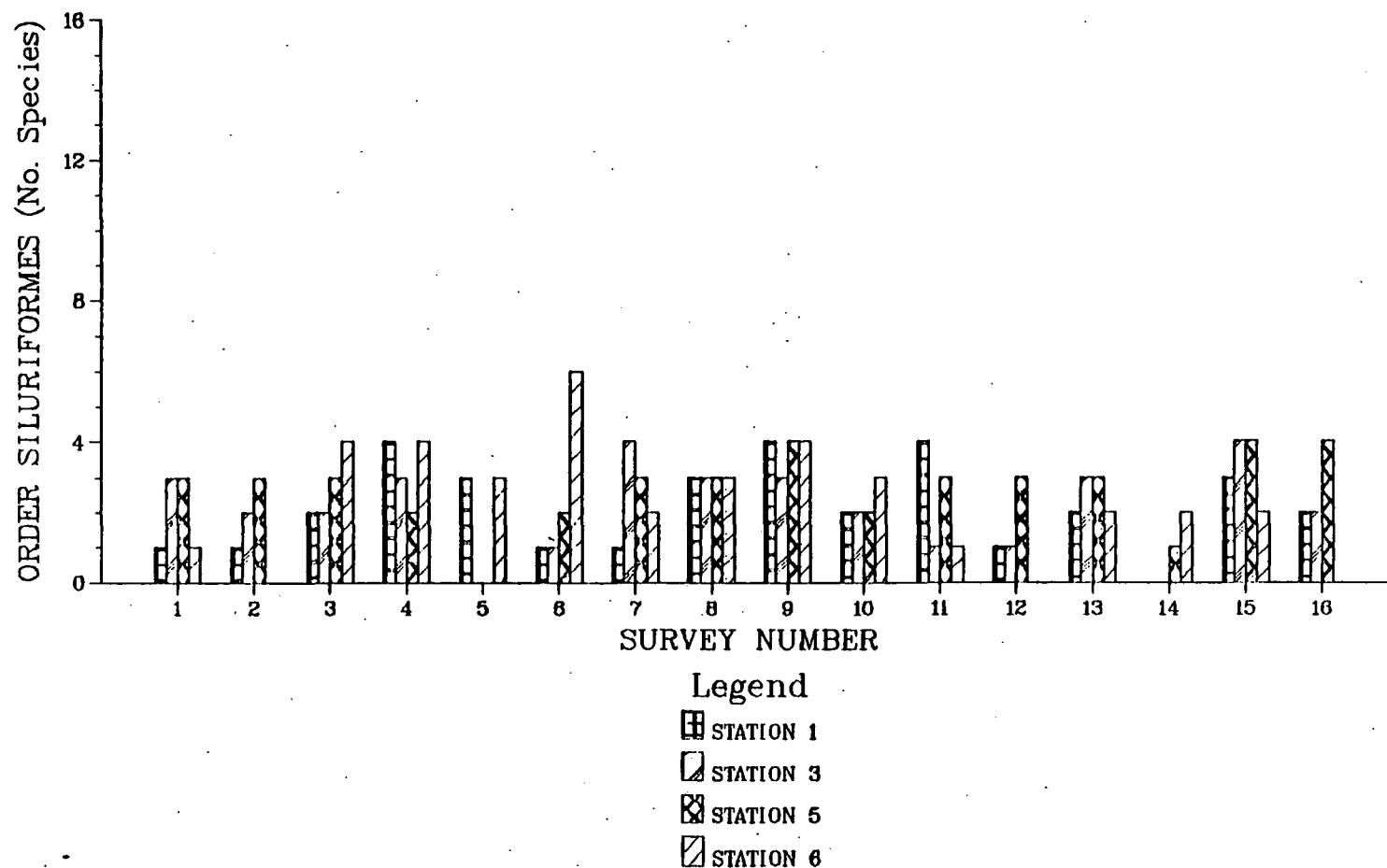


FIGURE B.1.36

B.2. Sample Presence/Absence Data for Two Diatom Species

A complete Presence/Absence list is available for each biological organism identified during the surveys. This list is quite extensive. Two representative species from the diatom genus, Gomphonema, have been chosen to show the general format:

G. parvulum Kutz.

Survey	1 Station 1,3,5,6
Survey	2 Station 1,3,5,6
Survey	3 Station 1,3,5,6
Survey	4 Station 1,3,5,6
Survey	5 Station 1,6
Survey	6 Station 1,3,5,6
Survey	7 Station 1,3,5,6
Survey	8 Station 1,3,5,6
Survey	9 Station 1,3,5,6
Survey	10 Station 1,3,5,6
Survey	11 Station 1,3,5,6
Survey	12 Station 1,3,5,6
Survey	13 Station 1,3,5,6
Survey	14 Station 1,3,5,6
Survey	15 Station 1,3,5,6
Survey	16 Station 1,3,5,6

G. truncatum v. capitatum (Ehr.)

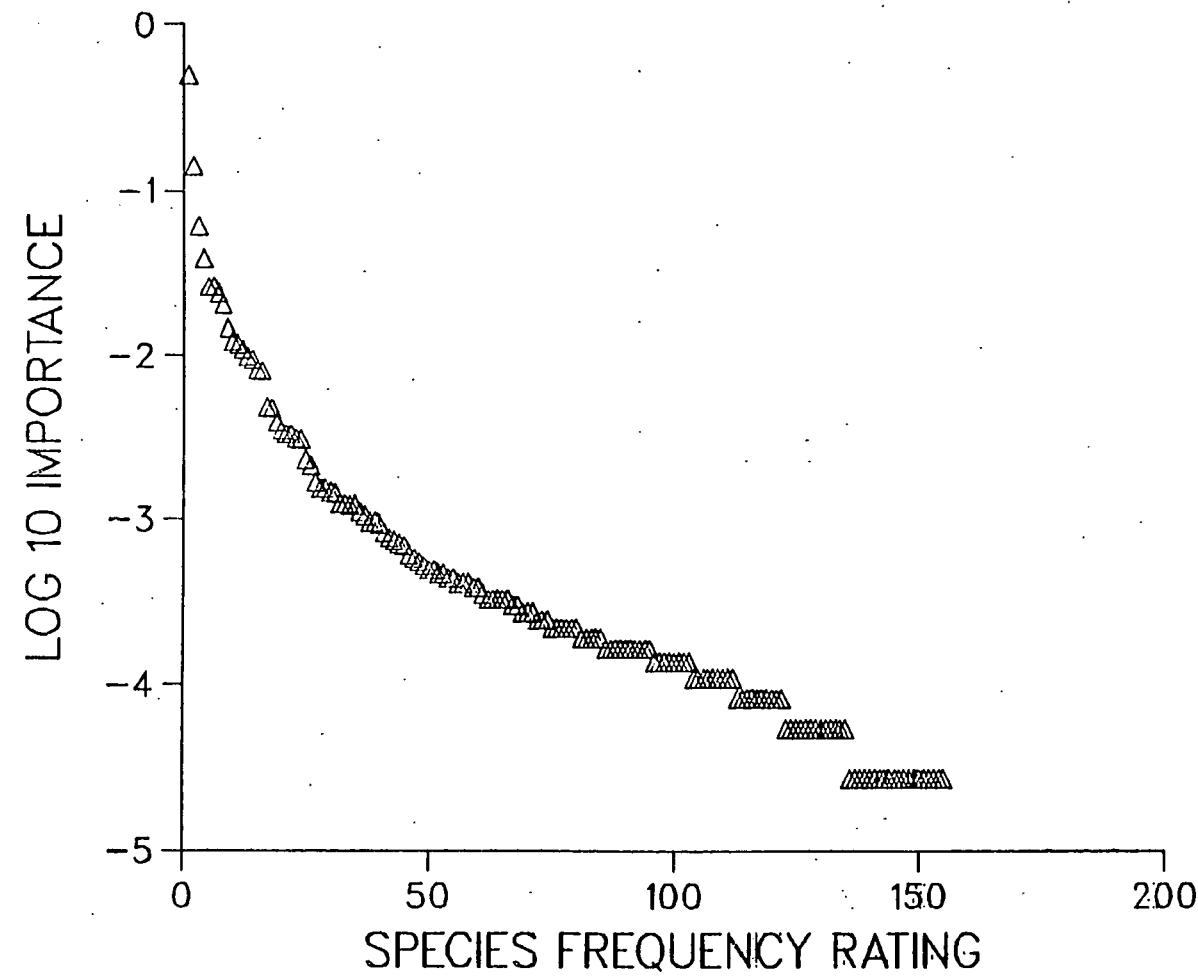
Patr.	
	Survey 7 Station 1,
	Survey 8 Station 1,5

These data indicate that Gomphonema parvulum was present during all the surveys, at nearly all stations. By contrast, G. truncatum v. capitatum was rarely encountered.

B.3. Sample Diatometer Data

One way to format the diatometer frequency data is to graph species frequency distributions for each station by survey. Two examples of this type of format are shown in Figures B.3.1 and B.3.2. These figures reflect two different community dominance patterns. Figure B.3.1 indicates that the diatom community was dominated by Gomphonema parvulum, which represented about 50 percent of the total number of individuals counted. Figure B.3.2 shows a diatom community that contains several dominant species, none of which have frequencies greater than about 12 percent.

FIGURE B.3.1 DIATOMETER FREQUENCY DISTRIBUTION
STATION 1 (FALL, 1953)



LOG 10 IMPORTANCE = $\log(N_i/N)$

WHERE N_i = DIATOM COUNT AT INDEX i , N = TOTAL DIATOM COUNT

FIGURE B.3.1. Diatom Frequency Distribution at Station 1 (Fall 1953)

SPECIES FREQUENCY RATING	% OF TOTAL DIATOM COUNT
1 Gomphonema parvulum Kutz.....	.50.0780
2 G. intermedia Grun.....	.14.0500
3 Melosira varians Ag.....	.6.1164
4 Synedra rumpens Kutz.....	.3.9408
5 Gomphonema grunowii Patr.....	.2.6600
6 Synedra socia Wall.....	.2.6600
7 S.ulna (Nitz.) Ehr.....	.2.4247
8 S.minuscula Grun.....	.2.0689
9 Fragilaria vaucheriae (Kutz.) Peters.....	.1.4778
10 Melosira alpigena Grun.....	.1.2288
11 Cymbella minuta Hilse ex Rabh.....	.1.1740
12 Synedra familiaris (Kutz.) Hust.....	.1.0837
13 S.meneghiniana Grun.....	.9852
14 Frustulia rhomboides (Ehr.) DeT.....	.9469
15 Cyclotella stelligera Cl. & Grun.....	.8237
16 Navicula nr. canalis Patr.....	.8073
17 Synedra scotica Grun.....	.4926
18 Tabellaria flocculosa (Roth) Kutz.....	.4789
19 Gomphonema gracile Ehr. emend. V. H.....	.3941
20 Eunotia naegelii Migula.....	.3503
21 Melosira angustissima Mull.....	.3284
22 Navicula lateropunctata Wallace.....	.3284
23 Cymbella affinis Kutz.....	.3092
24 Achnanthes minutissima Kutz.....	.3065
25 Gomphonema clevei Fricke.....	.2299
26 Eunotia maior (W. Sm.) Rabh.....	.2107
27 Cymbella pseudogracilis (Choln.) Reim.....	.1669
28 Navicula germainii (Wall.) Patr.....	.1533
29 Frustulia crassinervia (Breb. ex W. Sm.) Ross.....	.1533
30 Nitzschia kutzingiana Hilse.....	.1450
31 Frustulia vulgaris (Thwaites) DeT.....	.1423
32 Eunotia minor (Kutz.) Rabh.....	.1231
33 Cymbella silesiaca (Bleisch ex Rabh.) Reim.....	.1231
34 C. lunata W. Sm.....	.1204
35 Synedra tenera W. Sm.....	.1204
36 Tabellaria fenestrata (Lyngb.) Kutz.....	.1095
37 Navicula mutica Kutz.....	.1040
38 Amphipleura pellucida Kutz.....	.0957
39 Surirella delicatissima Lewis.....	.0957
40 Gyrosigma spencerii (Quek.) Griff. & Henfr.....	.0930
41 Eunotia curvata (Kutz.) Lagerst.....	.0821
42 Pinnularia biceps Greg.....	.0766
43 Eunotia tenella (Grun.) Cl.....	.0738
44 Navicula symmetrica Patr.....	.0711
45 N.escambia Patr.....	.0684
46 Capartogramma crucicula (Grun. ex Cl.) Ross.....	.0602
47 Coccconeis lineata (Ehr.) V. H.....	.0574
48 Frustulia weinholdii Hust.....	.0547
49 Nitzschia frustulum Kuta.....	.0520
50 Gomphonema capitatum (Ehr.) Patr.....	.0492

FIGURE B.3.1. (Contd)

SPECIES FREQUENCY RATING	% OF TOTAL DIATOM COUNT
51 <i>Pinnularia abaujensis</i> (Pant.) Ross.....	.0492
52 <i>Nitzschia clausii</i> Hantz.....	.0465
53 <i>Synedra oxyrhynchus</i> (Kutz.) V. H.....	.0465
54 <i>Nitzschia filiformis</i> (W. Sm.) Hust.....	.0437
55 <i>Navicula linearis</i> Hust.....	.0437
56 <i>Eunotia undulata</i> (Ralfs) Rabh.....	.0410
57 <i>Synedra angustissima</i> Grun.....	.0410
58 <i>Fragilaria subcapitata</i> Freng.....	.0410
59 <i>Achnanthes heterovalva</i> Krasske.....	.0383
60 <i>Navicula menisculus</i> Schum.....	.0383
61 <i>Achnanthes abbreviata</i> Reim.....	.0355
62 <i>Navicula pupula</i> Kutz.....	.0328
63 <i>Cymbella naviculiformis</i> Auersw. ex Heib.....	.0328
64 <i>Navicula biceps</i> (Arn.) V. H.....	.0328
65 <i>Opephora martyi</i> Herib.....	.0328
66 <i>Bacillaria paradoxa</i> Gmel.....	.0328
67 <i>Eunotia zasuminensis</i> (Cabej.) Korn.....	.0301
68 <i>Melosira italica</i> (Ehr.) Kutz.....	.0301
69 <i>Nitzschia intermedia</i> Hantz. ex Cl. & Grun.....	.0273
70 <i>Cymbella aspera</i> (Ehr.) H. Per.....	.0273
71 <i>Navicula cryptocephala</i> Kutz.....	.0273
72 <i>Cyclotella meneghiniana</i> Kutz.....	.0246
73 <i>Pinnularia obscura</i> Krasske.....	.0246
74 <i>Surirella ovata</i> Kutz.....	.0246
75 <i>Navicula schizonemoides</i> (V. H.) Patr.....	.0218
76 <i>Epithemia adnata</i> (Kutz.) Breb.....	.0218
77 <i>E.turgida</i> (Ehr.) Kutz.....	.0218
78 <i>Achnanthes stewartii</i> Patr.....	.0218
79 <i>Nitzschia amphibia</i> Grun.....	.0218
80 <i>Synedra</i> sp. l.....	.0218
81 <i>Anomoeoneis vitrea</i> (Grun.) Ross.....	.0191
82 <i>Coccconeis thumensis</i> A. May.....	.0191
83 <i>Navicula grimmiei</i> Krasske.....	.0191
84 <i>Diploneis oblongella</i> (Naeg. ex Kutz.) Ross.....	.0191
85 <i>Melosira undulata</i> (Ehr.) Kutz.....	.0191
86 <i>Achnanthes lanceolata</i> (Breb.) Grun.....	.0164
87 <i>Stauroneis phoenicenteron</i> (Nitz.) Ehr.....	.0164
88 <i>Amphora pediculus</i> (Kutz.) V. H. ex DeT.....	.0164
89 <i>Navicula capitata</i> Skv. & Mey.....	.0164
90 <i>N.canalis</i> Patr.....	.0164
91 <i>N. confervacea</i> (Kutz.) Grun.....	.0164
92 <i>Neidium amphirhynchus</i> (Ehr.) Cl.....	.0164
93 <i>Nitzschia tenuis</i> (W. Sm.) Grun.....	.0164
94 <i>Coccconeis fluviatilis</i> Wall.....	.0164
95 <i>Caloneis bacillum</i> (Grun.) Cl.....	.0164
96 <i>Navicula aikenensis</i> Patr.....	.0136
97 <i>N. anglica</i> Ralfs.....	.0136
98 <i>Cyclotella pseudostelligera</i> Hust.....	.0136
99 <i>Gomphonema brasiliense</i> Grun.....	.0136
100 <i>Nitzschia parvula</i> Lewis.....	.0136

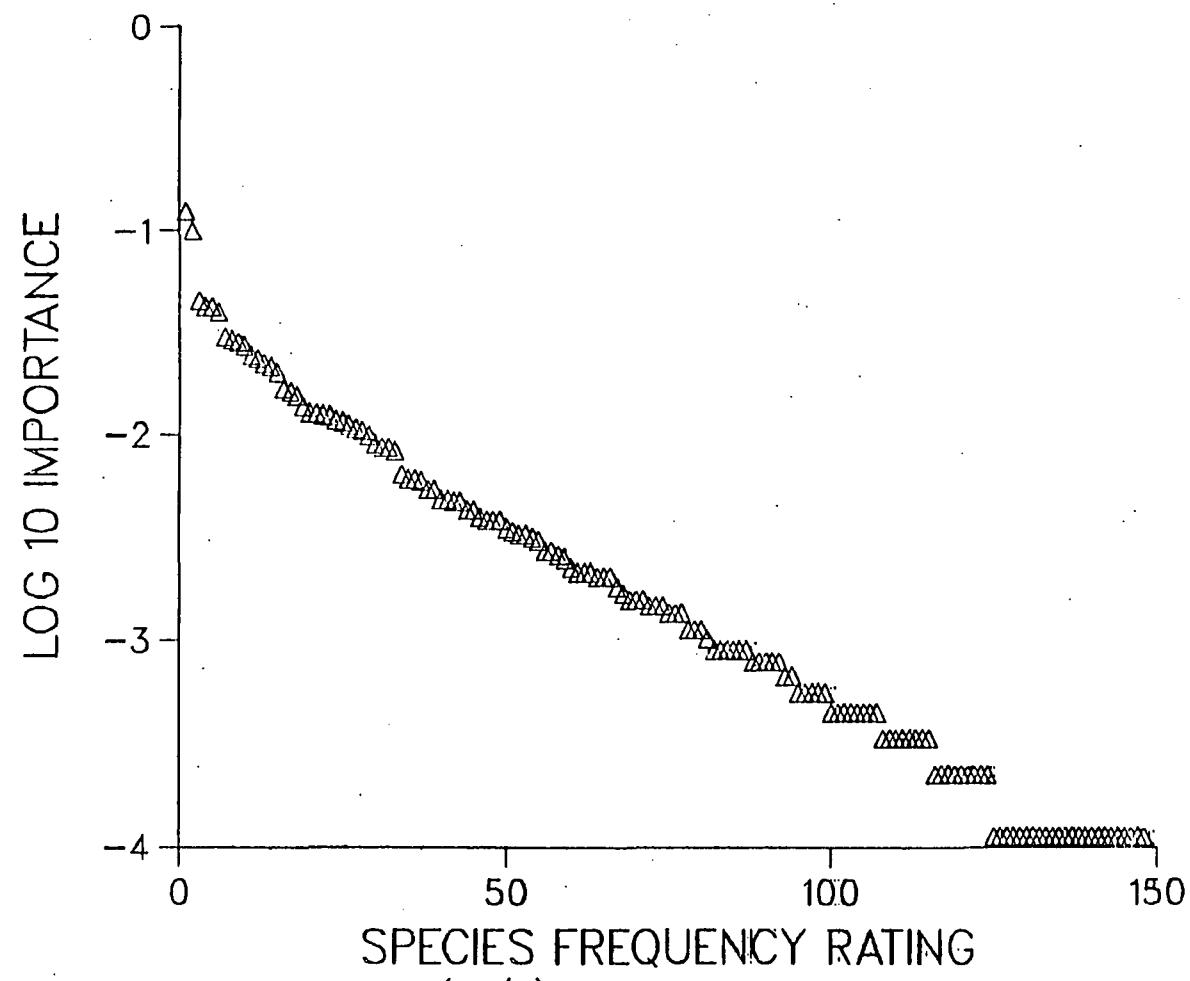
FIGURE B.3.1. (Contd)

SPECIES FREQUENCY RATING	% OF TOTAL DIATOM COUNT
101 <i>N.dissipata</i> (Kutz.) Grun.....	.0136
102 <i>Pinnularia mesogongyla</i> Ehr.....	.0136
103 <i>Surirella suecica</i> Zell.....	.0136
104 <i>Achnanthes dubia</i> Grun.....	.0109
105 <i>Coccconeis euglypta</i> (Ehr.) Cl.....	.0109
106 <i>Frustulia amphipleuroides</i> (Grun.) Cl.....	.0109
107 <i>Hantzschia amphioxys</i> Grun.....	.0109
108 <i>Melosira tenella</i> Nyg.....	.0109
109 <i>Fragilaria pinnata</i> Ehr.....	.0109
110 <i>Navicula seminoidea</i> Cl. & Grun.....	.0109
111 <i>Amphora submontana</i> Hust.....	.0109
112 <i>Surirella nervosa</i> A. Schm.....	.0109
113 <i>Synedra ramesi</i> (Herib.) Hust.....	.0082
114 <i>Pinnularia maior</i> (Kutz.) Rabh.....	.0082
115 <i>Navicula minima</i> Grun.....	.0082
116 <i>Nitzschia palea</i> (Kutz.) W. Sm.....	.0082
117 <i>Gyrosigma nodiferum</i> (Grun.) Reim.....	.0082
118 <i>Surirella angusta</i> Kutz.....	.0082
119 <i>Navicula caroliniana</i> Patr.....	.0082
120 <i>Neidium hitchcockii</i> (Ehr.) Cl.....	.0082
121 <i>Achnanthes apiculata</i> Patr.....	.0082
122 <i>Cymbella tumida</i> (Breb. ex Kutz.) V. H.....	.0082
123 <i>Amphora perpusilla</i> (Grun.) Grun.....	.0054
124 <i>Entomoneis ornata</i> (J. W. Bail.) Reim.....	.0054
125 <i>Caloneis lewisi</i> Patr.....	.0054
126 <i>Navicula capitata</i> Patr.....	.0054
127 <i>Melosira tenuissima</i> (Grun.) Mull.....	.0054
128 <i>Navicula laevissima</i> Kutz.....	.0054
129 <i>Nitzschia debilis</i> (Arn.) A. May.....	.0054
130 <i>N. victoriae</i> Grun.....	.0054
131 <i>N.acicularis</i> W. Sm.....	.0054
132 <i>Stauroneis livingstonii</i> Reim.....	.0054
133 <i>Gomphonema grovei</i> M. Schmidt.....	.0054
134 <i>Navicula tridentula</i> Krasske.....	.0054
135 <i>Caloneis hyalina</i> Hust.....	.0054
136 <i>Navicula capitata</i> Ehr.....	.0027
137 <i>Diploneis oculata</i> (Breb.) Cl.....	.0027
138 <i>Achnanthes clevei</i> Grun.....	.0027
139 <i>Nitzschia gracilis</i> Hantz.....	.0027
140 <i>Melosira granulata</i> (Ehr.) Ralfs.....	.0027
141 <i>Navicula thienemannii</i> Hust.....	.0027
142 <i>N.contraria</i> Patr.....	.0027
143 <i>Nitzschia subtilis</i> Grun.....	.0027
144 <i>N.mediastalsis</i> Hohn & Hell.....	.0027
145 <i>Pinnularia nodosa</i> (Ehr.) W. Sm.....	.0027
146 <i>Achnanthes flexella</i> (Kutz.) Brun.....	.0027
147 <i>Rhopalodia gibberula</i> (Ehr.) O. Mull.....	.0027
148 <i>Navicula nivalis</i> (Ehr.) Hust.....	.0027
149 <i>N.cuspidata</i> (Kutz.) Kutz.....	.0027
150 <i>Stauroneis smithii</i> Grun.....	.0027

FIGURE B.3.1. (Contd)

SPECIES	FREQUENCY	RATING	% OF TOTAL DIATOM COUNT
151	<i>Surirella linearis</i> W. Sm.....		.0027
152	<i>Amphora normanii</i> Rabh.....		.0027
153	<i>Surirella stalagma</i> Hohn & Hell.....		.0027
154	<i>Synedra danica</i> (Kutz.) V. H.....		.0027
155	<i>Nitzschia subconstricta</i> Grun.....		.0027

FIGURE B.3.2 DIATOMETER FREQUENCY DISTRIBUTION
STATION 6 (WINTER, 1954)



LOG 10 IMPORTANCE = LOG(N_i/N)
WHERE N_i = DIATOM COUNT AT INDEX i , N = TOTAL DIATOM COUNT

FIGURE B.3.2. Diatom Frequency Distribution at Station 6 (Winter 1954)

SPECIES FREQUENCY RATING	% OF TOTAL DIATOM COUNT
1 <i>Synedra ulna</i> (Nitz.) Ehr.....	.12.4112
2 <i>Gomphonema parvulum</i> Kutz.....	.9.9650
3 <i>Melosira alpigena</i> Grun.....	.4.5316
4 <i>Navicula cryptocephala</i> Kutz.....	.4.2273
5 <i>Synedra oxyrhynchus</i> (Kutz.) V. H.....	.4.1934
6 <i>S.rumpens</i> Kutz.....	.3.9680
7 <i>Melosira varians</i> Ag.....	.2.9985
8 <i>Fragilaria mesolepta</i> Rabh.....	.2.8971
9 <i>Gomphonema affine</i> Kutz.....	.2.8182
10 <i>Achnanthes minutissima</i> Kutz.....	.2.7054
11 <i>Nitzschia palea</i> (Kutz.) W. Sm.....	.2.4349
12 <i>Synedra danica</i> (Kutz.) V. H.....	.2.3447
13 <i>Gomphonema gracile</i> Ehr. emend. V. H.....	.2.2094
14 <i>Synedra augustissima</i> Grun.....	.2.1418
15 <i>Navicula nr. canalis</i> Patr.....	.2.0065
16 <i>Cyclotella stelligera</i> Cl. & Grun.....	.1.6684
17 <i>Synedra familiaris</i> (Kutz.) Hust.....	.1.6120
18 <i>Tabellaria fenestrata</i> (Lyngb.) Kutz.....	.1.5331
19 <i>Nitzschia filiformis</i> (W. Sm.) Hust.....	.1.3640
20 <i>Gomphonema capitatum</i> (Ehr.) Patr.....	.1.2851
21 <i>G.productum</i> Grun.....	.1.2738
22 <i>Synedra socia</i> Wall.....	.1.2513
23 <i>Melosira italica</i> (Ehr.) Kutz.....	.1.2400
24 <i>Gomphonema grunowii</i> Patr.....	.1.1836
25 <i>Nitzschia acicularis</i> W. Sm.....	.1.1498
26 <i>Pinnularia biceps</i> Greg.....	.1.1160
27 <i>Synedra tenera</i> W. Sm.....	.1.0709
28 <i>Cymbella minuta</i> Hilse ex Rabh.....	.1.0371
29 <i>Eunotia naegelii</i> Migula.....	.9807
30 <i>Navicula lateropunctata</i> Wallace.....	.9018
31 <i>Frustulia vulgaris</i> (Thwaites) DeT.....	.8680
32 <i>Navicula mutica</i> Kutz.....	.8680
33 <i>N.rhynchocephala</i> Kutz.....	.8342
34 <i>Surirella angusta</i> Kutz.....	.6425
35 <i>Fragilaria vaucheriae</i> (Kutz.) Peters.....	.6087
36 <i>Cyclotella meneghiniana</i> Kutz.....	.6087
37 <i>Synedra meneghiniana</i> Grun.....	.5975
38 <i>Nitzschia clausii</i> Hantz.....	.5411
39 <i>Surirella ovata</i> Kutz.....	.5411
40 <i>Navicula capitata</i> Ehr.....	.4847
41 <i>Nitzschia parvula</i> Lewis.....	.4847
42 <i>Synedra minuscula</i> Grun.....	.4735
43 <i>Navicula linearis</i> Hust.....	.4735
44 <i>Amphipleura pellucida</i> Kutz.....	.4284
45 <i>Synedra scotica</i> Grun.....	.4284
46 <i>Eunotia maior</i> (W. Sm) Rabh.....	.3945
47 <i>Navicula biceps</i> (Arn.) V. H.....	.3833
48 <i>Cymbella silesiaca</i> (Bleisch ex Rabh.) Reim.....	.3833
49 <i>Frustulia rhomboides</i> (Ehr.) DeT.....	.3833
50 <i>Gyrosigma spencerii</i> (Quek.) Griff. & Henfr.....	.3495

FIGURE B.3.2. (Contd)

SPECIES	FREQUENCY	RATING	% OF TOTAL DIATOM COUNT
51 <i>Eunotia minor</i> (Kutz.) Rabh.....			.3382
52 <i>Cymbella affinis</i> Kutz.....			.3269
53 <i>Nitzschia amphibia</i> Grun.....			.3269
54 <i>Gomphonema clevei</i> Fricke.....			.3156
55 <i>Surirella suecica</i> Zell.....			.3044
56 <i>Achnanthes heterovalva</i> Krasske.....			.2705
57 <i>Navicula germainii</i> (Wall.) Patr.....			.2705
58 <i>N. minima</i> Grun.....			.2593
59 <i>Cymbella tumida</i> (Breb. ex Kutz.) V. H.....			.2480
60 <i>Tabellaria flocculosa</i> (Roth) Kutz.....			.2255
61 <i>Coccconeis lineata</i> (Ehr.) V. H.....			.2142
62 <i>Navicula cohnii</i> (Hilse) Grun.....			.2142
63 <i>Achnanthes lanceolata</i> (Breb.) Grun.....			.2142
64 <i>A. linearis</i> (W. Sm.) Grun.....			.2029
65 <i>Fragilaria spinosa</i> Skv.....			.2029
66 <i>Achnanthes dubia</i> Grun.....			.2029
67 <i>Melosira tenella</i> Nyg.....			.1804
68 <i>Frustulia crassinervia</i> (Breb. ex W. Sm.) Ross.....			.1691
69 <i>Cymbella aspera</i> (Ehr.) H. Per.....			.1578
70 <i>Nitzschia sibirica</i> Skv.....			.1578
71 <i>Navicula rectangularis</i> (Greg.) Grun.....			.1578
72 <i>N. tropica</i> Hust.....			.1465
73 <i>Gomphonema intermedia</i> Grun.....			.1465
74 <i>Eunotia curvata</i> (Kutz.) Lagerst.....			.1465
75 <i>Nitzschia frustulum</i> Kutz.....			.1353
76 <i>Surirella stalagma</i> Holm & Hell.....			.1353
77 <i>Eunotia incisa</i> W. Sm. ex Greg.....			.1353
78 <i>Opephora martyi</i> Herib.....			.1127
79 <i>Melosira tenuissima</i> (Grun.) Mull.....			.1127
80 <i>Synedra ramesi</i> (Herib.) Hust.....			.1127
81 <i>Navicula capitata</i> Skv. & Mey.....			.1015
82 <i>Achnanthes clevei</i> Grun.....			.0901
83 <i>Stauroneis smithii</i> Grun.....			.0901
84 <i>Fragilaria subcapitata</i> Freng.....			.0901
85 <i>Bacillaria paradoxa</i> Gmel.....			.0901
86 <i>Navicula pupula</i> Kutz.....			.0901
87 <i>Nitzschia kutzingiana</i> Hilse.....			.0901
88 <i>Meridion circulare</i> (Grev.) Ag.....			.0789
89 <i>Surirella minuta</i> Breb.....			.0789
90 <i>Pinnularia mesogongyla</i> Ehr.....			.0789
91 <i>Cymbella lunata</i> W. Sm.....			.0789
92 <i>Pinnularia abaujensis</i> (Pant.) Ross.....			.0789
93 <i>Entomoneis ornata</i> (J. W. Bail.) Reim.....			.0676
94 <i>Capartogramma crucicula</i> (Grun. ex Cl.) Ross.....			.0676
95 <i>Gomphonema intricatum</i> Kutz.....			.0563
96 <i>Epithemia adnata</i> (Kutz.) Breb.....			.0563
97 <i>Pinnularia mesolepta</i> (Ehr.) W. Sm.....			.0563
98 <i>Melosira angustissima</i> Mull.....			.0563
99 <i>Nitzschia subtilis</i> Kutz.....			.0563
100 <i>Cymbella naviculiformis</i> Auersw. ex Heib.....			.0450

FIGURE B.3.2. (Contd)

SPECIES	% OF TOTAL
FREQUENCY	DIATOM COUNT
RATING	
101 <i>Amphora ovalis</i> (Kutz.) Kutz.....	.0450
102 <i>Nitzschia bacata</i> Hust.....	.0450
103 <i>Coccconeis diminuta</i> Pant.....	.0450
104 <i>Navicula schonfeldii</i> Hust.....	.0450
105 <i>Pinnularia viridis</i> (Nitz.) Ehr.....	.0450
106 <i>Stauroneis phoenicenteron</i> (Nitz.) Ehr.....	.0450
107 <i>Pinnularia maior</i> (Kutz.) Rabh.....	.0450
108 <i>Eunotia pectinalis</i> (O. F. Mull.?) Rabh.....	.0338
109 <i>Nitzschia recta</i> Hantz.....	.0338
110 <i>Achnanthes abbreviata</i> Reim.....	.0338
111 <i>Cymbella pseudogracilis</i> (Choln.) Reim.....	.0338
112 <i>Anomoeoneis vitrea</i> (Grun.) Ross.....	.0338
113 <i>Synedra parasitica</i> (W. Sm.) Hust.....	.0338
114 <i>Coccconeis euglypta</i> (Ehr.) Cl.....	.0338
115 <i>Gomphonema acuminatum</i> Ehr.....	.0338
116 <i>Pinnularia microstauron</i> (Ehr.) Cl.....	.0225
117 <i>Navicula accomoda</i> Hust.....	.0225
118 <i>N. atomus</i> (Kutz.) Grun.....	.0225
119 <i>Eunotia exigua</i> (Breb. ex Kutz.) Rabh.....	.0225
120 <i>Surirella linearis</i> W. Sm.....	.0225
121 <i>Fragilaria pinnata</i> Ehr.....	.0225
122 <i>Nitzschia gracilis</i> Hantz.....	.0225
123 <i>Coccconeis placentula</i> Ehr.....	.0225
124 <i>Navicula capitata</i> Patr.....	.0225
125 <i>Caloneis bacillum</i> (Grun.) Cl.....	.0112
126 <i>Gyrosigma nodiferum</i> (Grun.) Reim.....	.0112
127 <i>Navicula grimmiei</i> Krasske.....	.0112
128 <i>N. lacustris</i> Greg.....	.0112
129 <i>Nitzschia subconstricta</i> Grun.....	.0112
130 <i>Gomphonema brasiliense</i> Grun.....	.0112
131 <i>Gyrosigma scalpoides</i> (Rabh.) Cl.....	.0112
132 <i>Nitzschia perminuta</i> Grun.....	.0112
133 <i>Amphora pediculus</i> (Kutz.) V. H. ex DeT.....	.0112
134 <i>Gomphonema grovei</i> M. Schmidt.....	.0112
135 <i>Nitzschia subtilis</i> Grun.....	.0112
136 <i>Caloneis hyalina</i> Hust.....	.0112
137 <i>Neidium amphirhynchus</i> (Ehr.) Cl.....	.0112
138 <i>N. bisulcatum</i> (Lagerst.) Cl.....	.0112
139 <i>Gomphonema subclavatum</i> (Grun.) Grun.....	.0112
140 <i>Nitzschia debilis</i> (Arn.) A. May.....	.0112
141 <i>N. victoriae</i> Grun.....	.0112
142 <i>Stenopterobia intermedia</i> (Lewis) V. H.....	.0112
143 <i>Coccconeis thumensis</i> A. May.....	.0112
144 <i>Diploneis oblongella</i> (Naeg. ex Kutz.) Ross.....	.0112
145 <i>Denticula tenuis</i> Kutz.....	.0112
146 <i>Navicula bicephala</i> Hust.....	.0112
147 <i>Melosira undulata</i> (Ehr.) Kutz.....	.0112
148 <i>Synedra</i> sp. 1.....	.0112

B.4. Savannah River Water Chemistry Summary

The water chemistry data may be formated separately from, or in combination with, the biological data. Figures B.4.1-B.4.23 show one type of independent format. In these figures, the water chemistry data are plotted by station for each parameter over the entire survey period. This type of format may be used to look for long-term trends or unusual station values.

Cross formatting between water chemistry data and biological data may be used for a variety of purposes. One type of format lists the chemical ranges encountered for each survey organism. For example, the diatom species Gomphonema parvulum was found in the following chemical ranges:^{*}

0.145E+02	<	ALKALMO	<	0.336E+02
0.0	<	ALKALP	<	0.0
0.100E+00	<	BOD	<	0.290E+01
0.200E+01	<	CA	<	0.584E+01
0.600E+01	<	CACO3	<	0.160E+02
0.600E+00	<	CL	<	0.130E+02
0.900E+01	<	CLEAR	<	0.360E+02
0.155E+02	<	COD	<	0.635E+02
0.360E+01	<	CO2	<	0.106E+02
0.0	<	CO3	<	0.0
0.585E+01	<	DO	<	0.107E+02
0.600E-02	<	FE	<	0.141E+01

* Chemical abbreviations are identified in Chapter VIII.D.

B.4 (Contd)

0.160E+02	<	FIXR	<	0.158E+03
0.800E+01	<	HARD	<	0.340E+02
0.200E+02	<	HC03	<	0.390E+02
0.120E+01	<	K	<	0.140E+01
0.340E+00	<	MG	<	0.333E+01
0.300E+01	<	MGC03	<	0.180E+02
0.600E-01	<	MN	<	0.195E+00
0.570E+01	<	NA	<	0.990E+01
0.100E-02	<	NH3-N	<	0.270E+00
0.100E-02	<	NO2-N	<	0.154E-01
0.300E-01	<	NO3-N	<	0.539E+00
0.650E+01	<	PH	<	0.780E+01
0.400E-02	<	P04-P	<	0.860E-01
0.434E+01	<	SI02	<	0.146E+02
0.100E+01	<	S04	<	0.110E+02
0.290E-04	<	SPCOND	<	0.683E+03
0.900E+01	<	SS	<	0.130E+02
0.995E+02	<	TDS	<	0.187E+03
0.110E+02	<	TEMP	<	0.295E+02
0.700E-01	<	TKN	<	0.118E+01
0.160E+00	<	TOTP	<	0.220E+00
0.280E+02	<	TS	<	0.250E+03
0.105E+02	<	TURB	<	0.548E+03
0.00	<	VS	<	0.234E+03

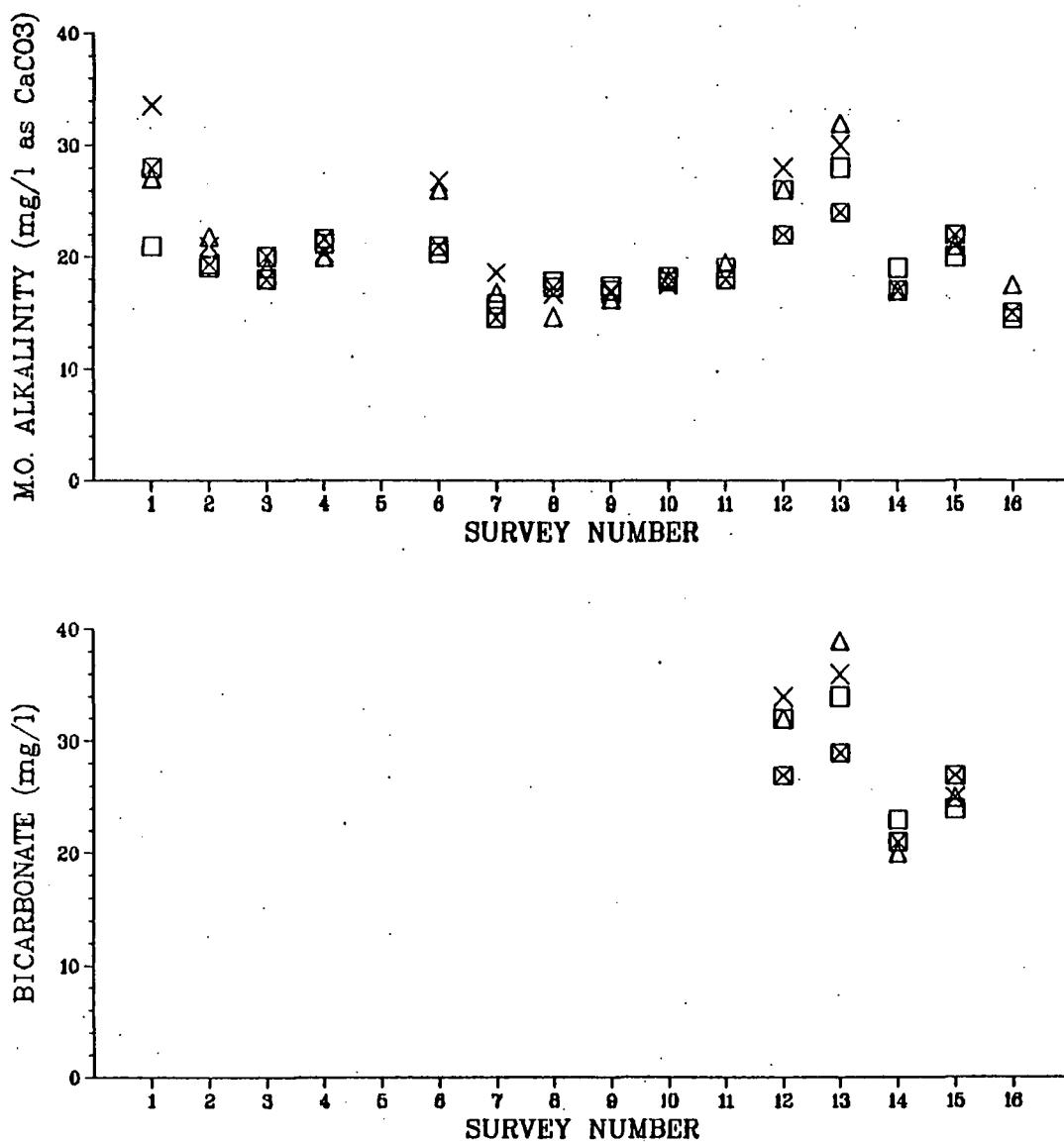


FIGURE B.4.1

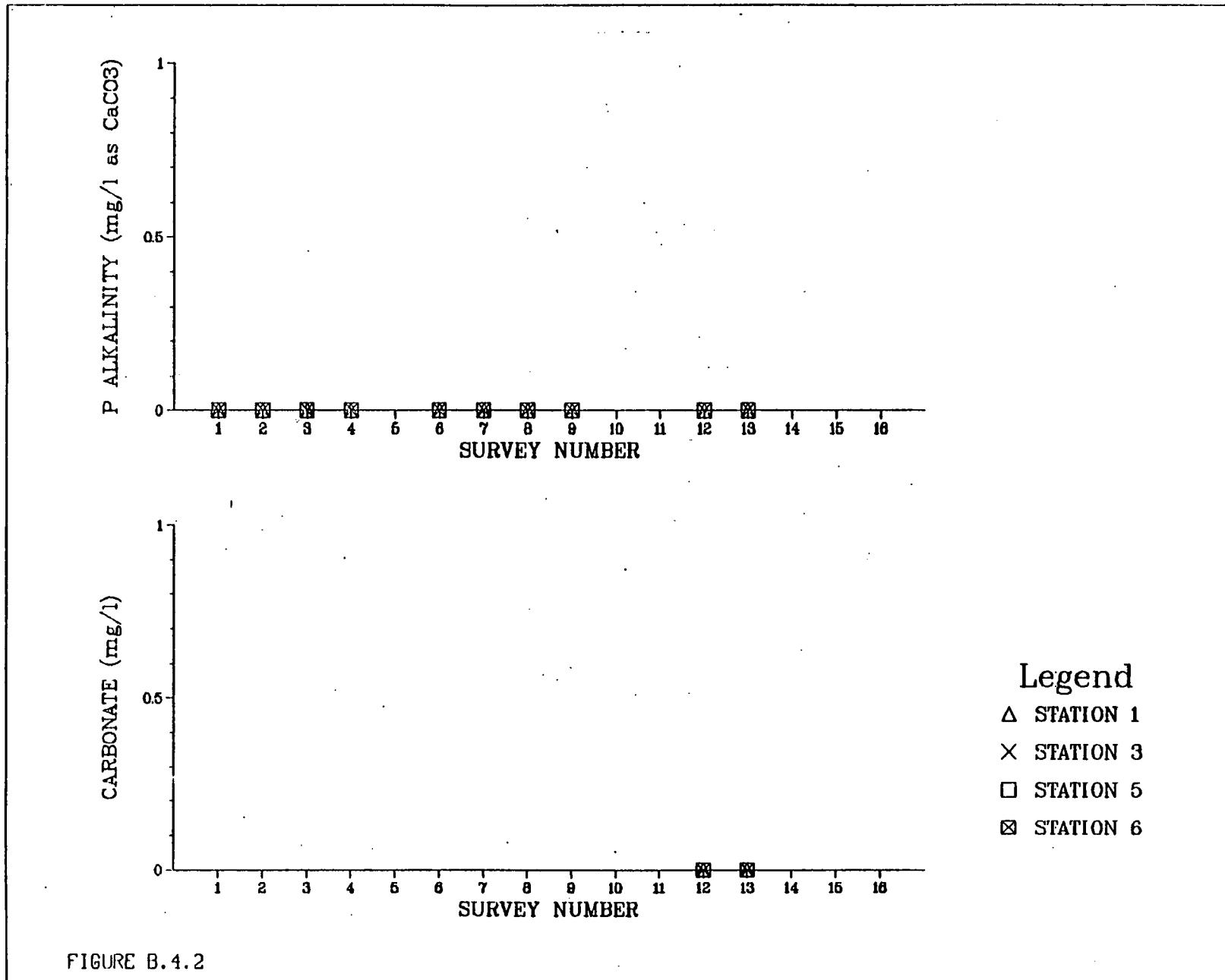


FIGURE B.4.2

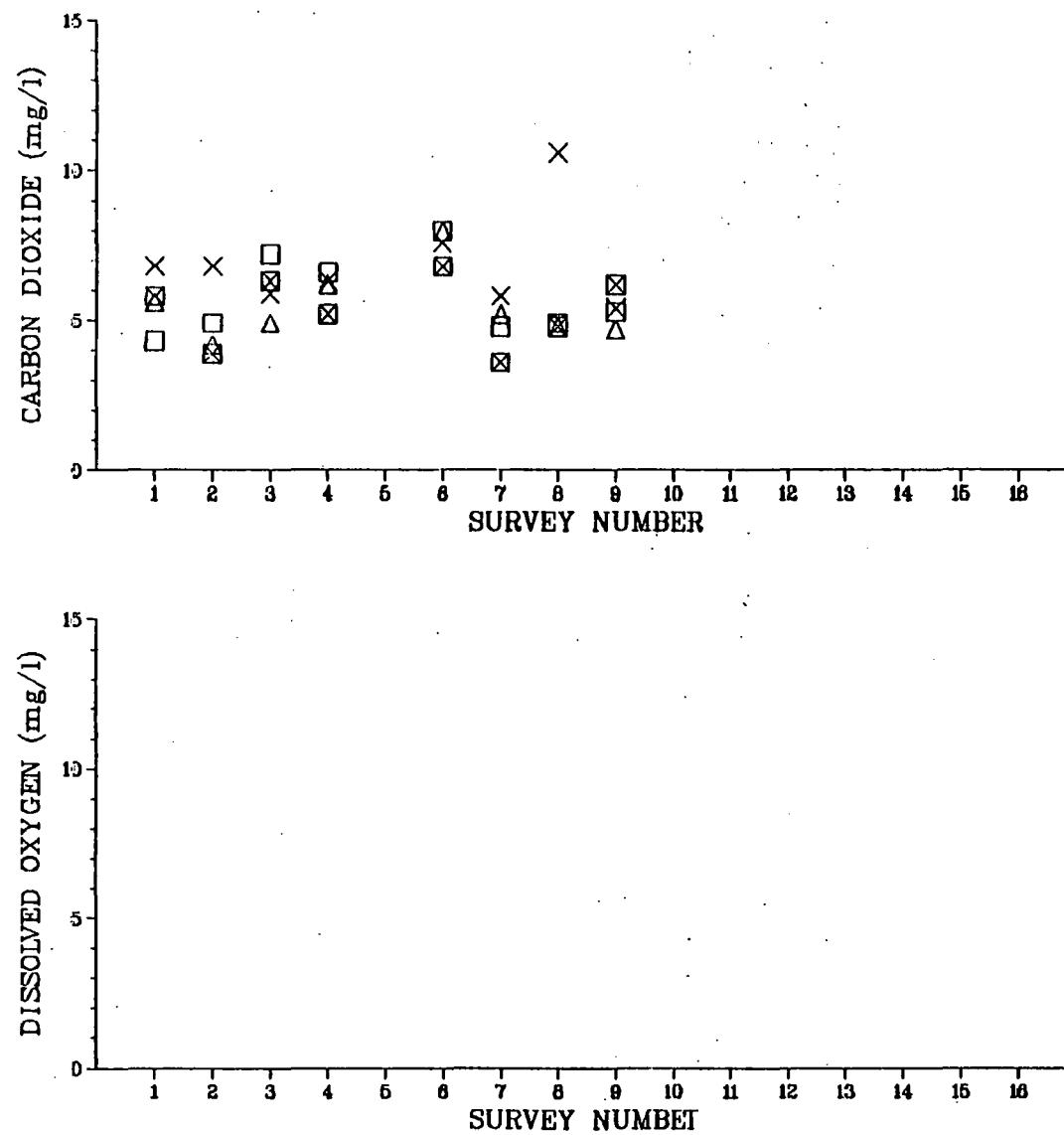


FIGURE B.4.3

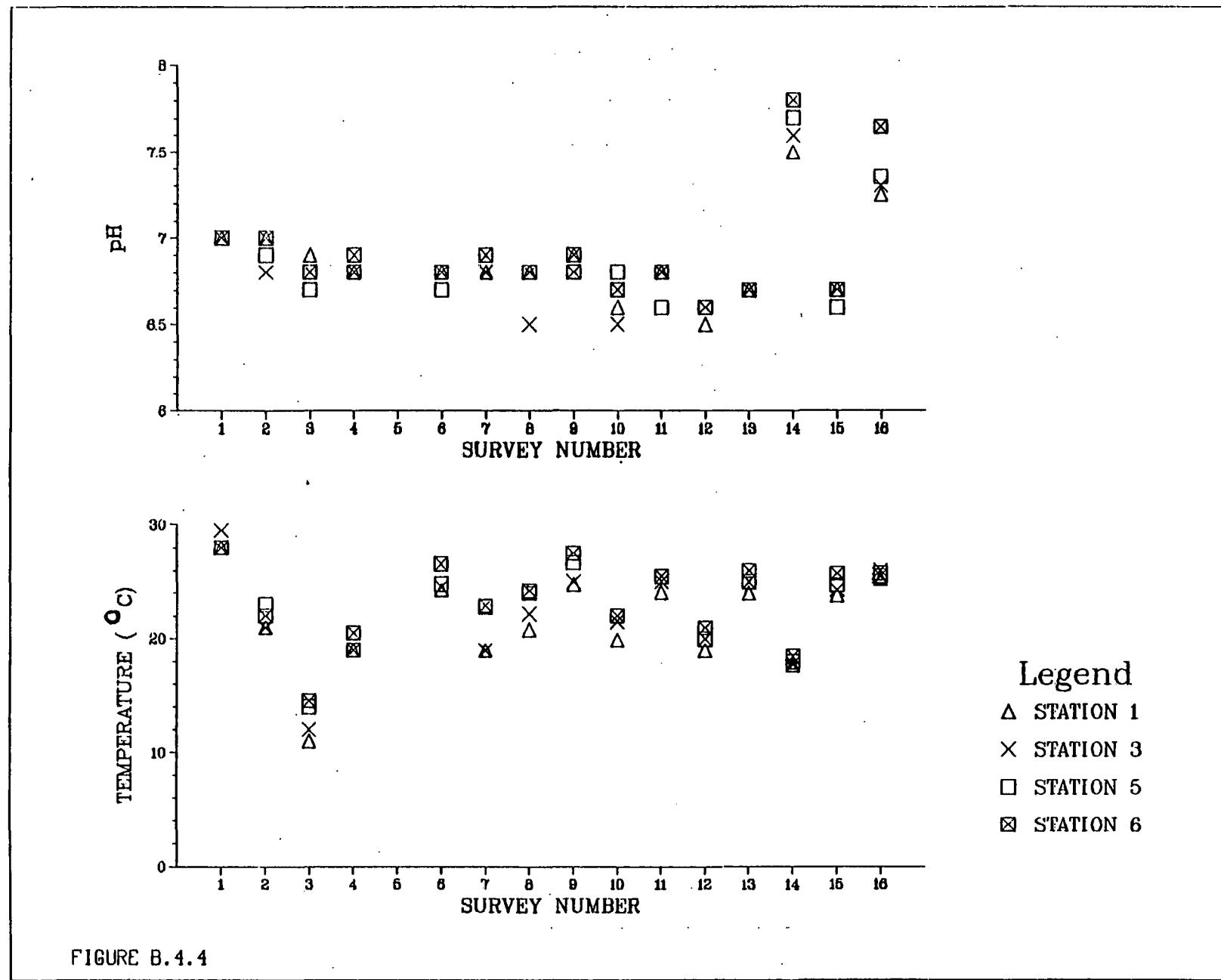


FIGURE B.4.4

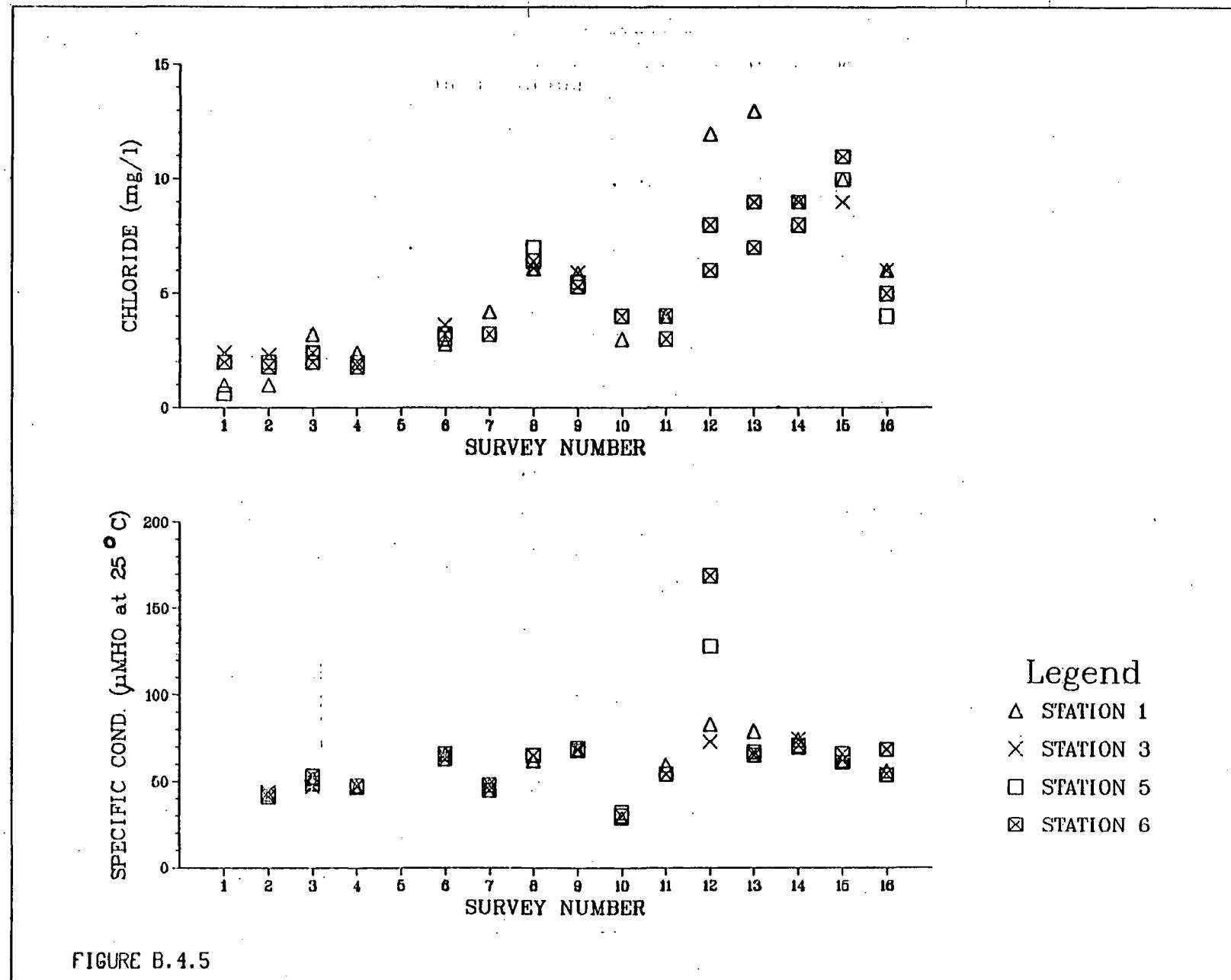


FIGURE B.4.5

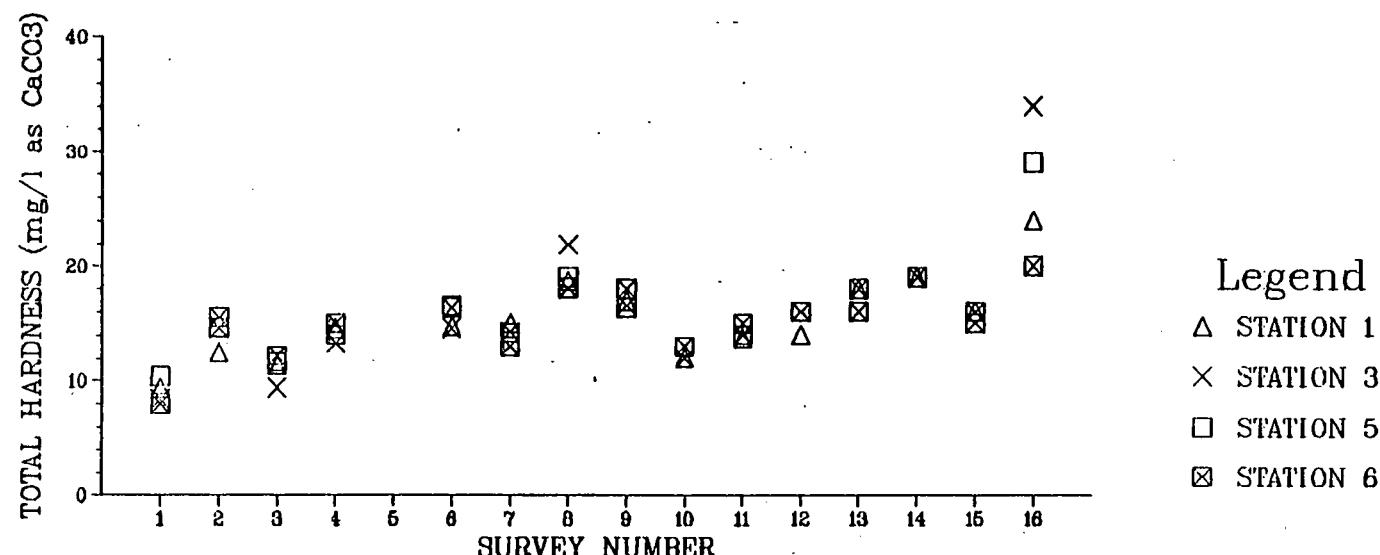


FIGURE B.4.6

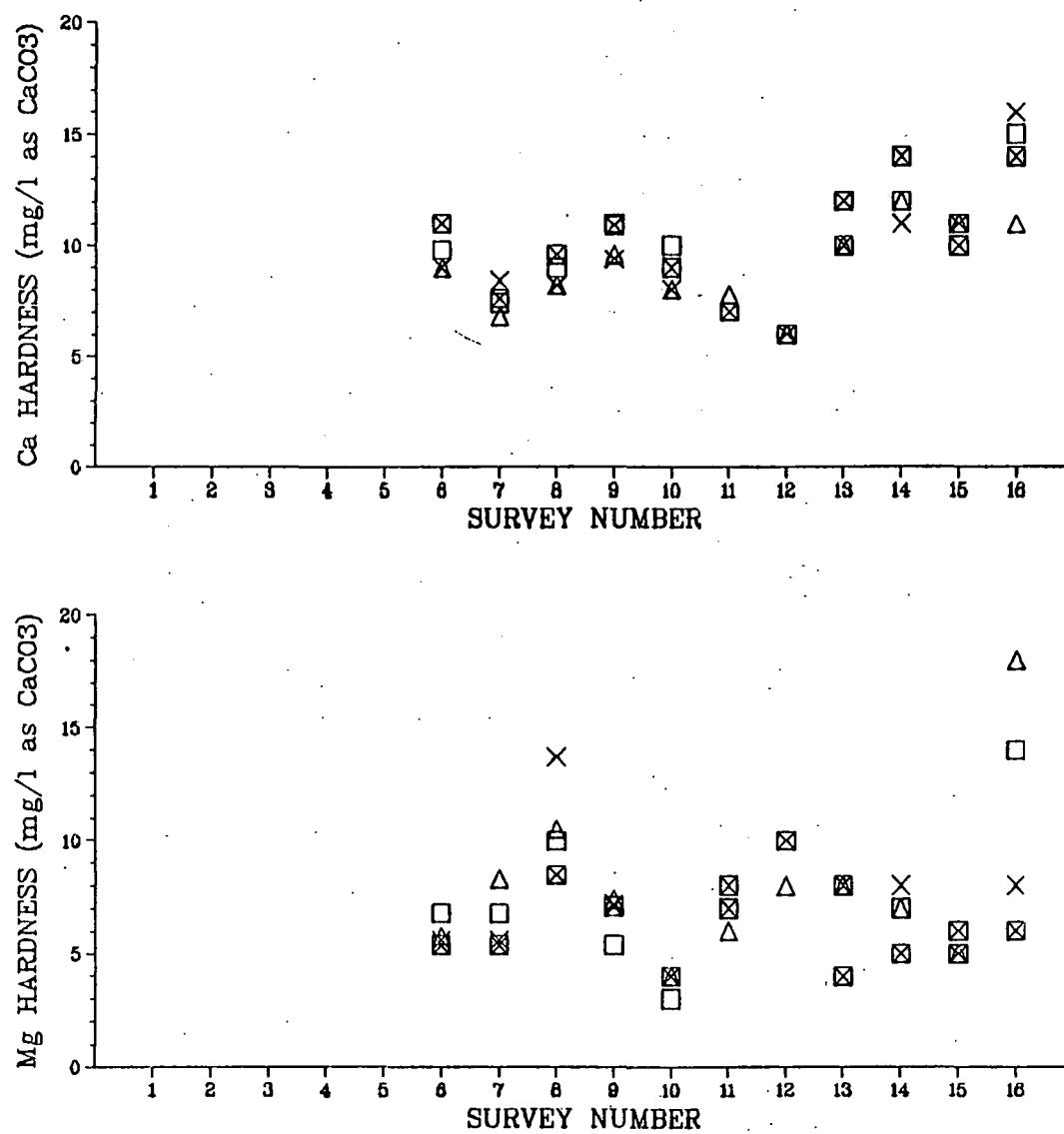


FIGURE B.4.7

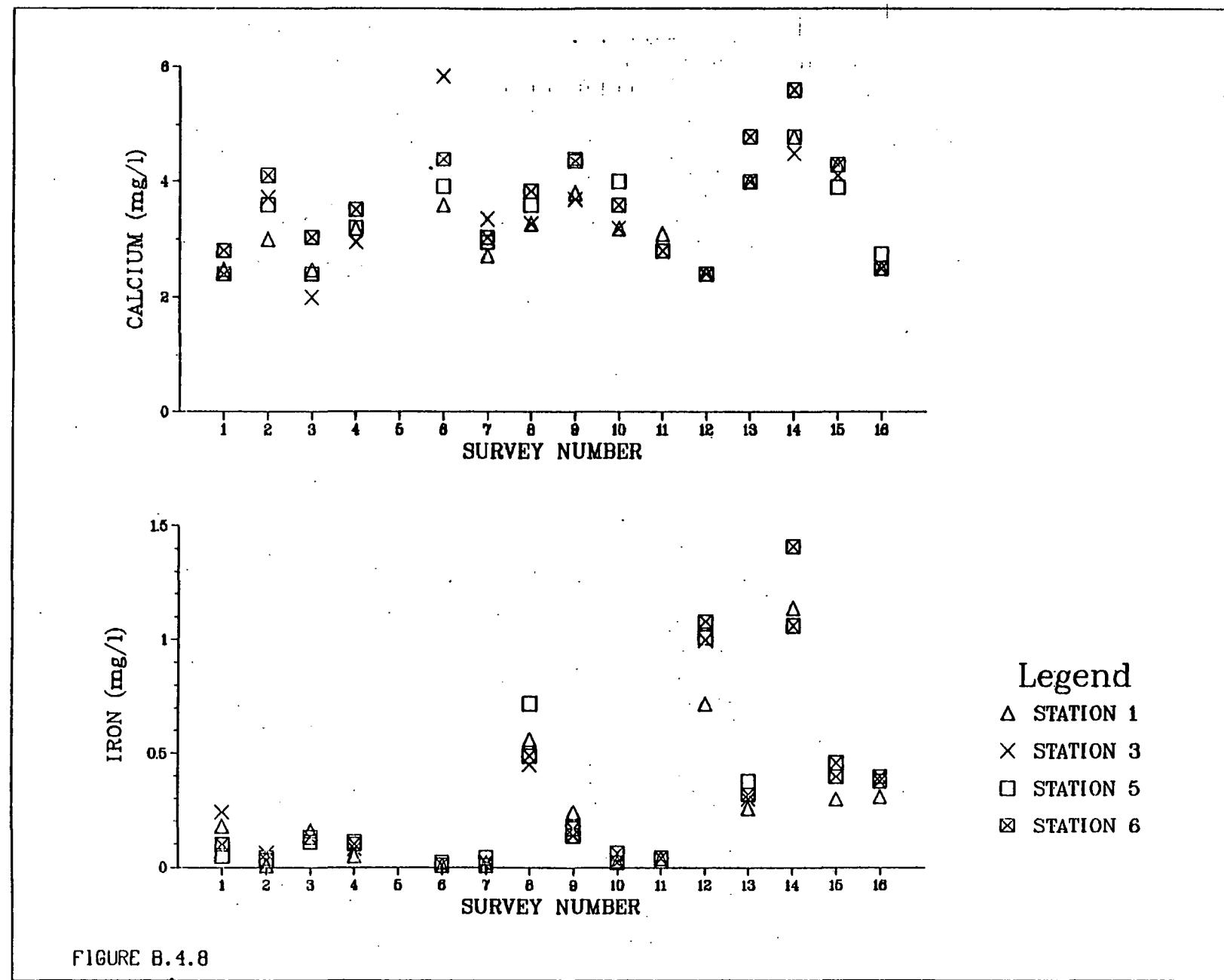


FIGURE B.4.8

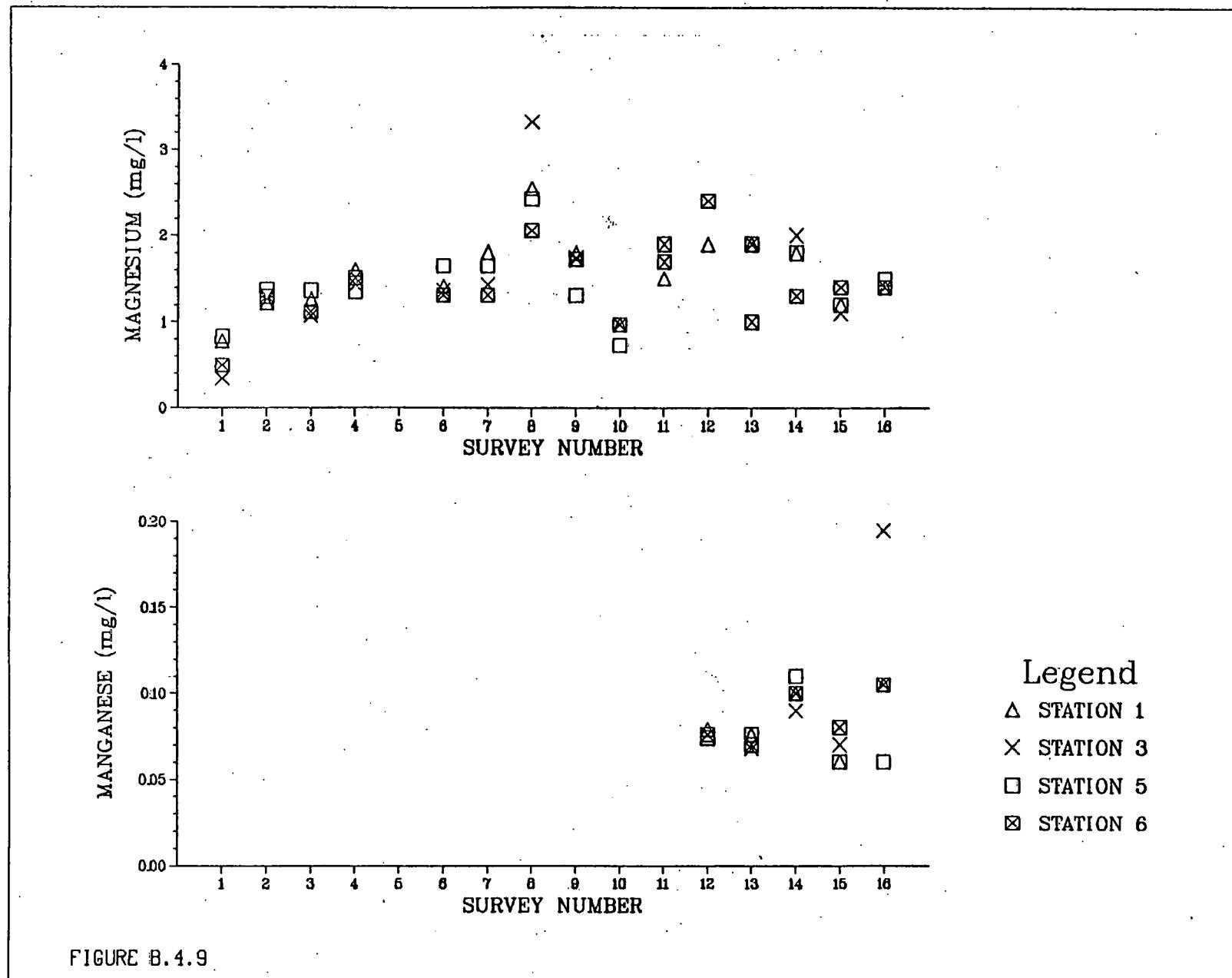


FIGURE B.4.9

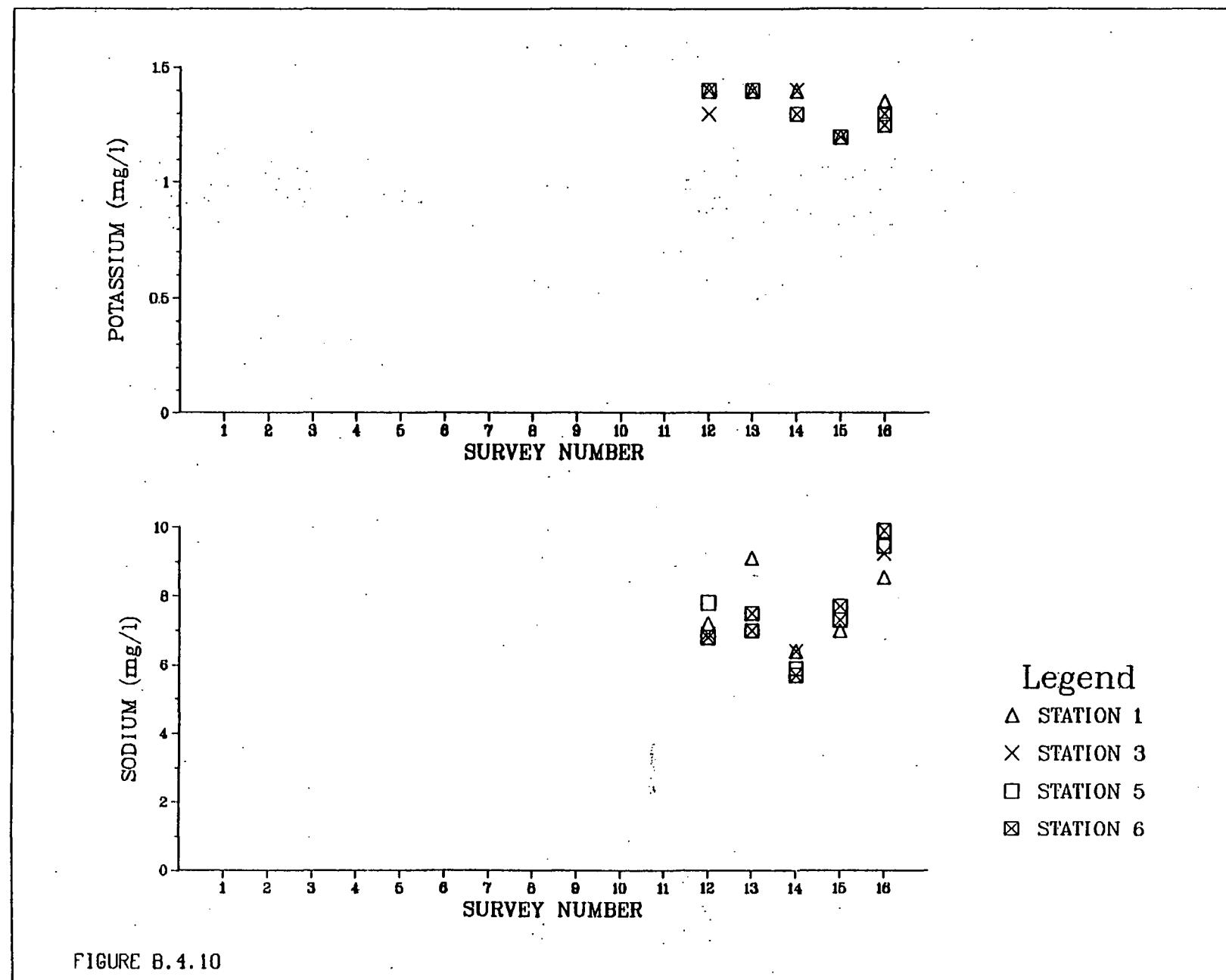


FIGURE B.4.10

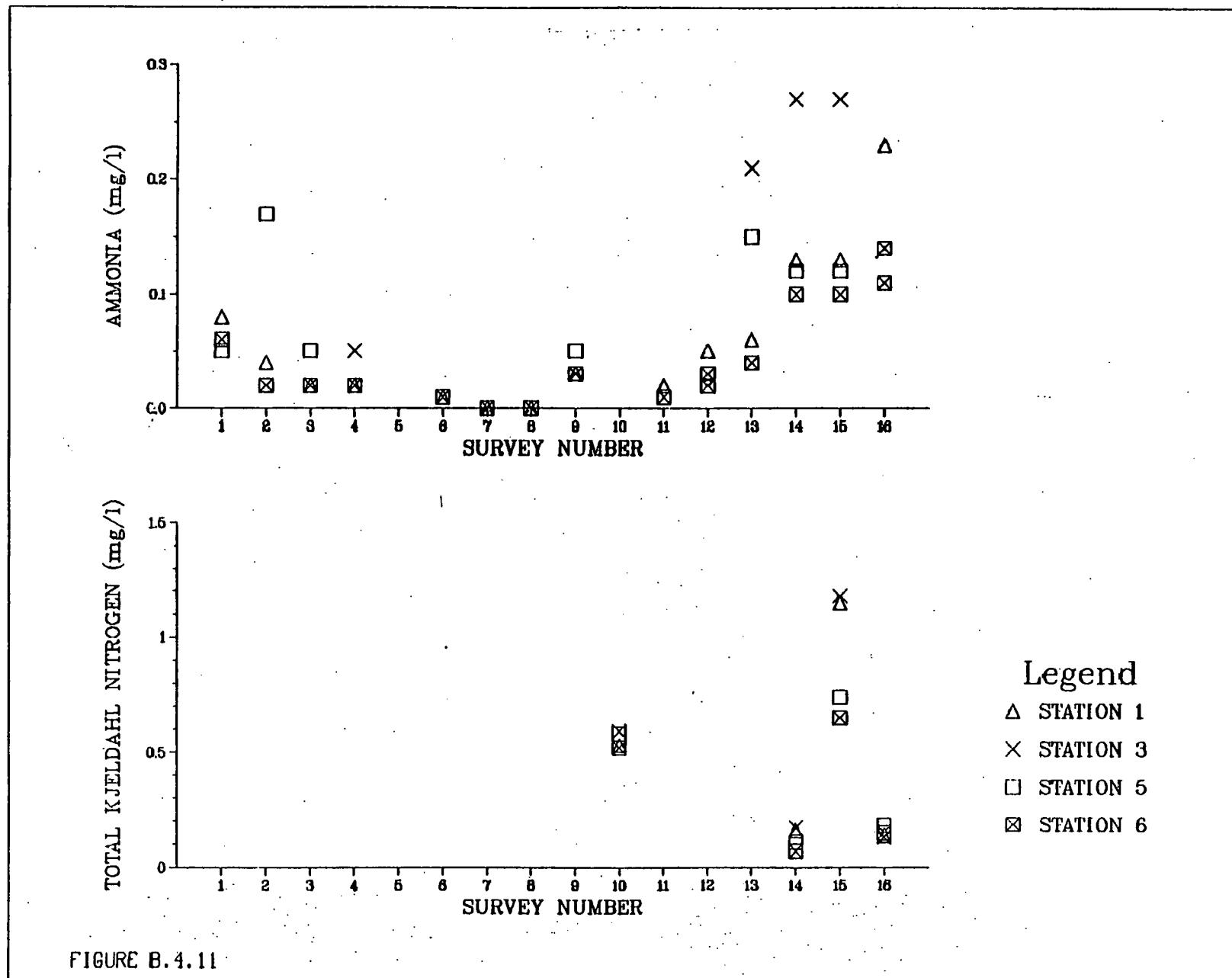


FIGURE B.4.11

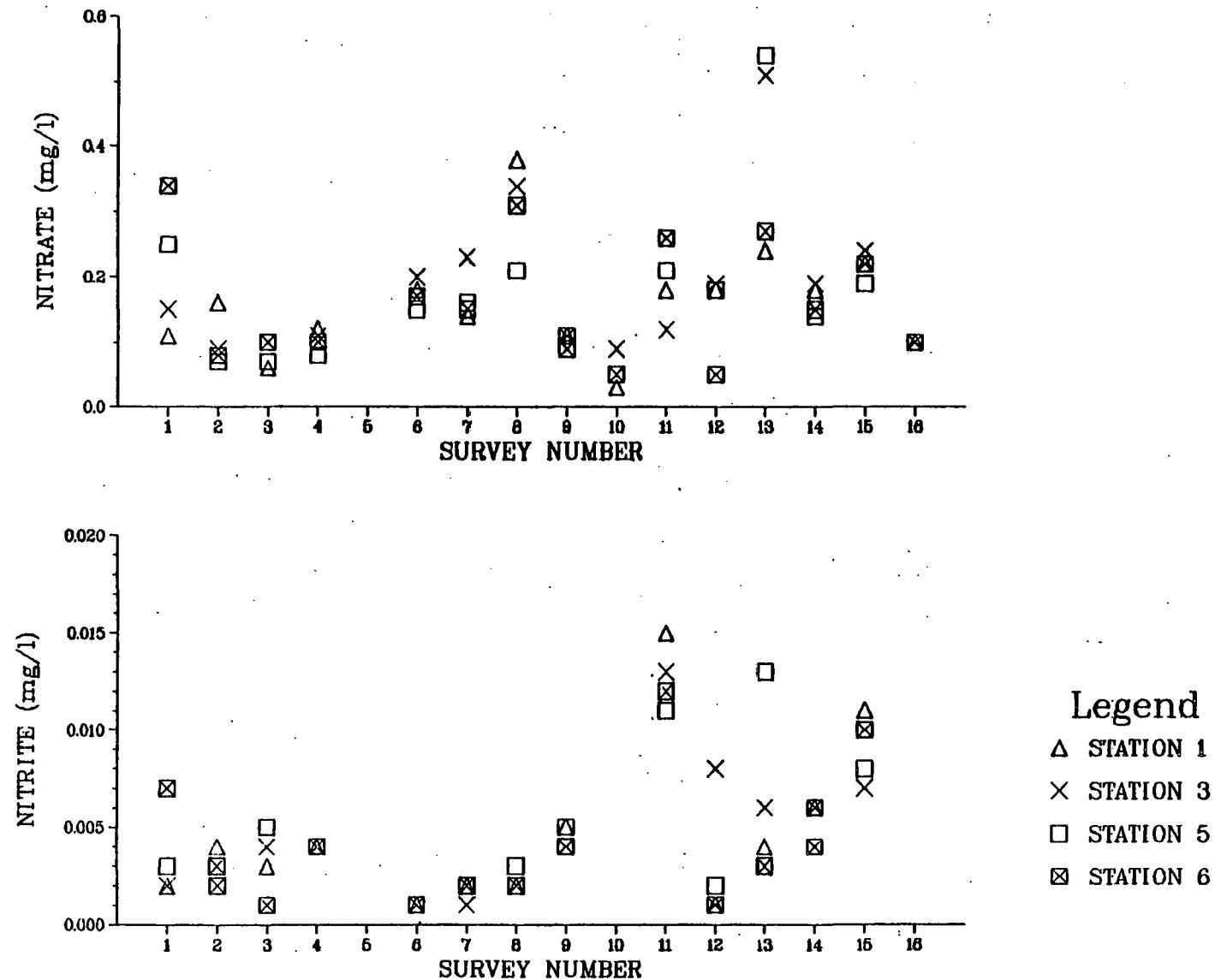


FIGURE B.4.12

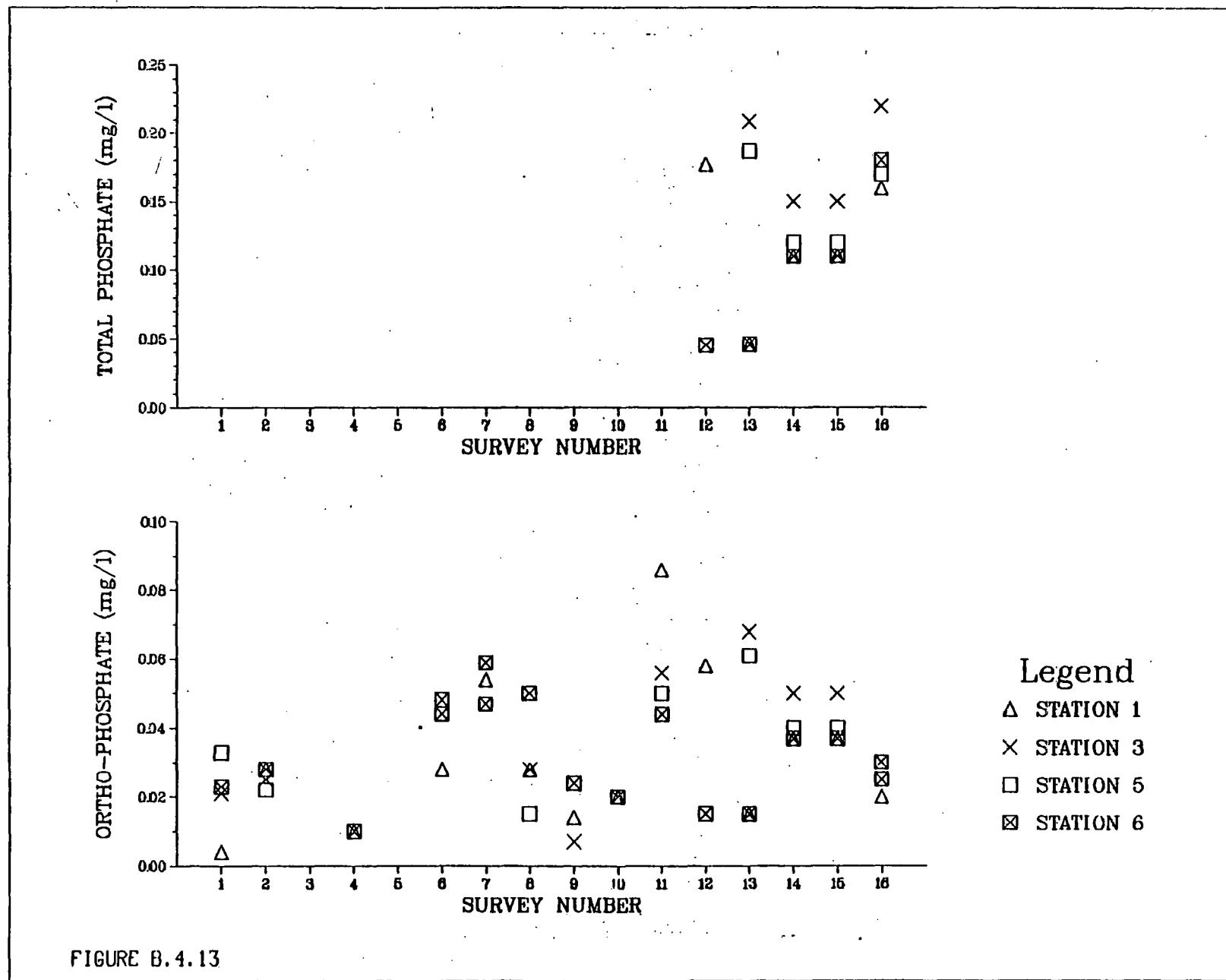


FIGURE B.4.13

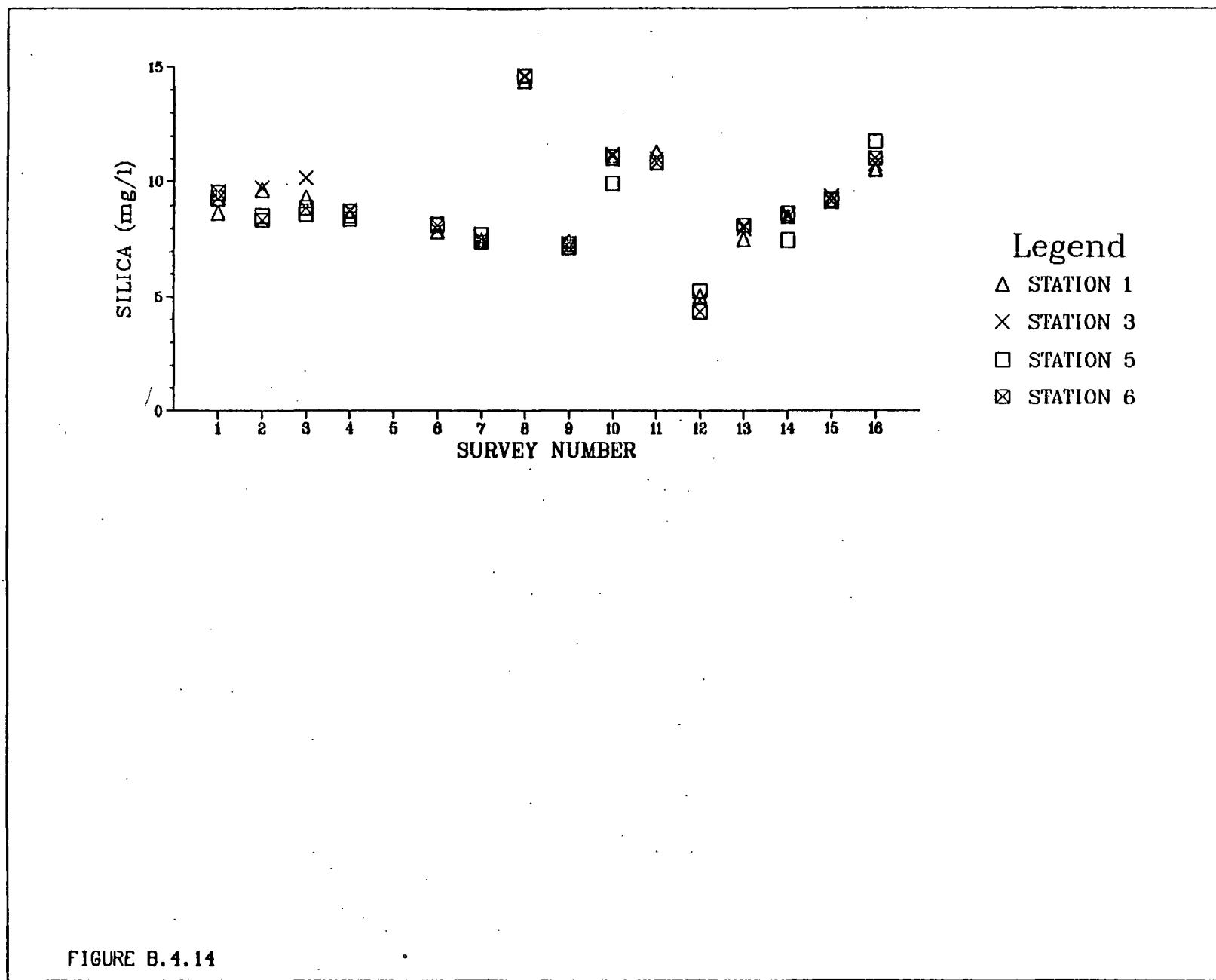


FIGURE B.4.14

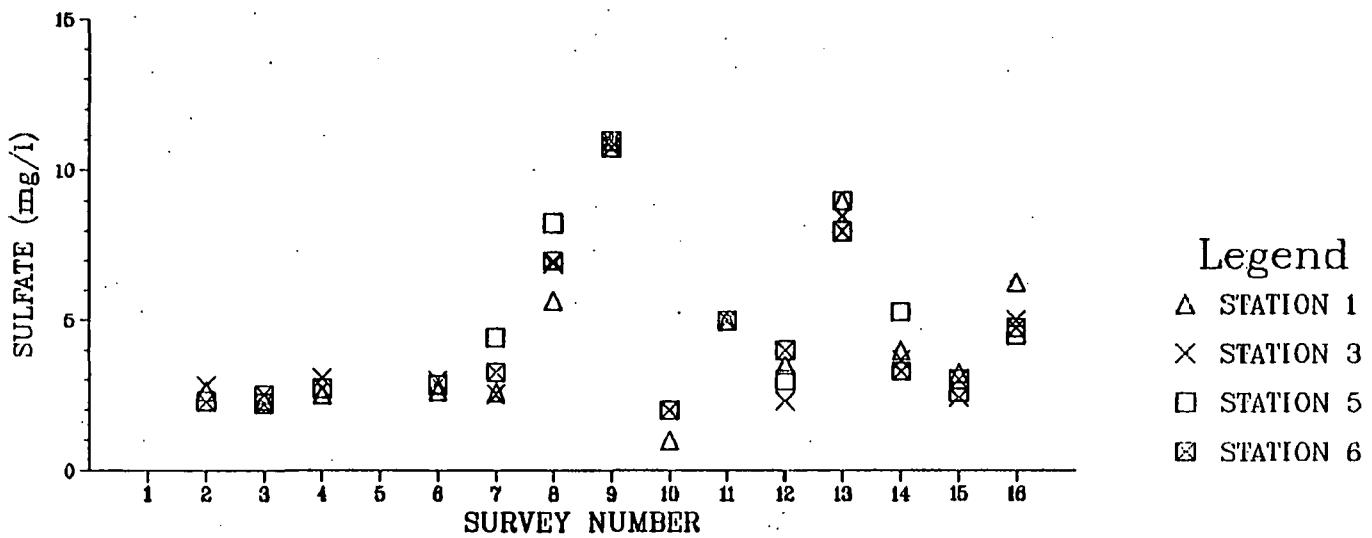


FIGURE B.4.15

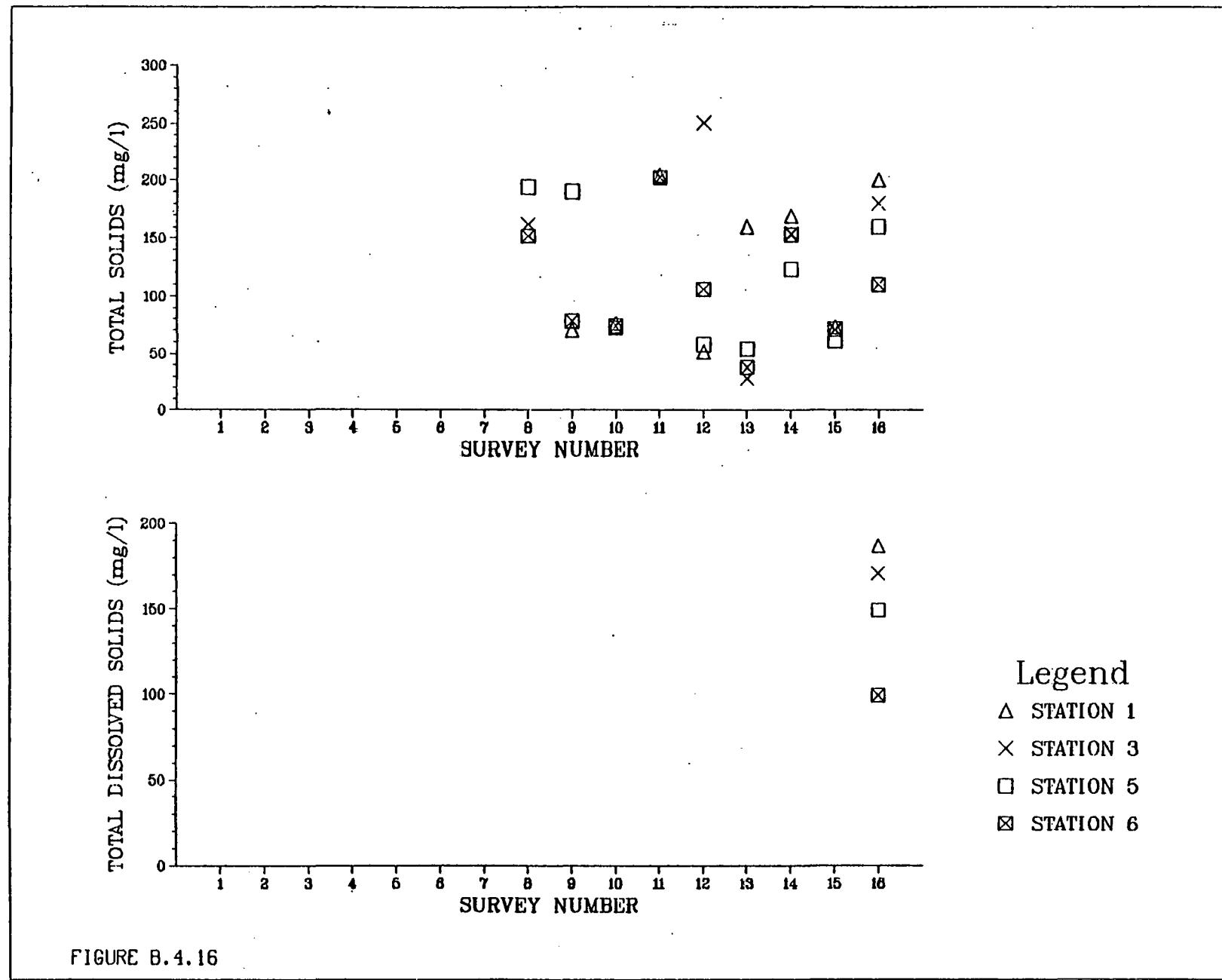


FIGURE B.4.16

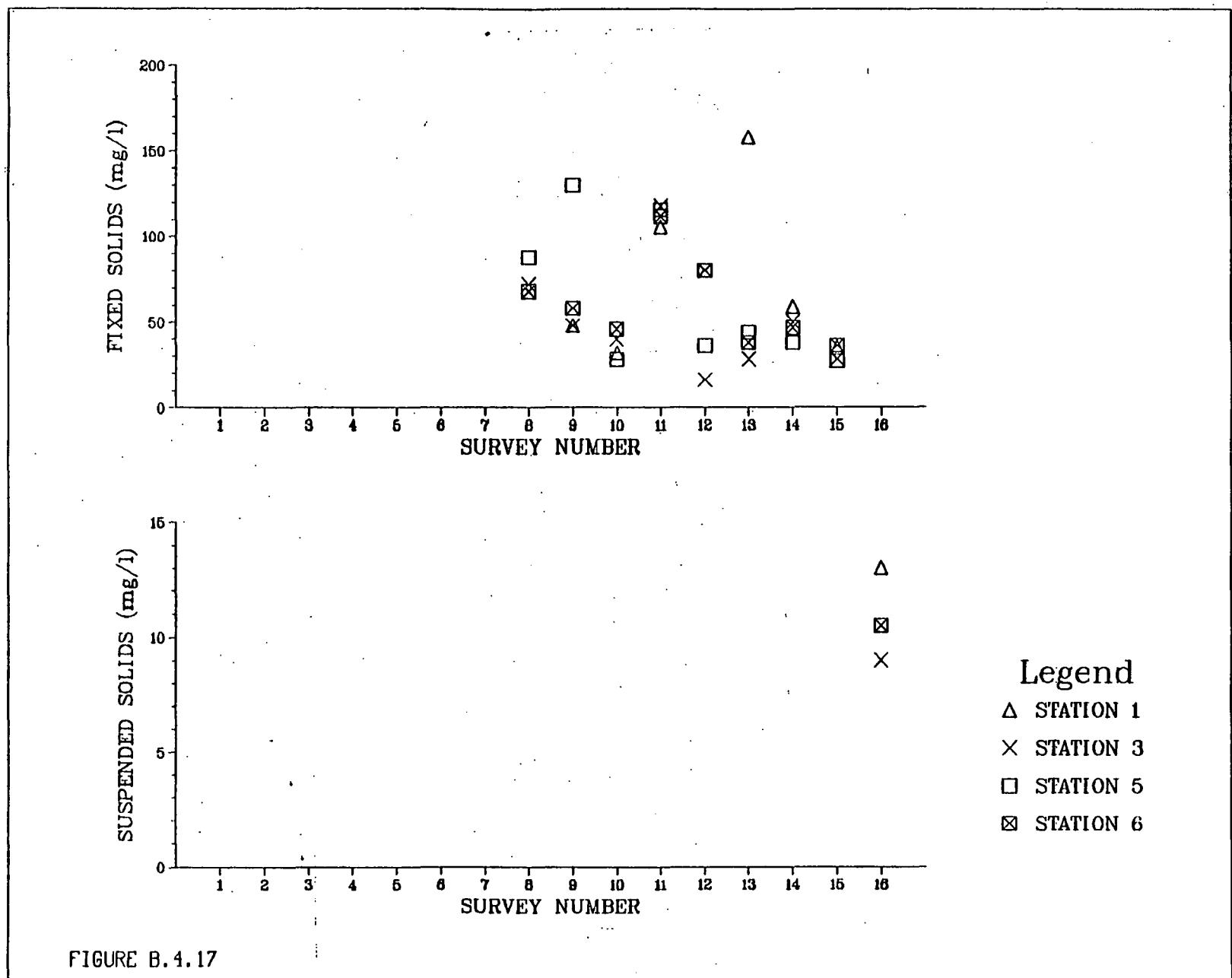


FIGURE B.4.17

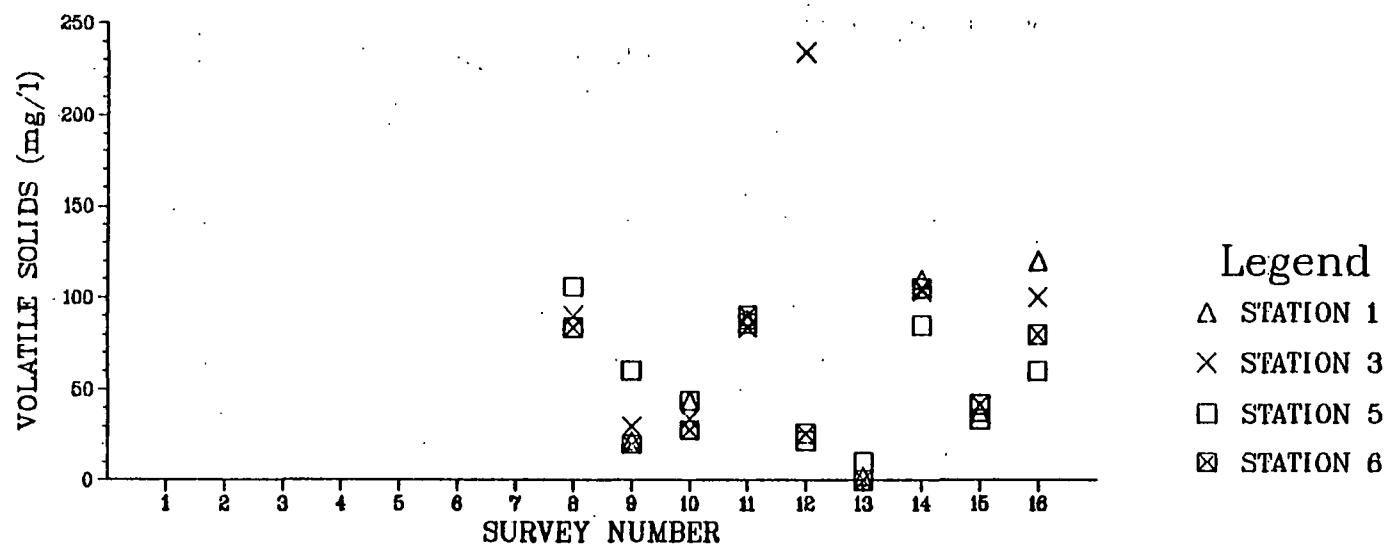


FIGURE B.4.18

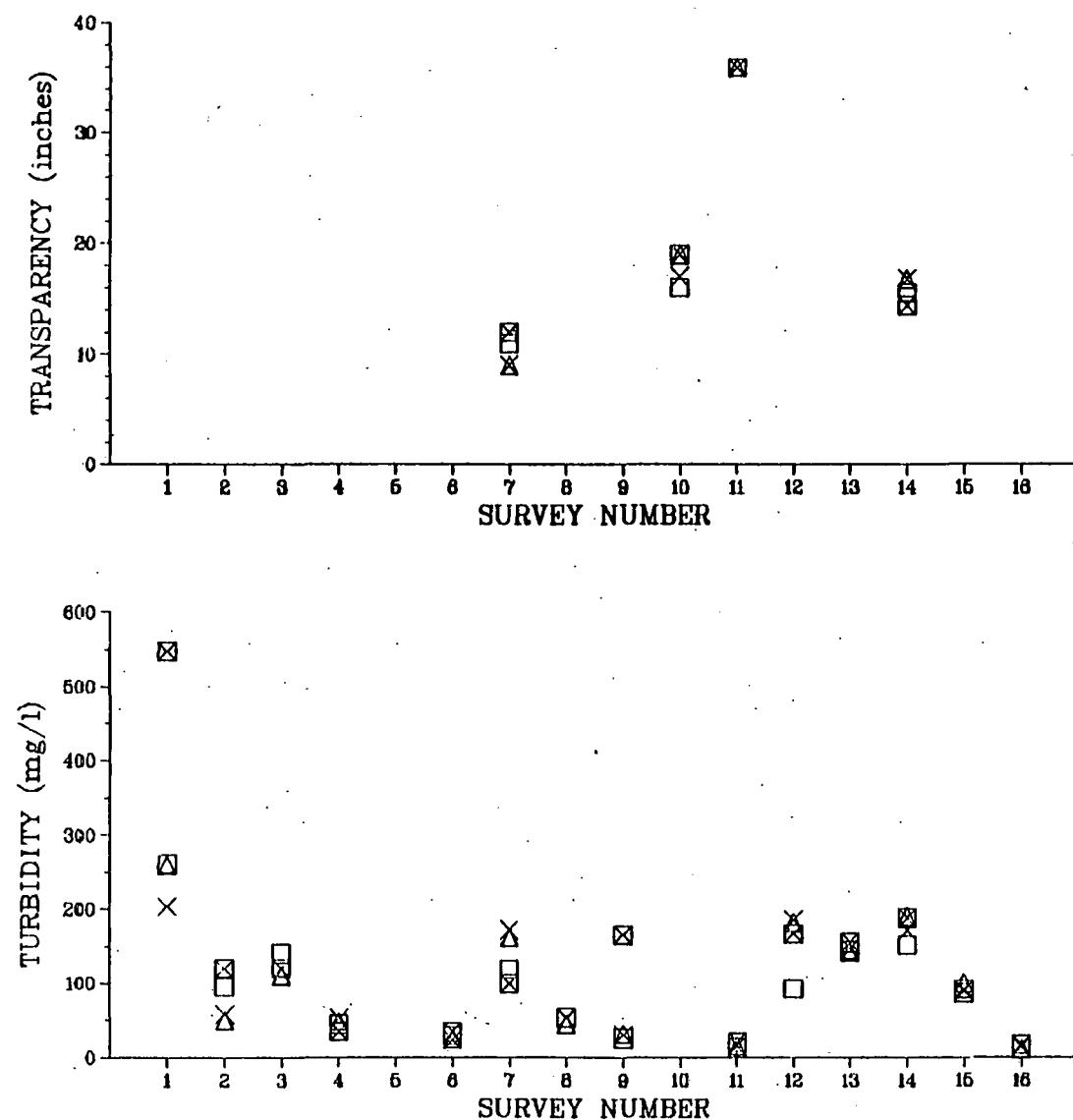


FIGURE B.4.19

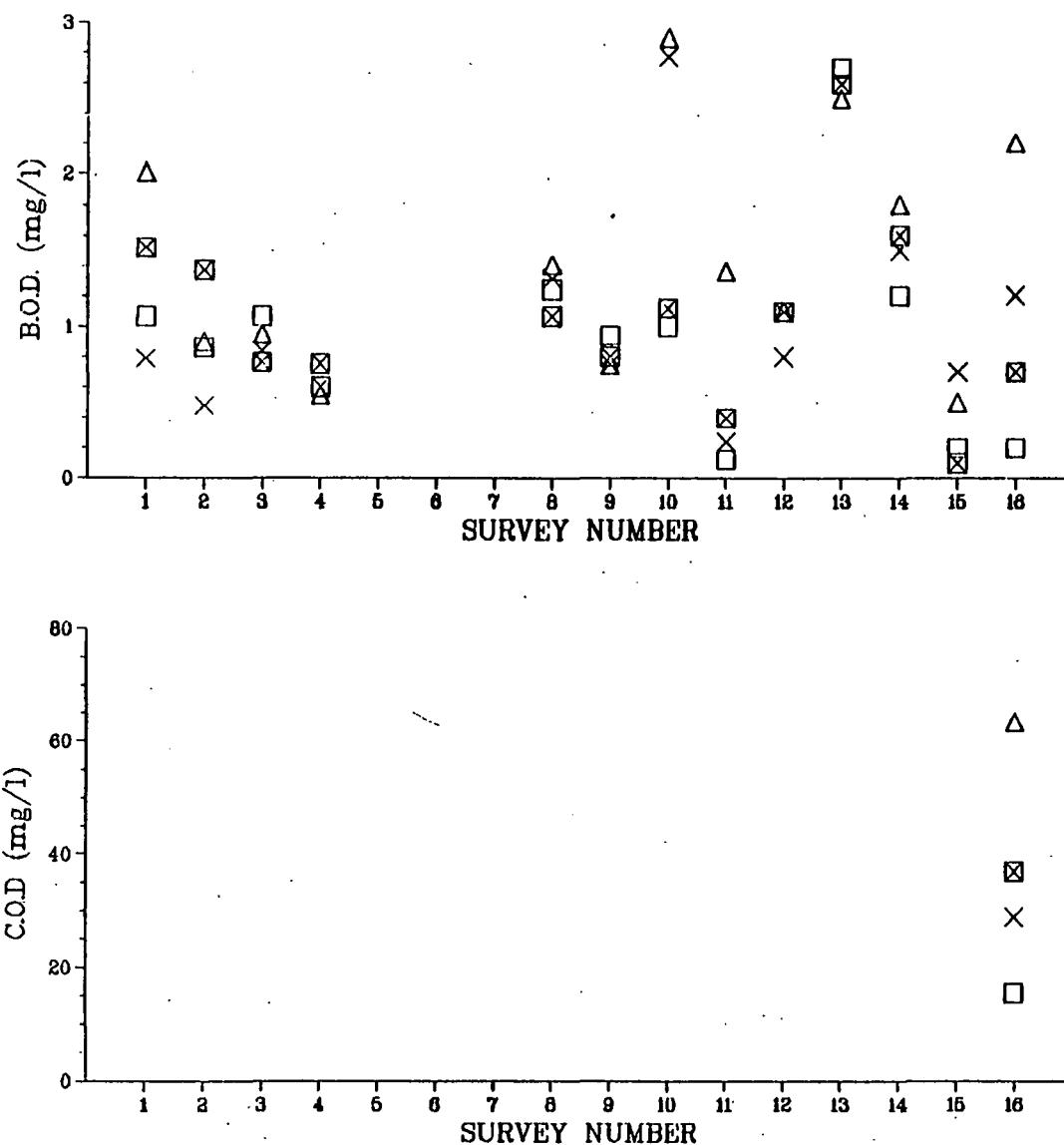


FIGURE B.4.20

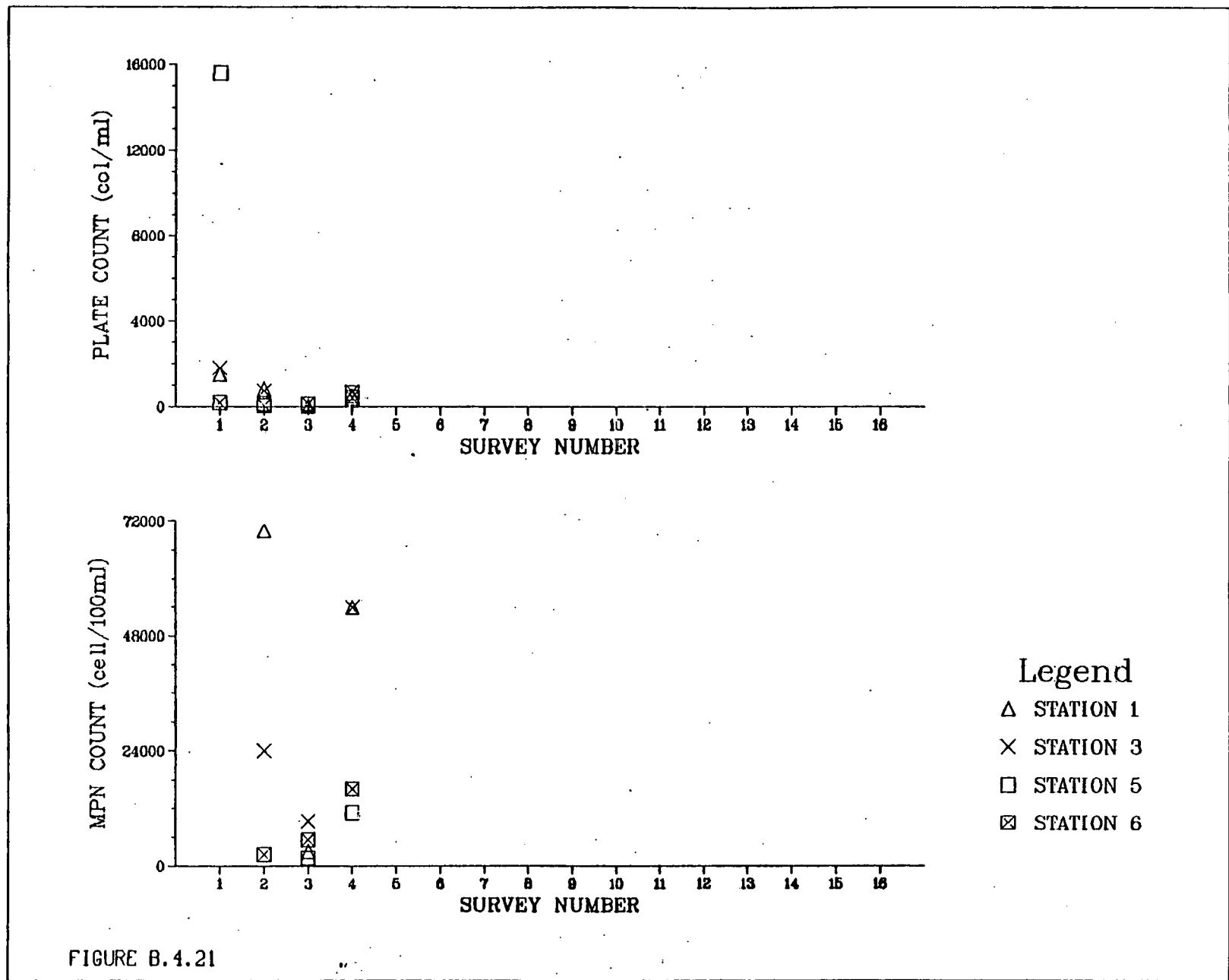
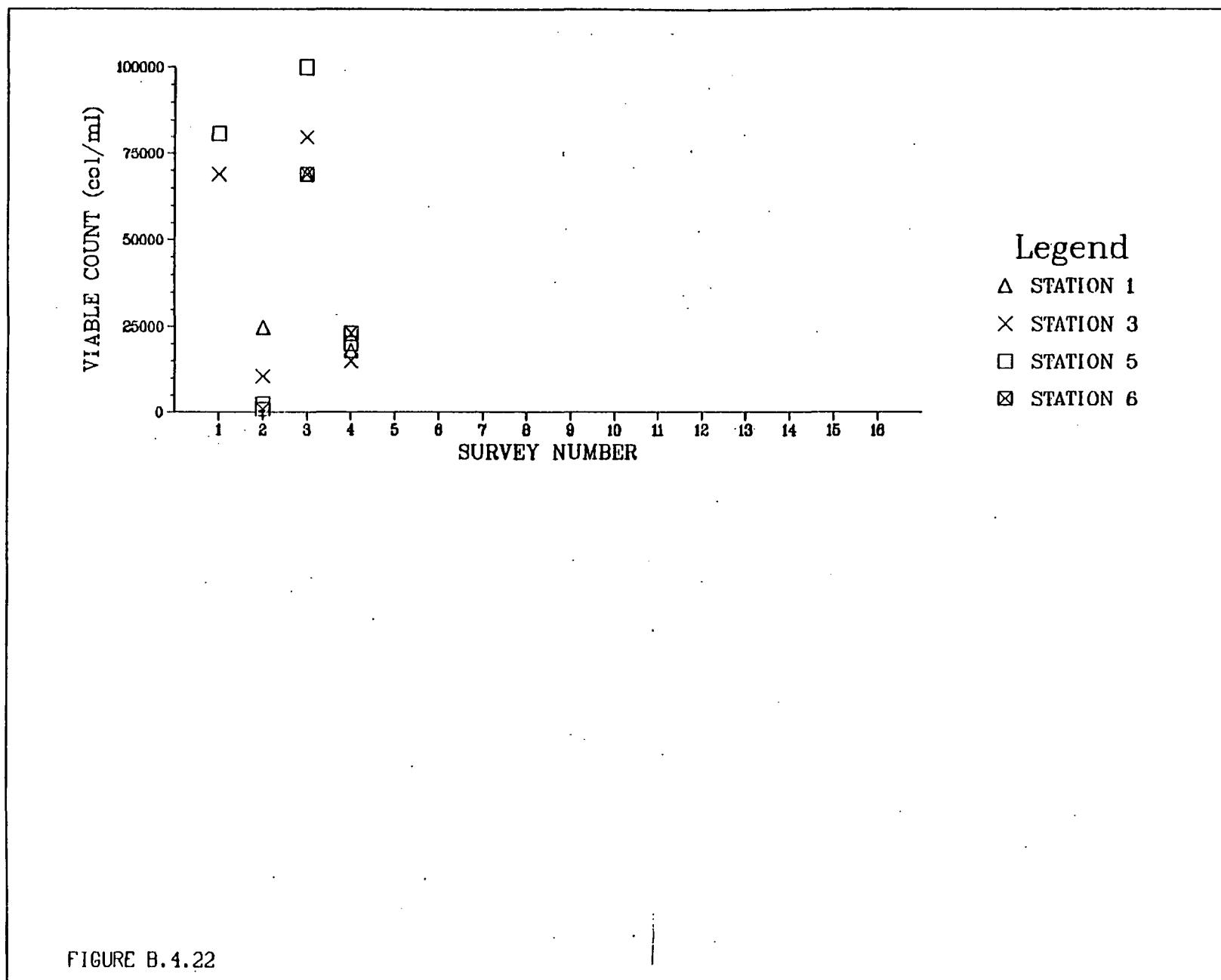
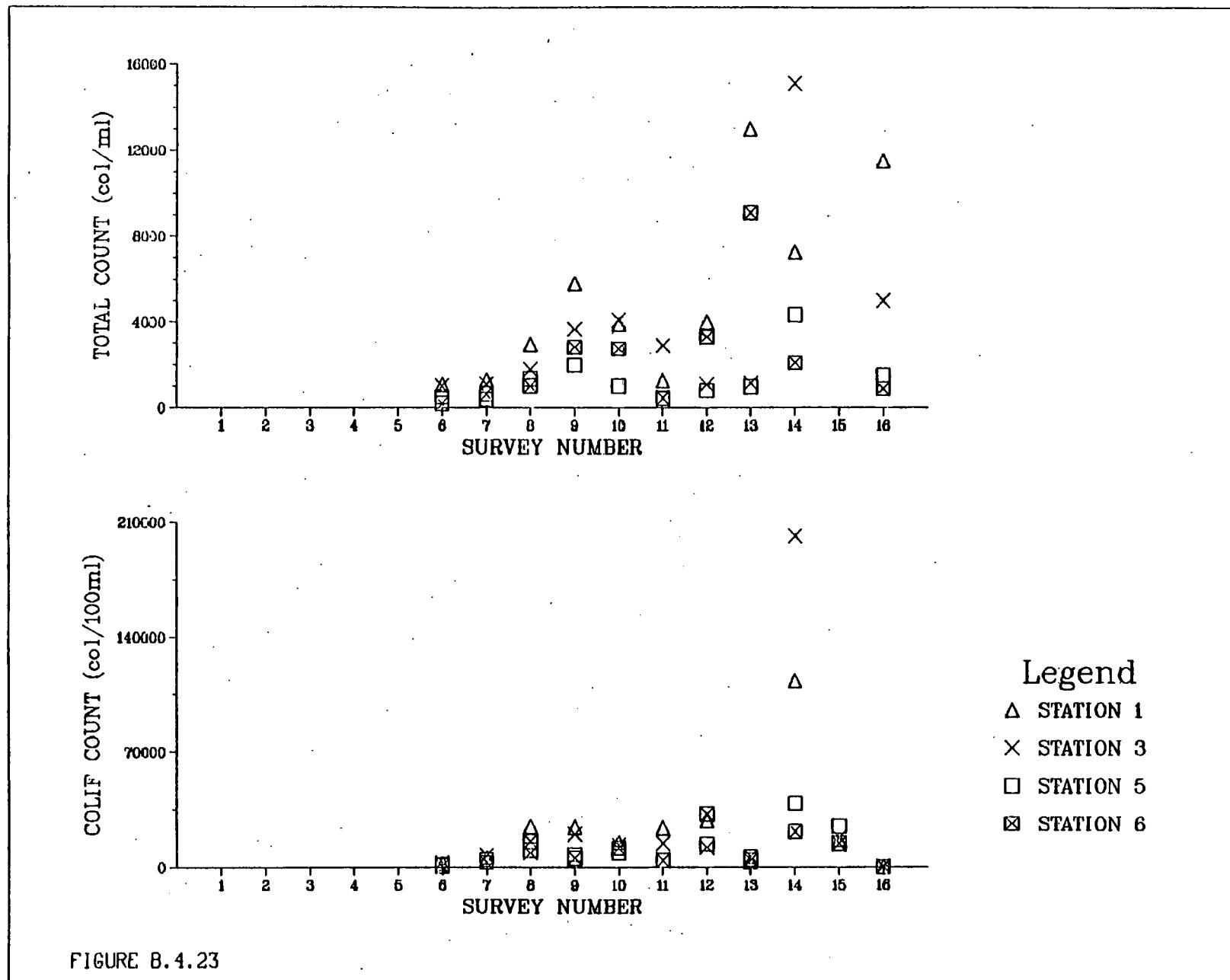


FIGURE B.4.21





Appendix C. Savannah River Flow and Temperature Data

- C.1 Savannah River Flow and Temperature Data at Augusta (A) or Jackson (J).
- C.2 Savannah River Daily Flow at Augusta during the Academy of Natural Sciences of Philadelphia Surveys.

Appendix C.1. Savannah River Flow and Temperature Data at Augusta (A) or Jackson (J)*

Year	Oct.	Nov.	Dec.	Jan.	Feb.	Mar.	Apr.	May	June	July	Aug.	Sep.	Water Year Ave.
1976-77													
flow (A)**	8880	9583	20530	13210	8924	13020	20180	8396	7745	7612	7235	6909	11030
temp (A)***	20.0	14.0	10.0	6.5	7.0	11/5	14/5	18.0	19.5	22.0	23.5	24.0	16.0
temp (J)	19.5	m	m	6.0	7.5	12.5	15.5	19.5	22.0	23.0	24.5	24.5	17.0
1975-76													
flow (A)	11100	13150	12680	12260	10420	16750	12600	11120	16950	13200	7380	7614	12110
temp (A)	21.0	18.0	13.0	9.0	10.0	12.5	16.0	17.5	m	21.5	23.5	22.5	17.0
temp (J)	21.0	17.5	12.5	9.0	11.0	13.5	16.5	18.5	20.5	22.5	24.0	23.0	17.5
1974-75													
flow (A)	6451	6814	7043	12171	18143	28493	21382	13425	9235	8230	7545	7882	12196
temp (A)	21.0	17.0	11.0	10.5	11.5	15.0	17.5	m	22.0	23.0	22.5	16.5	16.5
temp (J)	19.5	16.0	11.0	10.5	11.0	12.0	15.5	18.5	21.0	22.5	24.0	23.0	17.0
1973-1974													
flow (A)	6076	6153	7845	16160	22350	8762	13900	7865	7093	7302	8181	7238	9822
temp (A)	21.5	17.5	13.0	12.0	m	m	m	19.5	21.5	m	23.0	23.0	18.5
temp (J)	21.0	17.0	12.5	12.5	m	m	17.0	20.5	22.5	23.5	23.5	23.0	18.5
1972-1973													
flow (A)	6581	6386	12550	15260	19080	18180	25620	11030	22830	7906	7469	6345	13200
temp (J)	20.5	16.5	12.5	9.5	9.0	13.0	15.0	18.0	20.0	23.5	23.5	24.0	17.0
1971-1972													
flow (A)	7198	7504	14160	19260	16160	8569	7737	9347	10390	8429	7129	7078	10240
temp (J)	20.5	16.5	12.5	m	9.5	13.0	16.5	18.0	21.0	23.5	24.5	23.5	17.5
1970-1971													
flow (A)	6460	6283	6536	7152	7314	21580	8658	9374	7339	7248	8471	7306	8668
temp (J)	m	m	m	m	m	m	m	m	m	m	m	m	m
1969-1970													
flow (A)	6602	6587	6867	6945	7093	8552	8093	6582	6548	7059	6889	6562	7032
temp (J)	20.6	15.0	11.5	8.5	9.7	12.8	16.6	19.7	m	m	m	m	m

* Augusta measurements are taken 0.2 mi (0.3 km) upstream from Butler Creek at mile 187.4.
 Jackson measurements are taken 1.4 mi (2.3 km) downstream from Upper Three Runs Creek at mile 156.8. References 19, 30.

** Flow values are given in cfs.

*** Temperature values are given in °C.

Appendix C.1. (cont)

Year	Oct.	Nov.	Dec.	Jan.	Feb.	Mar.	Apr.	May	June	July	Aug.	Sep.	Water Year Ave.
1968-1969													
flow (A)	6649	6834	6469	10210	13590	11350	20800	13680	7370	6942	7128	7177	9812
temp (J)	19.7	15.0	10.2	7.8	7.9	9.8	15.2	19.3	21.9	23.2	22.3	21.9	
1967-1968													
flow (A)	6286	7593	15090	18440	7175	7199	7555	7620	9607	7366	7500	6809	9043
temp (J)	19.9	15.6	12.7	8.0	8.2	11.2	15.1	17.3	20.0	22.7	23.4	21.7	
1966-1967													
flow (A)	6478	6478	6795	8718	8439	9228	6870	7036	14440	8713	8625	8740	8372
temp (J)	20.3	17.0	12.2	9.5	9.7	13.1	16.6	18.4	18.7	22.2	22.9	21.7	
1965-1966													
flow (A)	7057	6958	7060	8783	13610	23610	7201	10480	9031	6830	6731	6896	9509
temp (J)	20.2	17.8	13.7	9.3	8.3	10.3	15.5	17.2	20.0	22.9	22.9	21.9	
1964-1965													
flow (A)	17740	10950	17670	17610	10120	17450	16370	9574	12760	7652	9027	7959	12940
temp (J)	18.5	16.4	12.5	9.8	9.6	10.5	14.4	18.4	19.3	21.8	22.0	22.1	
1963-1964													
flow (A)	6559	6826	9818	16360	16720	27510	43850	27050	7143	10970	11900	14480	16580
temp (J)	20.1	16.4	9.6	7.1	7.3	12.0	14.1	17.7	22.5	21.9	22.6	22.3	
1962-1963													
flow (A)	5960	5852	5865	9178	9885	18460	7675	14900	10090	11220	7875	7488	9554
temp (J)	20.0	15.1	10.5	8.4	7.5	11.0	16.7	17.5	20.8	22.4	23.6	21.8	
1961-1962													
flow (A)	5680	5537	12700	14960	9978	13180	15420	7963	8190	5676	5992	6050	9276
temp (J)	19.6	17.8	12.7	8.6	10.2	10.7	14.1	20.2	21.5	23.8	23.7	22.5	
1960-1961													
flow (A)	6514	5867	5943	6198	9951	11980	21770	7425	6783	8840	7700	7835	8873
temp (J)	22.0	16.5	10.3	8.4	9.3	12.7	14.4	19.5	22.1	22.5	22.9	22.9	

* Augusta measurements are taken 0.2 mi (0.3 km) upstream from Butler Creek at mile 187.4.
 Jackson measurements are taken 1.4 mi (2.3 km) downstream from Upper Three Runs Creek at mile 156.8. References 19, 30.

** Flow values are given in cfs.

*** Temperature values are given in °C.

Appendix C.1. (cont)

Year	Oct.	Nov.	Dec.	Jan.	Feb.	Mar.	Apr.	May	June	July	Aug.	Sep.	Water Year Ave.
1959-1960													
flow (A)	12680	12400	9298	15070	23480	18660	18020	9029	6267	6414	7344	6538	12450
temp (J)	22.3	16.3	11.7	9.9	9.0	8.6	15.0	18.2	22.1	23.5	24.2	24.7	
1958-1959													
flow (A)	5722	5675	5763	6026	7395	7322	6721	5781	13060	6613	7070	8559	7125
temp (J)	21.1	17.4	11.1	9.6	11.2	12.8	17.7	20.9	21.8	25.3	26.3	24.9	
1957-58													
flow (A)	7347	9310	13130	12780	14030	15120	23520	12260	6609	8796	7835	5879	11360
temp (J)	19.2	15.8	11.2	8.2	6.8	9.4	13.6	18.1	21.8	23.5	24.9	25.4	
1956-1957													
flow (A)	5437	5622	5962	6105	5988	6772	8225	9802	6138	6143	6171	6452	
temp (J)	19.5	15.5	12.5	10.2	12.6	13.0	17.6	19.5	22.6	24.2	24.9	25.0	
1955-1956													
flow (A)	4976	5076	5150	4418	4861	5668	6171	6031	5425	5266	5547	6189	5398
temp (J)	21.4	15.7	11.2	8.8	11.7	13.5	16.2	19.9	21.9	23.9	24.1	22.7	
1954-1955													
flow (A)	5818	5846	4982	4600	5278	5767	7119	5804	5227	5206	5226	4995	5487
temp (J)	data not available												
1953-1954													
flow (A)	7172	6498	7115	7247	7269	9420	11460	7306	6575	6230	5677	5584	7293
1952-1953													
flow (A)	3385	4017	3751	4084	5889	11390	6460	15150	5778	5750	5696	7232	6561
1951-1952													
flow (A)	2728	4196	8662	8654	5842	29090	21810	6782	4342	3627	3889	3332	8596
1950-1951													
flow (A)	8541	5166	8319	6581	7451	9764	11440	6580	5963	4570	3465	3389	6766

* Augusta measurements are taken 0.2 mi (0.3 km) upstream from Butler Creek at mile 187.4. Jackson measurements are taken 1.4 mi (2.3 km) downstream from Upper Three Runs Creek at mile 156.8. References 19, 30.

** Flow values are given in cfs.

*** Temperature values are given in °C.

Appendix C.2. Savannah River Daily Flow at Augusta during the Academy of Natural Sciences of Philadelphia Surveys*

<u>Survey</u>	<u>Date</u>	Flow (cfs)	<u>Survey</u>	<u>Date</u>	Flow (cfs)
1. 1951:	Jun. 25	6430	3. 1952:	Jan. 9	12800
	26	6830		10	11600
	27	6230		11	10500
	28	5640		12	9990
	29	5360		13	9510
	30	5080		14	11400
	Jul. 1	5260		15	13200
	2	5840		16	8320
	3	8080		17	4980
	4	6030		18	4540
	5	5550		19	5170
	6	5640		20	4360
	7	5460		21	5410
	8	5360		22	5040
	9	5460		23	5030
	10	4900		24	6930
	11	4620		25	5360
	12	4100		26	5080
	13	3580		27	4980
	14	3580		28	5260
				29	6730
2. 1951:	Oct. 15	2360		30	6730
	16	2280		31	5740
	17	1960			
	18	1770			
	19	3250			
	20	2170			
	21	1770			
	22	1830			
	23	1890			
	24	1960			
	25	2150			
	26	2860			
	27	3420			
	28	3670			
	29	3840			
	30	3260			
	31	2860			

* Reference 19

Appendix C.2. (cont)

Discharge at Augusta (cfs)

Survey	Date	Discharge	Survey	Date	Discharge	Survey	Date	Dis-	charge
4. 1952: May	5	9990	6. 1955: Aug	24	5170	8. 1960: May	23	5570	
	6	9990		25	5080		24	6010	
	7	9270		26	4900		25	6890	
	8	9270		27	4900		26	6010	
	9	9870		28	4900		27	6890	
	10	8030		29	5260		28	5900	
	11	7590		30	5170		29	5790	
	12	8360		31	4980		30	5790	
	13	7480		Sept 1	4900		31	6120	
	14	7590		2	4980		June 1	7880	
	15	6310		3	5460		2	7550	
	16	4540		4	4980		3	7770	
	17	4360		5	4980		4	6120	
	18	4360		6	5170				
	19	4540		7	5170	9. 1960: Aug	31	7990	
	20	4900					Sept 1	7990	
	21	4620	7. 1956: May	6	4980		2	8100	
	22	4540		7	5910		3	6010	
				8	7100		4	5790	
5. 1954: Aug	15	5460		9	7260		5	5790	
	16	5550		10	6630		6	5790	
	17	5640		11	5930		7	6560	
	18	5640		12	5840		8	7770	
	19	5640		13	5170		9	7220	
	20	5640		14	6420		10	6120	
	21	5640		15	6790		11	5900	
	22	5640		16	5930		12	5790	
	23	5640		17	5260		13	5900	
	24	5550		18	5080		14	6670	
	25	5550		19	5840		15	7220	
	26	5550		20	5360				
	27	5640		21	6260				
	28	5840		22	7590				
	29	5740							
	30	5740							

Appendix C.2. (cont)

Discharge at Augusta (cfs)

<u>Survey</u>	<u>Date</u>	<u>Discharge</u>	<u>Survey</u>	<u>Date</u>	<u>Discharge</u>	<u>Survey</u>	<u>Date</u>	<u>Dis-</u> <u>charge</u>
10. 1965:	May 30	6550	13. 1968	Aug 24	8230	16. 1976:	Aug 10	6900
	31	6780		25	6900		11	7430
	June 1	6960		26	6870		12	7650
	2	6900		27	7040		13	7630
	3	6640		28	7100		14	7850
	4	6620		29	7300		15	7640
	5	6600		30	7770		16	6930
	6	6560		31	7520			
	7	6530		Sept 1	6890			
	8	7070		2	6810			
11. 1965:	Sept 21	6460	14. 1972:	May 22	9610			
	22	6530		23	14700			
	23	7240		24	17100			
	24	7120		25	16500			
	25	6900		26	13700			
	26	6650		27	9940			
	27	6580		28	7440			
	28	6490		29	7320			
	29	7150		30	7250			
	30	6890		31	7310			
				June 1	7530			
12. 1968:	May 27	6750		2	7500			
	28	6670						
	29	6650	15. 1972:	Sept 13	6850			
	30	6650		14	7020			
	31	6700		15	9170			
	June 1	6480		16	7520			
	2	6730		17	7110			
	3	6860		18	6970			
	4	6840		19	6660			
	5	6570		20	7060			
				21	7090			
				22	7190			

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